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EFFECT OF HEAVY METALS ON BACTERIA

Elena Vratonjic

The Effect of Heavy Metals on Bacteria

By

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Columbus State University
The College of science
The Graduate Program In Environmental Science

Effect of Heavy Metals on Bacteria

A Thesis in
Environmental Science

By
Elena Vratonjic

Submitted in Partial Fulfillment
of the requirements
for the Degree of
Master of Science

December 2003

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I have submitted this thesis in partial fulfillment of the requirements for the degree of Master of Science

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ABSTRACT

Heavy metals have become devastating environmental contaminants due to their widespread use in the manufacture of electronics, plastics, batteries and dyes. The discharge of heavy metals into the environment has caused concern about their effects on the ecosystem. Many metals are essential for growth in small amounts. However, at higher concentrations heavy metals are toxic because they bind to organic compounds. This accounts for their effects on some important parts of the cell structure, like their ability to denature protein molecules. The initial objective of this study was to find out the growth rate of *E.coli* and *M.roseus* in the presence of 1mM and 10 μ M concentrations of cadmium and lead. *E.coli* and *M.roseus* were used due to their ability to grow in a controlled environment and ease of usage. The growth rates of the bacteria and viable counts were obtained via light scattering. Bacteria were grown on rich media (LB broth) as well as mineral M9 media supplemented with and without casamino acids and growth rates were compared. Another objective in this study was to identify genes in *E. coli* mutants that were more resistant or sensitive to cadmium. *E.coli* was mutagenized with a transposon to identify mutants with altered sensitivities to Cd. The growth rates of the mutants and wild type were obtained with and without Cd. Results suggest that Cd^{+2} , at 10 μ M and 1mM concentrations as well as Pb^{+2} at 1mM concentration, had a significant effect on *E.coli* and *M.roseus* in all media types. Insignificant effects were found in *M.roseus* and *E.coli* growth rates when exposed to 10 μ M Pb. The responses of bacteria to cadmium salts were 100 times more toxic in comparison to lead salts, suggesting slightly different biological effect of these heavy metals. The isolation of mutants resistant to specific toxic agents such as Tc and Rif antibiotics was not a very useful genetic approach in this study. It was unfortunate that it cannot be proven that real mutants were created. Instead, the donor was already cadmium sensitive, and the Rif^r donor appeared to be even more sensitive to cadmium and therefore behaved like a mutant. UV-light exposure, % survival, growth rates, and appearance of plasmids confirmed exactly the same behavior of Rif resistant donor and suspected mutant. Therefore it seemed like our donor strain was already a "mutant" that has very high cadmium sensitivity. A different approach was taken in order to identify *E. coli* genes involved with cadmium resistance, by simply increasing concentrations of Cd^{+2} . *E.coli* strain was found that is resistant to 4.5 mM Cd^{+2} .

TABLE OF CONTENTS

iv

ABSTRACT	iii
TABLE OF CONTENTS	iv
LIST OF FIGURES	vi
LIST OF TABLES	viii
INTRODUCTION	1
The effect of heavy metals on bacteria	1
Bacterial growth	7
Objectives	13
MATERIALS AND METHODS	14
Bacterial strains and growth conditions	14
Measurements of growth rates	15
Viable count procedure	15
Experimental protocol for preparing different media	16
Testing for sensitivity to Cd and Pb	16
Anova and Duncan's multiple test	17
Mutagenesis	18
Determination of minimal inhibitory concentration	19
<i>E. coli</i> plasmid mini-prep	19
Preparation of DNA electrophoresis gel	20
Selection of a Cd ⁺² resistant <i>E. coli</i> strain	20
UV exposure	21

RESULTS	23 ^v
<i>E. coli</i> and <i>M. roseus</i> growth rates with glucose and lactose at 1x 10 ⁻³ M cadmium and lead	23
<i>M. roseus</i> growth rates in different media at 1x 10 ⁻³ M and 1x 10 ⁻⁵ M cadmium and lead	24
<i>E. coli</i> growth rates in different media at 1x 10 ⁻³ M and 1x 10 ⁻⁵ M cadmium and lead	24
Mutagenesis	25
DISCUSSION AND CONCLUSION	99
REFERENCES	106
APPENDIX A - <i>E. coli</i> doubling times	109
APPENDIX B - <i>M. roseus</i> doubling times	199
APPENDIX C - Suspected mutant, donor and recipient doubling times	278

LIST OF FIGURES

Figure		Page
1-1	Average doubling times of <i>E. coli</i> vs. Cd and Pb from absorbance vs. time	33
1-2	Average doubling times of <i>E. coli</i> vs. glucose or lactose from absorbance vs. time	34
1-3	Summary of average doubling times of <i>E. coli</i> from absorbance vs. time	35
1-4	Average doubling times of <i>M. roseus</i> vs. Cd and Pb from absorbance vs. time	41
1-5	Average doubling times of <i>M. roseus</i> vs. glucose or lactose from absorbance vs. time	42
1-6	Summary of average doubling Times of <i>M. roseus</i> from absorbance vs. time	43
1-7	Average doubling times of <i>E. coli</i> vs. Cd and Pb from cell count vs. time	49
1-8	Average doubling times of <i>E. coli</i> vs. glucose or lactose from cell count vs. time	50
1-9	Summary of average doubling times of <i>E. coli</i> from cell count vs. time	51
2-1	Average absorbance of <i>E. coli</i> vs. Cd and Pb from cell count vs. absorbance	57
2-2	Average absorbance of <i>E. coli</i> vs. glucose or lactose from cell count vs. absorbance	58
2-3	Summary of average absorbance's of <i>E. coli</i> from cell count vs. absorbance	59
2-4	Average doubling times of <i>M. roseus</i> vs. Cd and Pb from cell count vs. time	65
2-5	Average doubling times of <i>M. roseus</i> vs. glucose or lactose from cell count vs. time	66

Figure		Page
2-6	Summary of average doubling times of <i>M. roseus</i> from cell count vs. time	67
2-7	Average absorbance of <i>M. roseus</i> vs. Cd and Pb from cell count vs. absorbance	73
2-8	Average absorbance of <i>M. roseus</i> vs. glucose or lactose from cell count vs. absorbance	74
2-9	Summary of average absorbance's of <i>M. roseus</i> from cell count vs. absorbance	75
3-1	Average doubling times of <i>M. roseus</i> vs. Cd and Pb from absorbance vs. time	81
3-2	Average doubling times of <i>M. roseus</i> vs. different media from absorbance vs. time	82
3-3	Summary of average doubling times of <i>M. roseus</i> from absorbance vs. time	83
3-4	Average doubling times of <i>E. coli</i> vs. Cd and Pb from absorbance vs. time	89
3-5	Average Doubling Times of <i>E. coli</i> vs. different media from absorbance vs. time	90
3-6	Summary of average doubling times of <i>E. coli</i> from absorbance vs. time	91
3-7	Average % survival for wild type, Tc ^r donor, Rf ^r donor and mutant	96

LIST OF TABLES

Table		Page
1-1	Doubling times for <i>E. coli</i> absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	28
1-2	ANOVA: Two-Factor With Replication for <i>E. coli</i> absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	29
1-3	Duncan's Multiple Range Test for <i>E. coli</i> absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	30
1-4	Mean Comparisons and Rp Comparisons for <i>E. coli</i> absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	31
1-5	Summary of doubling times for <i>E. coli</i> absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	32
1-6	Doubling times for <i>M. roseus</i> absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	36
1-7	ANOVA: Two-Factor With Replication for <i>M. roseus</i> absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	37
1-8	Duncan's Multiple Range Test for <i>M. roseus</i> absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	38
1-9	Mean Comparisons and Rp Comparisons for <i>M. roseus</i> absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	39
2-1	Summary of doubling times for <i>M. roseus</i> absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	40

Table		Page
2-2	Doubling times for <i>E. coli</i> cell count vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	44
2-3	ANOVA: Two-Factor With Replication for <i>E. coli</i> cell count vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	45
2-4	Duncan's Multiple Range Test for <i>E. coli</i> cell count vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	46
2-5	Mean Comparisons and Rp Comparisons for <i>E. coli</i> cell count vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	47
2-6	Summary of doubling times for <i>E. coli</i> cell count vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	48
2-7	Doubling times for <i>E. coli</i> cell count vs. absorbance in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	52
2-8	ANOVA: Two-Factor With Replication for <i>E. coli</i> cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	53
2-9	Duncan's Multiple Range Test for <i>E. coli</i> cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	54
3-1	Mean Comparisons and Rp Comparisons for <i>E. coli</i> cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	55
3-2	Summary of doubling times for <i>E. coli</i> cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	56
3-3	Doubling times for <i>M. roseus</i> cell count vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	60

Table		Page
3-4	ANOVA: Two-Factor With Replication for <i>M. roseus</i> cell count vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	61
3-5	Duncan's Multiple Range Test for <i>M. roseus</i> cell count vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	62
3-6	Mean Comparisons and Rp Comparisons for <i>M. roseus</i> cell count vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	63
3-7	Summary of doubling times for <i>M. roseus</i> cell count vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	64
3-8	Doubling times for <i>M. roseus</i> cell count vs. absorbance in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	68
3-9	ANOVA: Two-Factor With Replication for <i>M. roseus</i> cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	69
4-1	Duncan's Multiple Range Test for <i>M. roseus</i> cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	70
4-2	Mean Comparisons and Rp Comparisons for <i>E. coli</i> cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	71
4-3	Summary of doubling times for <i>M. roseus</i> cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	72
4-4	Doubling times for <i>M. roseus</i> absorbance vs. time in M9 and LB media exposed to 1×10^{-3} M and 1×10^{-5} M Cd and Pb	76

Table		Page
4-5	ANOVA: Two-Factor With Replication for <i>M. roseus</i> absorbance vs. time grown in M9 and LB media exposed to 1×10^{-3} M and 1×10^{-5} M Cd and Pb	77
4-6	Duncan's Multiple Range Test for <i>M. roseus</i> absorbance vs. time grown in M9 and LB media exposed to 1×10^{-3} M 1×10^{-5} M Cd and Pb	78
4-7	Mean Comparisons and Rp Comparisons for <i>M. roseus</i> absorbance vs. time grown in M9 and LB media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb	79
4-8	Summary of doubling times for <i>M. roseus</i> absorbance vs. time grown in M9 and LB media exposed to 1×10^{-3} M 1×10^{-5} M Cd and Pb	80
4-9	Doubling times for <i>E. coli</i> absorbance vs. time in M9 with and without casamino acids and LB media exposed to 1×10^{-3} M and 1×10^{-5} M Cd and Pb	84
5-1	ANOVA: Two-Factor With Replication for <i>E. coli</i> absorbance vs. time grown in M9 with and without casamino acids and LB media exposed to 1×10^{-3} M and 1×10^{-5} M Cd and Pb	85
5-2	Duncan's Multiple Range Test for <i>E. coli</i> absorbance vs. time grown in M9 with and without casamino acids and LB media at 1×10^{-3} M and 1×10^{-5} M Cd and Pb	86
5-3	Mean Comparisons and Rp Comparisons for <i>E. coli</i> absorbance vs. time grown in M9 with and without casamino acids and LB media at 1×10^{-3} M and 1×10^{-5} M Cd and Pb	87
5-4	Summary of doubling times for <i>E. coli</i> absorbance vs. time grown in M9 with and without casamino acids and LB media exposed to 1×10^{-3} M and 1×10^{-5} M Cd and Pb	88
5-5	Exposure of Rf ^r wild type, Tc ^r , Rf ^r donor, and mutant strain to UV-light	92

Table		Page
5-6	% Survival of wild type, Tc ^r donor, Rf ^r donor, and mutant strain	93
5-7	Single Factor ANOVA for % survival of wild type, Tc ^r donor, Rf ^r donor and mutant strain	94
5-8	Analysis of Variance for % survival of wild type, Tc ^r donor, Rf ^r donor and mutant strain	95
5-9	Duncan's Multiple Range Test for % survival of wild type, Tc ^r donor, Rf ^r donor and mutant strain	97
6-1	Mean Comparisons and Rp Comparisons for % survival of wild type, Tc ^r donor, Rf ^r donor and mutant strain	98

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INTRODUCTION

The effect of heavy metals on bacteria

Heavy metals have become significant environmental contaminants due to their widespread use in the manufacture of electronics, plastics, batteries and dyes. The discharge of heavy metals into the environment has caused concern about their effects on the ecosystem (Shapiro and Keasling, 1996). Though many metals are essential for growth in small amounts, at higher concentrations heavy metals are toxic because they bind to organic compounds. This accounts for their effects on some important parts of the cell structure, like their ability to denature protein molecules (Shapiro and Keasling, 1996, Morozzi et al., 1982). Because of their intrinsically persistent nature and toxicity of the metals to living systems, the mechanisms of resistance and repair of damage caused by heavy metals is an important and unresolved problem (Pazirandeh, 1996). Some of the earliest attempts to control microorganisms by chemical means were based on the use of heavy metals. For example, copper sulphate was used as a plant fungicide, and mercury salts were used in the treatment of certain infectious diseases (Sadler and Trudinger, 1967).

Data from numerous studies indicates that the presence of high concentrations of metals provides a selective medium favoring metal resistant bacteria (Mietz and Sjogren, 1982). Some bacteria are resistant to concentrations of heavy metals that are toxic to higher organisms. These

bacteria may concentrate or change the valence of heavy metals and remove them from the environment. Each species may be affected in different ways, so that a particular concentration of a metal may exert no, little, or a dramatic effect, depending on the microorganism's characteristics (Gadd, 1990).

Most bacteria exhibit a biphasic response to a number of heavy metals. At low concentrations of the metal there is a stimulation of growth, but as the metal concentration is increased growth becomes progressively inhibited and finally ceases. The "cross-over" zone between stimulation and inhibition by metals is generally over a very narrow range. The actual concentrations at which the responses occur depend upon the organism, the form of the metal, and the chemical and physical composition of the medium in which the responses are determined (Gadd, 1990). An understanding of the mechanisms by which microbes survive heavy metal exposure is critical for understanding heavy metal toxicity in all organisms and for the use of such organisms for the removal of metals from the environment (Shapiro and Keasling, 1996). Furthermore, the biological effects of such pollutants on microorganisms and the frequency of appearance of heavy metal resistant bacteria can be useful as an indicator in bioassay systems (Gelmi et al., 1994).

An increasing frequency of appearance of bacteria resistant to toxic metals such as cadmium seems to be correlated with increasing loads of metals in the environment (Gelmi et al., 1994). However, the interaction of inorganic lead (Pb) and cadmium (Cd) with well-studied bacteria such as *Escherichia coli* has received little attention. One reason for this may be because *E. coli* is not

a normal member of soil flora, so it is usually not isolated from metal contaminated sites. Another reason for the lack of study of effects of heavy metals on *E. coli* may be that phosphate found in most standard media combine with many metal ions to form an insoluble precipitate. Metal solubility is critical for uptake into the bacterial cell. Avoidance of precipitation by the phosphate found in most standard media is necessary prior to analysis of growth inhibition. Metals reduce growth if a modified medium containing glycerol-2-phosphate is used to lower the concentration of precipitate (LaRossa et al., 1994). Chelating compounds, such as citrate and EDTA have been shown to slightly increase the toxicity of heavy metals (Gadd and Griffiths, 1978).

The extent to which a metallic cation interacts in soil environments with organic and inorganic surfaces determines the concentration of metal in solution. Bacteria have a high surface area to volume ratio and a high capacity for sorbing metals from solution. Many studies have shown that large quantities of metallic cations are complexed by bacteria (Mullen et al., 1989). Previous studies concluded that cell walls of gram-positive bacteria such as *B. subtilis* bind larger quantities of several metals than cell envelopes of the *E. coli* (gram-negative bacterium) (Beverage and Fyfe, 1985).

The objective of a microbe study by Mullen et al. (1989) was to determine the metal binding capacities of whole cells of two-gram positive bacteria and two-gram negative bacteria. Concentrations of $1\mu\text{M}$ - 1mM Ag^+ , Cd^{2+} , Cu^{2+} , and La^{3+} were used as metal salt solutions. Examinations of the values of Cd sorption showed gram-negative *E. coli* and *P. aeruginosa* were more efficient at

Cd sorption than gram-positive bacteria. On the other hand, *B. subtilis* removed the most Cu^{2+} at an equilibrium concentration of $1\mu\text{M}$. High equilibrium concentrations affected gram-negative bacteria the most and gram-positive bacteria the least. This research evaluated the sorption capabilities of selected bacteria (Mullen et al., 1989).

Traditionally, adsorption, chemisorption, and ion exchange have been used to describe the removal of various cations. When using intact bacteria cells we should also consider other processes like active uptake of the metal into the cytoplasm through non-specific cation transport systems. Evidence suggests that the role of bacteria is not limited to short term metal immobilization and that bacteria cells are capable of binding large quantities of metal cations (Mullen et al., 1989). There is evidence suggesting that microbial biomasses have high heavy metal uptake capacities. One possible method of concentration may be due to the production of certain metal-binding proteins such as metallothioneins. These proteins are cysteine rich and bind heavy metals, such as Cd, with very high affinity (Pazirandeh, 1996). Pazirandeh (1996) suggests further that *E. coli* does not have any gene encoding metallothionein-like proteins. Even without a metallothionein gene, *E. coli* demonstrates considerable heavy metal resistance (Gelmi et al., 1994). The study of this resistance could identify the genes that will give *E. coli* improved ability to sequester heavy metals (Pazirandeh, 1996).

Cenci et al. (1985) used total dehydrogenase activity (TDH) as a means of measuring metal toxicity in *E. coli*. TDH was used since it is a measure of

bacterial activity closely related to energetic metabolic systems and beta-galactosidase as a typical inducible enzyme in *E. coli*. From the results it was possible to define the order of metal toxicity. The 50 percent inhibition of TDH-activity was: 7.4, 10, 30, and 50 μM , respectively for Hg, Zn, Cd, and Cu. In the presence of the metals incubation time of over 60 minutes consistently lead to the inhibition of TDH activity. Experiments were carried out to define some characteristics of the biological damage to the total cell population. Hg and Cu markedly inhibited growth in Trypticase soy agar (TSYA) medium, while the viability of cells treated with Cd and Zn remained constant in the same medium. This indicated that treatment with Cd and Zn, unlike that observed for Hg and Cu, did not cause metabolic injury to *E. coli* (Cenci et al., 1985).

Continued pollution of the biosphere with toxic heavy metals makes it clear that microbe-based technologies will have an important role in environmental remediation (Gadd, 1990). Technological applications of microbial metal concentration may depend on the ease of metal recovery either by biomass regeneration or by reclamation. If inexpensive biomass is used to reclaim valuable metals, then destructive recovery (incineration or dissolution in acids or alkalis) may be economically feasible. Microorganisms with high capacity for metal uptake removal such as *Staphylococcus aureus*, *Bacillus sphaericus*, *Bacillus licheniformis* and *Arthrobacter species* can be used as both biological monitors of environmental contamination and for remediation (El-Bestawy et al., 1998). These species were isolated from water heavily polluted with heavy metals. Mutagenesis, both physical and chemical,

and gradual increase in concentration of heavy metals significantly increased bacterial resistance against a wide range of metals and led to high efficiencies for their removal from soil. Physical mutation was induced by u.v light (254 nm) at 30 cm height, and at the dose that resulted in 90% mortality. Agar well diffusion technique was used for chemical mutation that was induced by 1% ethidium bromide. In this study, the highest removal efficiency achieved by the different selected mutants was Cd (89.9-100%), Cr (87.3-99.7%) and Pb (40.2-51%) (El-Bestawy et al., 1998).

Microbes are generally the first organisms exposed to heavy metals present in the environment. Although the presence of heavy metals is detrimental to them, toxic metals select resistant variants that confer the ability to tolerate higher levels of the toxic compounds (Cervantes et al., 1994). Heavy metal resistance has been shown to be plasmid mediated in some cases, and the genetic determinants responsible for heavy metal resistance often reside on plasmids that also mediate antibiotic resistance (Mietz and Sjogren, 1982).

Transposons are useful for making mutations since they can be helpful in gene identification. Transposons are specific DNA segments with the ability to move as a unit in a more or less random fashion from one genetic point to another. Complete loss of gene function often results upon insertion of a transposon within a gene. Therefore, transposons can be used as a mutagenic agent, if a replicon that carries the transposon can be efficiently introduced into the organism to be mutagenized. Selecting for transposon-mediated antibiotic resistance can easily isolate transposition events. Molecular cloning

of prokaryotic genes and manipulation and genetic analysis of bacteria have been greatly facilitated by the use of antibiotic-resistance transposons (Simon et al., 1983).

Genetic manipulation of bacteria may allow the development of new strains with better growth rates and improved resistance to toxic metals. Therefore, manipulation of wild type, mutant, and individual or mixed cultures may be a useful tool for decontamination of polluted effluents by metal bioremediation (Brierley et al., 1982).

Bacterial growth

The simplest measure of the effect of heavy metals on bacteria is obtained by examining their effect on growth rate. Bacterial cultures are exposed to different concentrations of various heavy metals and the growth rate is determined (Mandelstam and Mc Quillen, 1968).

When bacterial cells are placed in a nutritionally favorable environment they will grow and divide. At some point due to exhaustion of nutrients, depletion of oxygen, and/or accumulation of toxic products, the environment will become unfavorable for growth. Until this occurs, the bacteria grow in an unhindered manner, and each cell will grow, on the average, at the same rate as its predecessors. This is a geometric doubling called logarithmic growth (Ford and Mitchell, 1992).

Growth parameters most often used to determine growth rate are those that can be easily and accurately measured. One parameter easily measured is

turbidity of a liquid culture, which depends on the amount of light that is scattered by the organisms and is related to the bacterial mass (Collins and Stotzky, 1992). Measuring living or viable cells is also a useful method for determining growth rates. The number of viable bacteria is determined by colony count, which is performed by placing an appropriate dilution of the culture on or in a solid medium where the organisms are known to grow. Since colonies arise from single cells, the number of colonies multiplied by the dilution factor equals the number of viable bacteria originally present. Standard curves are used to relate the viable cells to the turbidity (Mandelstam and Mc Quillen, 1968).

E. coli grows well in chemically defined media. Some of the media are very simple, containing only mineral salts and a single organic component as a source of carbon and energy. When glucose is the organic component the bacterial mass may double every 45-60 minutes. Growth is slower when other substances such as acetate or succinate are used as the carbon energy and source. The growth rate is increased considerably by the addition of amino acids, purines and pyrimidines, and vitamins. These are best provided by use of a complex medium containing these substances (Ford and Mitchell, 1992). Since *E. coli* can take up sugars, proteins and amino acids from the complex medium and need not synthesize them *de novo*, growth is much more rapid than on mineral medium. Many different types of culture media have been employed in the study of bacteria, and some common laboratory species will grow in complex medium. Complex media is generally a solution composed of

peptones, concentrated meat or yeast extract, and salt. Extracts are composed of the water-soluble constituents of meat or yeast, which will diffuse out of the meat or yeast on standing in water, and it supplements the peptone and helps the growth of bacterial species (Ford and Mitchell, 1992).

The growth rate of bacteria can also be manipulated by altering the temperature. With *E. coli* at 37°C the growth rate ranges from 18-20 minutes in a complex medium to about 2 hours in media where the sole carbon and energy source is one of the several sugars which are metabolized inefficiently (Sadler and Trudinger, 1967).

If *E. coli* is inoculated into an appropriate growth medium in the evening, by the following morning there will be signs of considerable growth. In order to obtain growing bacteria in log phase, a portion of this overnight culture must be sub-cultured into fresh medium. Growth will not begin immediately, but will start slowly, reaching the maximum growth rate in a gradual manner. This period prior to the maximum growth rate is called the lag phase. The duration of this period depends on many considerations, including the organism, the medium, the conditions of previous cultivation, the degree of aeration, and the way in which bacterial growth is measured. At the end of the lag phase, the cells are in a physiological steady state, undergoing balanced growth as previously described. This state ceases more or less gradually when the cells begin to exhaust essential nutrients or when they accumulate toxic products (See Figure 1). The time at which the log phase ends is entirely dependent on the species of organism and the actual conditions of cultivation. When either

the carbon or the nitrogen source is limiting in a synthetic medium, growth of *E. coli* ends very abruptly (Mandelstam and Mc Quillen, 1968).

Heavy metals such as Cd^{+2} , Cr^{+6} , Hg^{+2} , Pb^{+2} , and Zn^{+2} , produce a decrease of *E. coli* growth rate, and reduced total biomass. All the concentrations of toxicants tested in the work done by Mariscal et al. (from 0.3 μM to 5 μM) produced at least a partial inhibition of *E. coli* growth rate. This inhibition was reflected in the decrease of the respective values of growth rate, in comparison with those of controls. Comparison of the values revealed that, Cr^{+6} , Hg^{+2} , Cd^{+2} , Pb^{+2} , and Zn^{+2} in that order, were the most to least toxic to *E. coli* (Mariscal et al., 1995).

Studies have resulted from the reduction of population growth of several microorganisms in bodies of water from industrial effluents containing heavy metals. Heavy metals probably bound with the cellular membrane of the bacteria by impairing certain physiological and biochemical parameters. The results suggested that the degree of growth population of *E. coli* varied among metal ions in the aquatic environment. Effects of different concentrations of metals on the population growth of *E. coli* after 7 and 28 days were analyzed in the treated and untreated experimental sets. A gradual decline in population growth of *E. coli* was noted with increasing incubation time and metal concentration. The harmful effects of the metals were found to be highest in Cd, then Pb, Cu, As, Hg, and lowest in Cr (Jana and Bhattacharya, 1987).

In a study by Pickett and Dean (1979), the effect of different concentrations of Cd^{+2} and Zn^{+2} on *Bacillus subtilis* subsp. *niger* and

Pseudomonas fluorescens were investigated. Organisms in this study that were cultured many generations in liquid medium at concentrations between 0.12 and 0.20 mM of Cd^{+2} or Zn^{+2} all survived on agar plates containing metal ions at that concentration, but at higher concentrations the viability decreased sharply. The minimum Cd^{+2} concentrations completely inhibiting the growth of *Bacillus subtilis* subsp. *niger* and *Pseudomonas fluorescens* were 0.62 and 0.13 mM respectively, suggesting that *Pseudomonas sp.* is a more sensitive strain. The same trend was seen for Zn^{+2} , which inhibited these strains at 1.53 and 0.12 mM respectively. As with many other toxic agents, the lag increased as the inhibitor concentration increased. There was also a distinct anomaly in the form of a plateau, after the lag phase, which occurred at Cd^{+2} concentrations between 0.12 and 0.2 mM. Evidence suggests that well-defined sites such as the cell walls on the organisms were initially attacked by metal ions. The degree of inhibition of growth rates was proportional to the concentration of metal ions present in the medium. When the cell walls were saturated with metal ions, small increases in concentration had no effect, and a relatively high concentration was required before other less accessible sites were affected. In liquid medium the lag and the mean generation time of *Bacillus subtilis* subsp. *niger* increased with increasing Cd^{+2} or Zn^{+2} concentrations, whereas only the total biomass was affected in *Pseudomonas fluorescens*. Nevertheless, the responses of both species indicted a specific action at low concentration and a more general toxic action at high concentrations (Pickett and Dean, 1979).

In a study by Morozzi et al. (1982) the effect of Hg^{2+} , Cd^{2+} , Cu^{2+} , Zn^{2+} , Cr^{2+} , and Pb^{2+} on *E. coli* were studied. The growth curves of *E. coli* in cultures containing sub-lethal concentrations of metal ions (ranging from 0.01 μM to 100 μM) show that there is no effect on the lag phase time until the threshold concentration is reached. The metals studied, and the corresponding values of threshold concentration that were derived from the graph, were as follow: 0.005 μM for Hg^{2+} , 0.5 μM for Cd^{2+} , 5 μM for Cu^{2+} , 23 μM for Cu^{2+} , 30 μM for Zn^{2+} and 45 μM for Pb^{2+} . In order to evaluate the persistence of the tolerance to metal ions, *E. coli* previously exposed to metals were subcultured into fresh media and then re-inoculated into media plus metal at different times (0-12 hours). Previous exposure to the single metals diminished the toxicity of Hg^{2+} , Cd^{2+} , Zn^{2+} , Cu^{2+} , and Cr^{2+} . The mid-log phase times of primary sub-cultures, after four, five, and six hours respectively, came near the values of the controls without metals. The acquired tolerance is lost in a very short time if bacteria grow in the absence of metal. In this experiment, the lag phase began to lengthen again when bacteria without the metal were sub-cultured into medium containing the corresponding ion. Loss of viability was only observed for microorganisms tested after the exposure to lead. The reduction of metallic ions indicated a physiological adaptation of microbes rather than the selection of mutants. In a study by Morozzi et al. (1982) cells that have become adapted to the presence of metal ions lost this adaptation after growing in a metal-free medium. In conclusion, metal ions Hg^{2+} , Cd^{2+} , Cu^{2+} ,

Cr^{2+} , Zn^{2+} did not cause selection of metal-resistant mutants of *E. coli* (Morozzi et al., 1982).

OBJECTIVES:

The initial objective of this study is to find out the growth rate of *E. coli* and *M. roseus* in the presence of 1mM and 10 μM concentrations of cadmium and lead. *E. coli* and *M. roseus* will be used due to their ability to grow in a controlled environment and ease of usage. Growth rates of *Escherichia coli* (gram-negative bacteria) and *Micrococcus roseus* (gram positive bacteria) will be observed before and after exposure to lead or cadmium. The growth rates of the bacteria will be measured via light scattering, and viable counts will be obtained. Bacteria will be grown on rich media (LB broth) as well as mineral M9 media supplemented with and without casamino acids and growth rates will be compared.

Another objective in this study is to identify genes in *E. coli* mutants that will be more resistant or sensitive to cadmium. *E. coli* will be mutagenized with a transposon to identify mutants with altered sensitivities to Cd. The growth rates of the mutants and wild type will be obtained with and without Cd. The null hypothesis in this study states, "Mutagenesis will have no effect in mean growth rates between wild type and mutant".

MATERIAL AND METHODS

Chemicals and instruments utilized were available in the Departments of Biology and Chemistry and Geology Department, College of Science, Columbus State University. Rifamycin (Rf) and tetracycline (Tc) were purchased from Sigma.

Bacterial strains and growth conditions

E. coli was supplied from Bactrol discs (ATCC 25922), and *M. roseus* from Microbank (29). For transposon mutagenesis *E. coli* S17 mini Tn5-Tc (Tc^r, Rif^s) (de Lorenzo et al., 1990) was the plasmid donor. *E. coli* (ATCC 25922) was grown on Rif plates and surviving colonies were used as the recipient (Tc^s, Rif^r). Selection for antibiotic resistance was performed on LA plates supplemented with Tc or Rf to final concentrations of 15µg/ml and 50µg/ml, respectively from sterile stock solutions. Antibiotic stock solutions were prepared by adding 50 mg of Rif per ml of Dimethyl Formamide (DMF), and 15 mg of Tc per ml of 70% ethanol followed by filter sterilization.

Escherichia coli and *Micrococcus roseus* were initially grown in minimal medium containing 4.85µg Na₂HPO₄, 3.0g NaCl, 1.0g NH₄Cl, 0.27g MgSO₄, and 100µL of 1.0M CaCl₂ per Liter of distilled water at pH 7.0. When necessary, 5.0g of casamino acids and 5g of glucose or lactose were added. Agar powder was added to 15 g/L to prepare solid medium. Bacteria were also grown in LB

broth (Difco Laboratories, Detroit, MI) prepared according to manufacturer's instructions, always supplemented with 2g/L of glucose. When necessary, Tc and Rif were added to LB. All media were sterilized before addition of antibiotics, by autoclaving at 121°C for 15 minutes.

Filter sterilized stock solutions of 0.1M of CdCl₂, PbNO₃, and EDTA were prepared in de-ionized water and metal solutions were simultaneously introduced to EDTA. Solutions were then introduced at final concentrations of 10⁻³ and 10⁻⁵ M.

Measurement of growth rates

Growth rates were monitored using a Milton Roy Spectronic 20D spectrophotometer (St. Petesburg, FL) set at a wavelength of 650 nm. Measurements were plotted against time to determine growth rate for each species.

Viable count procedure

Viable cell counts were performed by pour plate technique of tenfold serial dilutions of control and 1 mM Cd and Pb containing samples. 1 ml of culture dilutions was pipetted into a sterile petri plate, adding 20 ml of 45°C Tryptic Soy Agar and gently swirling the plate on the tabletop. Plates that have between 30 and 300 colonies were counted. Three replicate plates of each dilution were prepared.

Experimental protocol for preparing different media

Bacteria were inoculated into M9 broth media (with 5% glucose or lactose) with and without casamino acids and LB broth media (supplemented with 2 % glucose) and grown overnight. These bacteria were then used to inoculate standing tubes for the experiment. The experimental tubes included M9 media containing lactose only, glucose only, lactose with severely limited glucose, M9 media without casamino acids and LB broth. Cultures were inoculated on ice to prevent growth of the bacteria, and then transferred to a 37°C water bath for standing incubation. Turbidity was measured every 30 minutes with spectrophotometer at 650 nm.

Testing for sensitivity to Cd and Pb

After control readings of the four media types above were completed, ten-milliliter test tubes of differently supplemented broths were inoculated with bacteria using a sterile, plastic inoculating loop. Identical procedures were used as the control, but medium contained the following:

1. Glucose with casamino acid and cadmium
2. Glucose with casamino acid and lead
3. Lactose with casamino acid and cadmium
4. Lactose with casamino acid and lead
5. Lactose with casamino acid, limited glucose, and cadmium

6. Lactose with casamino acid, limited glucose, and lead
7. Glucose with cadmium
8. Glucose with lead
9. LB broth with glucose and cadmium
10. LB broth with glucose and lead

Controls were repeated at this stage. All tubes were incubated in a water bath at 37°C, and the absorbance was read every 30 minutes until bacterial growth was well into stationary phase. Three independent repetitions of each experiment were performed in this study.

ANOVA and Duncan's multiple test

Analysis of Variance (ANOVA) was used for analyzing data in this study. ANOVA is used to uncover the overall effects of independent variables on an interval dependent variable. The key statistic in ANOVA is the F-test of difference of group means, testing if the means of the groups formed by values of the independent variable are different enough not to have occurred by chance. If the group means do not differ significantly then it is inferred that the independent variables did not have an effect on the dependent variable. If the F test shows that overall the independent variables are related to dependent variables, then multiple comparisons tests of significance are used (Krebs, 1999). In this study, once it was determined that a significant difference exists within the level means of a variable, it was necessary to detect the location of significant differences. Duncan's Multiple Test was used

to distinguish exactly where the differences were significant for *E.coli* and *M.roseus*. Duncan's Multiple Test involves three steps consisting of calculating standard error (S_x), arraying or ranking the means for the treatment or interaction in question, and calculating confidence intervals and comparing them (Steel and Torrie, 1960).

Mutagenesis

Overnight cultures of donor and recipient strains (0.1 ml) were transferred into 10 ml of LB liquid cultures with appropriate antibiotics, and grown to log phase. 0.1 ml of each strain was centrifuged in separate Eppendorf tubes for 1 minute. The cell pellets were washed twice in 0.1 ml of LB and re-suspended in 0.1 ml of LB. The two strains were mixed together, centrifuged for 1 minute, the liquid discarded, and the cell pellets re-suspended in 20 μ l of LB broth. The mating mixture was spotted on LA plates and incubated overnight at 30°C. The overnight growth was scraped off the plate and suspended in 1 ml of LB, and 0.1 ml samples were then spread on LA plates with Tc and Rif. Controls of donor and recipient only were treated identically. Approximately 1000 colonies that acquired the transposon antibiotic resistance were obtained on these plates. Tc^r, Rif^r colonies were picked and replica-plated on LA/Tc/Rf, and LA/Tc/Rf/CdCl₂ plates. Putative mutants were cultured in liquid LB + 1 mM CdCl₂ to confirm Cd sensitivity. Mutants exhibiting abnormal growth were selected for further analysis.

Determination of minimal inhibitory concentration

Suspected mutants that didn't grow on plates in the presence of 1 mM of Cd were tested to find their minimal inhibitory concentration. Mutants were grown overnight at 37°C in LB medium + Tc + Rf +10 µM Cd⁺². Growth was observed after 24 hours, suggesting that the minimal inhibitory concentration was higher than 10 µM. Concentrations were incrementally increased until no cell growth was observed.

E. coli plasmid mini-prep

1-2 ml of cell culture was centrifuged for 1 minute in a microfuge, the pellet was collected and the culture fluid was discarded. The cell pellet was resuspended in 0.1 ml of TE buffer. Lysis solution (0.2 ml) was added and mixed thoroughly by inversion. Potassium acetate (3M) 0.15 ml was added and mixed thoroughly by inversion until a white precipitate formed. The solution was centrifuged for 5 minutes in a microfuge. The cleared lysate was transferred to a new tube, avoiding the white precipitate that was on the side and bottom of the tube. Cold 95-100% (0.8 ml) was added to the lysate and mixed. The solution was centrifuged for 5 minutes in microfuge. The liquid was discarded and the pellet was saved. The liquid was completely removed with a pipette. The pellet was washed with 0.5 ml of cold 70% ethanol, dried, and resuspended in 20-50 µl water or TE buffer. Instructions outlined in the Laboratory Manual by Sambrook et al. (1989) were used to prepare alkaline lysis buffers.

Preparation of DNA electrophoresis gel

Plasmid preparations were made for potential mutants, wild type, and donor cell (Tc^r). Pellets from the plasmid preps were dissolved in 30 μ L of TE buffer and divided into a control portion and a portion to be digested. These plasmid preps were then digested with restriction enzyme Pst I (Roche Diagnostic Corporation, Indianapolis, IN) according to manufacturer's instructions. Instructions for preparing 1% agarose gel and 10X loading buffer were extracted from the Laboratory Manual by Sambrook et al. (1989). Prior to electrophoresis, 2 μ L of 10X loading buffer was added to each tube. DNA Molecular Marker IV (Roche Diagnostic Corporation, Indianapolis, IN) was also run on the gel with all of the samples. After electrophoresis, the gel was stained in 10 μ g/ml ethidium bromide solution for 10 minutes. The gel was transferred to distilled water for five minutes, and then visualized by Foto//Phoreisis I apparatus by Fotodyne at wavelength of 312 nm. Photography of these gels was taken afterwards.

Selection of a Cd⁺² resistant *E. coli* strain

E. coli was grown on LA plates that were prepared with different concentrations of Cd⁺². The minimum starting concentration was 500 μ M. Flourishing colonies were selected and grown in LB with the same concentration of Cd⁺². Once the bacteria reached log phase, a loopful was spread on LA plates prepared with a higher concentration of Cd⁺². LA plates

were started with very low cadmium concentration of 0.5mM, followed by the intermediate concentrations of 1mM, 1.5mM, 2mM, 2.5mM, 3mM, 3.5mM, and 4mM. This process was replicated until suspected mutant reached its maximal growth with Cd^{+2} of 4.5 mM. At the end, wild type *E. coli* and mutant that was Cd^{+2} resistant were tested for Cd^{+2} sensitivity. They were grown in LB with and without maximum concentration of Cd^{+2} that mutant could grow on and results were recorded.

UV exposure

Wild type (Rif^r recipient), Tc^r donor, Rif^r donor, and suspected mutant were grown overnight in LB at 37°C. Once grown to same OD of 0.35, an equal volume of culture was placed on LA plates along five lines. Each line was exposed to UV light in five seconds increments ranging from 0 to 20 seconds. Uv mineralized lamp (UVS-54) was set five inches above the plates at a wavelength of 254 nm. The plates were incubated for 24 hours at 37°C, and then examined for growth.

The survival of bacterial cells exposed to 10 seconds of UV-light was then compared to controls that were not exposed to the UV-light. Wild type (Rif^r recipient), Rif^r donor, Tc^r donor, and suspected mutant were grown to mid-log phase, and samples of grown culture were diluted 10^1 to 10^7 times with sterile phosphate buffer (PBS). Spread plating on a series of LA plates was done by 100 μ l of diluted samples. After 24-hour incubation at 37°C, the number of

colonies formed before and after the UV-light exposure was counted and % survival was determined. Only plates that contained between 30 to 300 colonies were counted. Three replicate plates of each dilution were prepared.

RESULTS

E. coli and *M. roseus* growth rates with glucose and lactose at 1×10^{-3} M cadmium and lead.

Doubling times for *E. coli* and *M. roseus* were initially derived from absorbance vs. time. Two-Factor ANOVA with replication showed that there was a significant change in absorbance observed with increasing concentration of cadmium and lead (P-value < 0.001). Duncan's Multiple Test was used to distinguish where the differences are significant between *E. coli* and *M. roseus* when supplemented with glucose and lactose and exposed to Cd and Pb. Therefore there is no significant difference in doubling times between *M. roseus* and *E. coli* grown in M9 mineral medium supplemented with glucose and lactose. There were significant differences noted in *M. roseus* doubling times when exposed to Cd and Pb, but not between the heavy metals alone. All interactions between the heavy metals and control were significant in *E. coli* doubling times.

Doubling times were also found in *E. coli* and *M. roseus* from cell growth vs. time and cell growth vs. absorbance. Two-Factor ANOVA with replication showed that increasing concentration of cadmium and lead (P-value < 0.001) caused a significant reduction in growth rates. Duncan's Multiple Test replicates previously mentioned results for both *E. coli* and *M. roseus*. Therefore, only media with glucose was used since there was no significant

difference observed between media supplemented with glucose or lactose at 95% confidence level.

***M. roseus* growth rates in different media at 1×10^{-3} M and 1×10^{-5} M cadmium and lead.**

Graphs of absorbance vs. time for *M. roseus* illustrate growth rates in M9 mineral and rich media (LB) when exposed to 1×10^{-3} M and 1×10^{-5} M of Cd and Pb. Two-Factor ANOVA with replication showed that there was a significant decrease in growth rates observed with increasing concentration of cadmium and lead (P-value < 0.001). Duncan's Multiple Test indicated that the differences between 1×10^{-3} M and 1×10^{-5} M Cd and Pb were significant, while interactions between 1×10^{-3} M Pb and 1×10^{-5} M Cd, and 1×10^{-5} M Pb and control (bacteria that hasn't been exposed to either Pb or Cd) were not significant. Also, there is no significant difference between *M. roseus* in M9 mineral and rich media.

***E. coli* growth rates in different media at 1×10^{-3} M and 1×10^{-5} M cadmium and lead.**

Since *M. roseus* couldn't grow in M9 media without casamino acid, growth rates of *E. coli* were compared in M9 medium with and without casamino acid and LB, at 1×10^{-3} M and 1×10^{-5} M Cd and Pb. Two-Factor ANOVA with replication showed that there was a significant decrease in growth rates observed with increasing concentration of cadmium and lead (P-value < 0.001).

It can be observed from Duncan's Multiple Test that there is no significant difference between 1×10^{-5} M of Pb and control. All of the other interactions of growth rates in between the heavy metals, and heavy metals and controls were significant, as well as all of the growth rates of *E. coli* that were grown in M9 medium with and without casamino acid and LB at 95% confidence level.

FIGURES A-1 through A-125 in APPENDIX A, and FIGURES B-1 through B-99 in APPENDIX B, as well as TABLES 1-1 through 5-4 present all of the above raw data of *E. coli* and *M. roseus* and their complete analysis.

Mutagenesis

Approximately 1000 Tc^r , Rf^r colonies that acquired the transposon antibiotic resistance were obtained on LA plates. Colonies were replica-plated on LA/Tc/Rf, and LA/Tc/Rf/CdCl₂ plates. The effect of Tc^r , Rf^r colonies with and without Cd was different. Their appearance varied from small and dry to large and slimy in the presence of Cd. Eight different mutants were found that didn't grow at all on plates in the presence of Cd. Putative mutants, wild type *E. coli*, Rif^r recipient, Rif^r donor, and Tc^r donor were grown in LB, LB/Rif/Tc, and LB/Rif/Tc and 50 μ M, 100 μ M, and 300 μ M of Cd. Observed growth rates are presented in APPENDIX C, FIGURES C-1 to C-32, and in summary TABLE C-1. Rif^r donor, Tc^r donor, and suspected mutant demonstrated the same growth rates. Minimal Inhibitory concentration (MIC) was found to be 300 μ M in all mutants. It was possible that this Rif^r , Tc^r phenotype would arise from a Tc^r mutant or from Rif^r donor. If that were true suspected mutant would have Tc

donor plasmid that has never entered the chromosome. Therefore donor and recipient strains and their Cd sensitivity were tested and compared to the "mutants". Cd sensitivities in wild type, recipient, donor, and "mutant" strains in summary TABLE C-1 suggest the possibility of picking Rif resistant donor that was naturally more Cd sensitive than the recipient strain. Wild type, recipient, donor, and "mutant" strains were prepared and run on DNA electrophoresis gel. Figures C-33 to C-36 show the presence of transposons in the "mutant" strains.

To confirm our results, UV exposure was done on wild type *E. coli*, Rif^r recipient, Rif^r donor, Tc^r donor, and suspected mutant. UV-light exposure was necessary to see if suspected mutant behaved like the donor or wild type. After identical incubation the LA plates were inspected. Wild type *E. coli* and Rif^r recipient demonstrated the same UV resistance. Suspected mutant was sensitive to 5 seconds exposure to UV light. Tc^r donor was sensitive to longer exposure of 10 seconds to UV mineral light. Rif^r donor behaved in the same manner as suspected mutant illustrating the same 5 seconds sensitivity to UV light.

Since UV-light exposure couldn't tell us the exact number of cells that were affected, it was decided to validate the results by exposing bacteria to UV-light to determine % survival. Viable counts were compared to non-exposed samples of the same culture. The control invariably contained 100 ± 10 colonies. Percent survival of wild type (Rif^r recipient), Tc^r donor, Rif^r donor, and mutant was 78.7%, 13.5%, 6.2%, and 4.2 %, respectively. Single Factor

ANOVA showed that there was a significant change in % survival among tested bacteria (P -value < 0.001). The calculated F -value of 287 and F -critical value of 4 indicated that means of % survival are significantly different for two degrees of freedom. Analysis of Variance and Duncan's Multiple Test were used to distinguish exactly where the differences are significant. Analysis of Variance Test shows overlapping of confidence intervals between Rif^r donor and mutant. Therefore, both tests presented and supported that there was no significant difference in % survival between Rif^r donor and mutant. All of the other interactions in between the wild type, Tc^r donor, Rif^r donor, and mutant were significant at 95% confidence level. TABLES 5-5 through 6-1 and FIGURE 3-7 present all of the above analysis.

Wild type *E. coli* was exposed to different concentrations of Cd and strain that is cadmium resistant was discovered. *E. coli* strain that is cadmium resistant could grow at the maximum Cd concentration of 4.5 mM. Wild type *E. coli* and Cd resistant mutant were tested for Cd sensitivity. They were grown in LB with and without 4 mM Cd. Wild type *E. coli* grew in LB with no Cd, but demonstrated no growth in LB supplemented with 4 mM Cd. Cadmium resistant *E. coli* strain grew in both, LB with and without Cd.

Table 1-1. Doubling times for *E. coli* absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

	1×10^{-3} M Cd	1×10^{-3} M Pb	Control	Avg
Ec-glu	150	90	60	100
	150	90	60	100
	150	90	60	100
Ec-lac	150	90	30	90
	150	90	30	90
	150	90	60	100

Variable 1: Bacteria and nutrients (glucose/lactose)
Ec- *E.coli*

Variable 2: Treatment

Table 1-2. Anova: Two-Factor With Replication for *E. coli* absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

SUMMARY	1x10-3M	1x10-3M	Control	Total
<i>Ec-glu</i>				
Count	3	3	3	9
Sum	450	270	180	900
Average	150	90	60	100
Variance	0	0	0	1575

<i>Ec-lac</i>				
Count	3	3	3	9
Sum	450	270	120	840
Average	150	90	40	93.3333
Variance	0	0	300	2350

<i>Total</i>			
Count	6	6	6
Sum	900	540	300
Average	150	90	50
Variance	0	0	240

ANOVA						
Source of Variation	SS	df	MS	F	P-value	F crit
Sample	200	1	200	4	0.0687	4.7472
Columns	30400	2	15200	304	5E-11	3.8853
Interaction	400	2	200	4	0.0467	3.8853
Within	600	12	50			
Total	31600	17				

Table 1-3. Duncan's Multiple Range Test for *E. coli* absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

Treatment			Sw
df=2	Value of p	2	3
	SSR	6.09	50 EMS
	LSR	17.6	6 k
	Rp=LSR		8.3 EMS/k
			2.9 Sx

$$Sx = \frac{EMS}{k}$$

$$LSR = Sx * S$$

Media			Sw
df=1	Value of p	2	50 EMS
	SSR	18.0	9 k
	LSR	42.4	5.6 EMS/k
			2.4 Sx

p=number of means for range being tested
Protection level = 0.05

Table 1-4. Mean Comparisons and Rp Comparisons for *E. coli* absorbance vs. time grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

High Cd	High Pb	Control
150.0	90.0	50.0

Ec-glu	Ec-lac
100	93.3

	Difference	LSR		Difference	LSR	
HC-HP	60.0	17.6	42.4	6.7	42.4	-35.7
HC-Co	100.0	17.6	82.4			not sig
HP-Co	40.0	17.6	22.4			

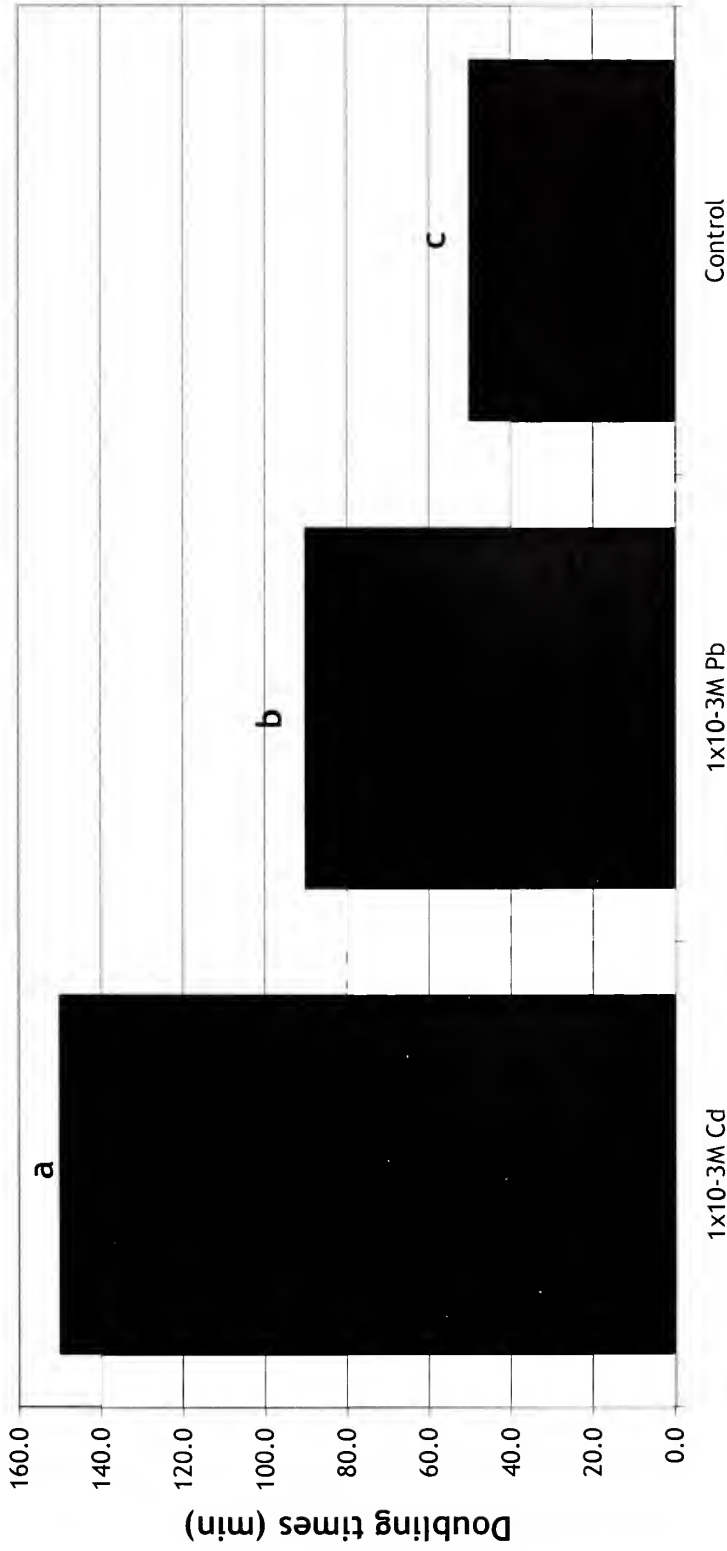
Table 1-5. Summary of doubling times for *E. coli* absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

1×10^{-3} M Cd	1×10^{-3} M Pb	Control	
150.0	90.0	50.0	
Ec-glu	100		
Ec-lac	93.3		
	1×10^{-3} M Cd	1×10^{-3} M Pb	Control
Ec-glu	150	90	60
Ec-lac	150	90	40

Variable 1: Bacteria and nutrients (glucose/lactose)
Ec- *E.coli*

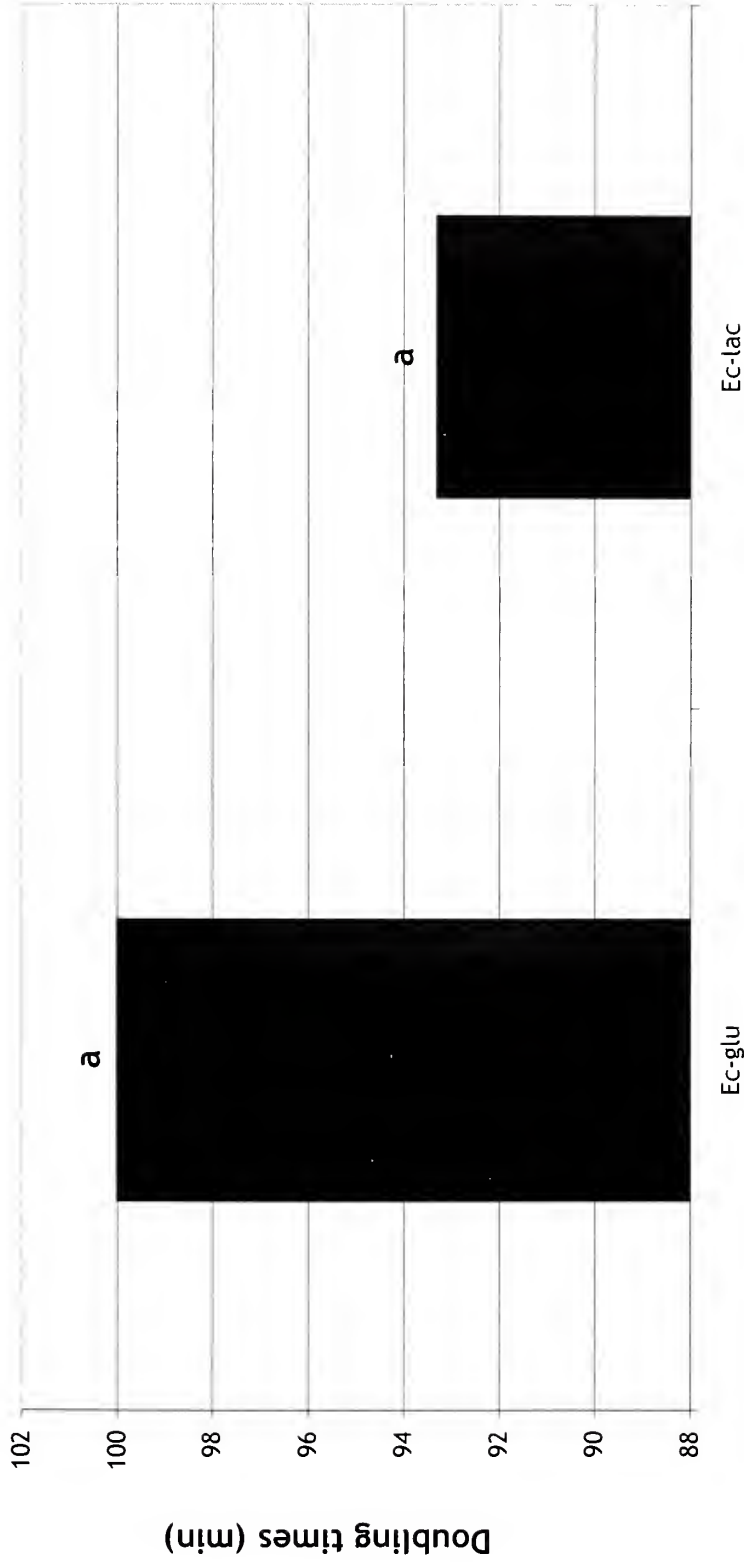
Variable 2: Treatment

Figure 1-1. Average doubling times of *E. coli* vs. Cd and Pb from absorbance vs. time



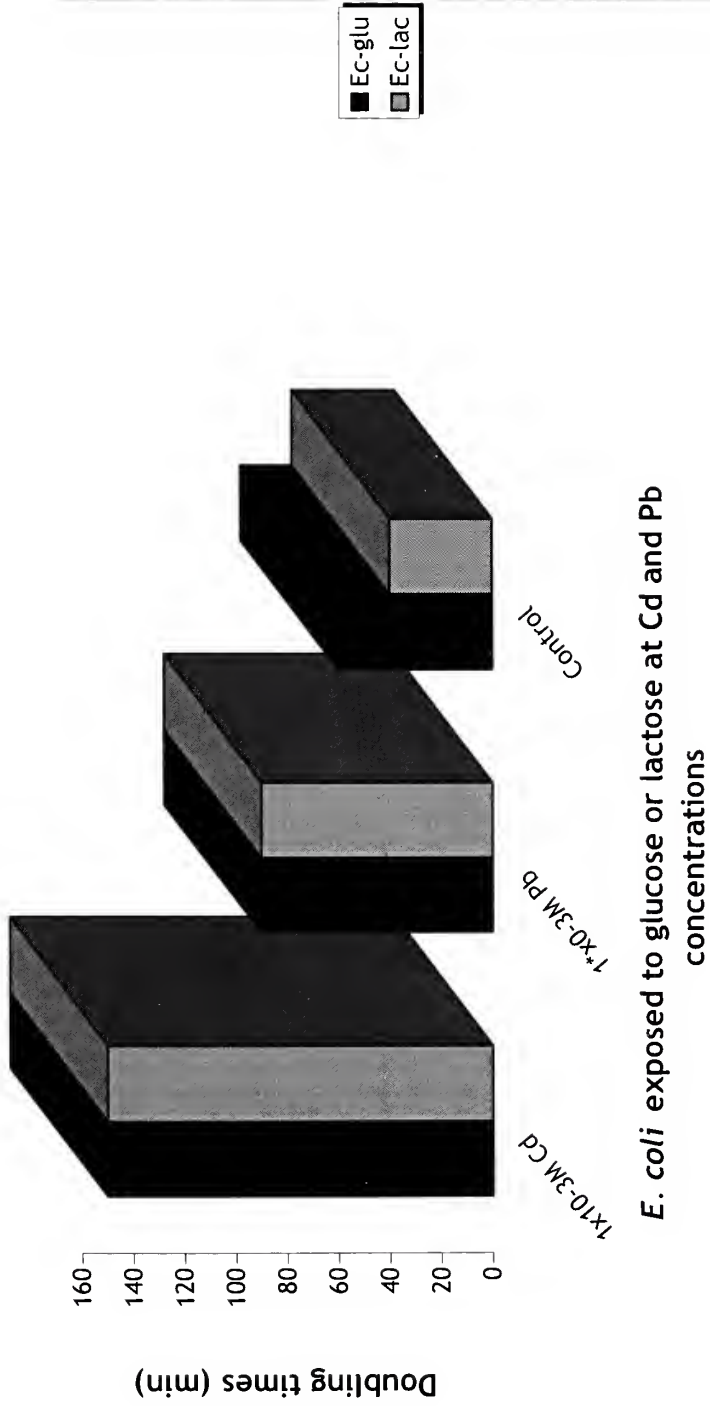
***E. coli* exposed to 1x10⁻³M Cd and Pb**

Figure 1-2. Average doubling times of *E. coli* vs. glucose or lactose from absorbance vs.time



***E. coli* exposed to glucose or lactose**

Figure 1-3. Summary of average doubling times of *E. coli* from absorbance vs. time



E. coli exposed to glucose or lactose at Cd and Pb concentrations

Table 1-6. Doubling times for *M. roseus* absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

	1×10^{-3} M Cd	1×10^{-3} M Pb	Control	Avg
Mr-glu	240	210	150	200
	240	210	150	200
	240	210	150	200
Mr-lac	240	240	120	200
	240	210	120	190
	270	210	150	210
Avg	245	215	140	

Variable 1: Bacteria and nutrients (glucose/lactose)
Mr - *M.roseus*

Variable 2: Treatment

Table 1-7. Anova: Two-Factor With Replication for *M. roseus* absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

SUMMARY	1x10-3M	1x10-3M	Control	Total
<i>Mr-glu</i>				
Count	3	3	3	9
Sum	720	630	450	1800
Average	240	210	150	200
Variance	0	0	0	1575

<i>Mr-lac</i>				
Count	3	3	3	9
Sum	750	600	390	1740
Average	250	200	130	193.333
Variance	300	300	300	2950

<i>Total</i>			
Count	6	6	6
Sum	1470	1230	840
Average	245	205	140
Variance	150	150	240

ANOVA						
Source of Variation	SS	df	MS	F	P-value	F crit
Sample	200	1	200	1.3333	0.2707	4.7472
Columns	33700	2	16850	112.33	2E-08	3.8853
Interaction	700	2	350	2.3333	0.1393	3.8853
Within	1800	12	150			
Total	36400	17				

Table 1-8. Duncan's Multiple Range Test for *M. roseus* absorbance vs. time grown in M9 media with glucose and lactose exposed to 1×10^{-3} M Cd and Pb

Treatment		Sw
df=2	2	150 EMS
SSR	6.09	6 k
LSR	30.5	25.0 EMS/k
Rp=LSR		5.0 Sx

$$Sx = \frac{EMS}{k}$$

Media		Sw
df=1	2	150 EMS
SSR	18.0	9 k
LSR	73.5	16.7 EMS/k
		4.1 Sx

$$LSR = Sx * SSR$$

p=number of means for range being tested
Protection level = 0.05

Table 1-9. Mean Comparisons and Rp Comparisons for *M. roseus* absorbance vs. time grown in M9 media with glucose and lactose exposed to 1×10^{-3} M Cd and Pb

High Cd	High Pb	Control
245.0	215.0	140.0

Mr-glu	Mr-lac
200	200

	Difference	LSR		Difference	LSR				
HC-HP	30.0	30.5	-0.4	not sig	Mr-glu-Mr-lac	0.0	73.5	-73.5	not sig
HC-Co	105.0	30.5	74.5	sig					
HP-Co	75.0	30.5	44.5	sig					

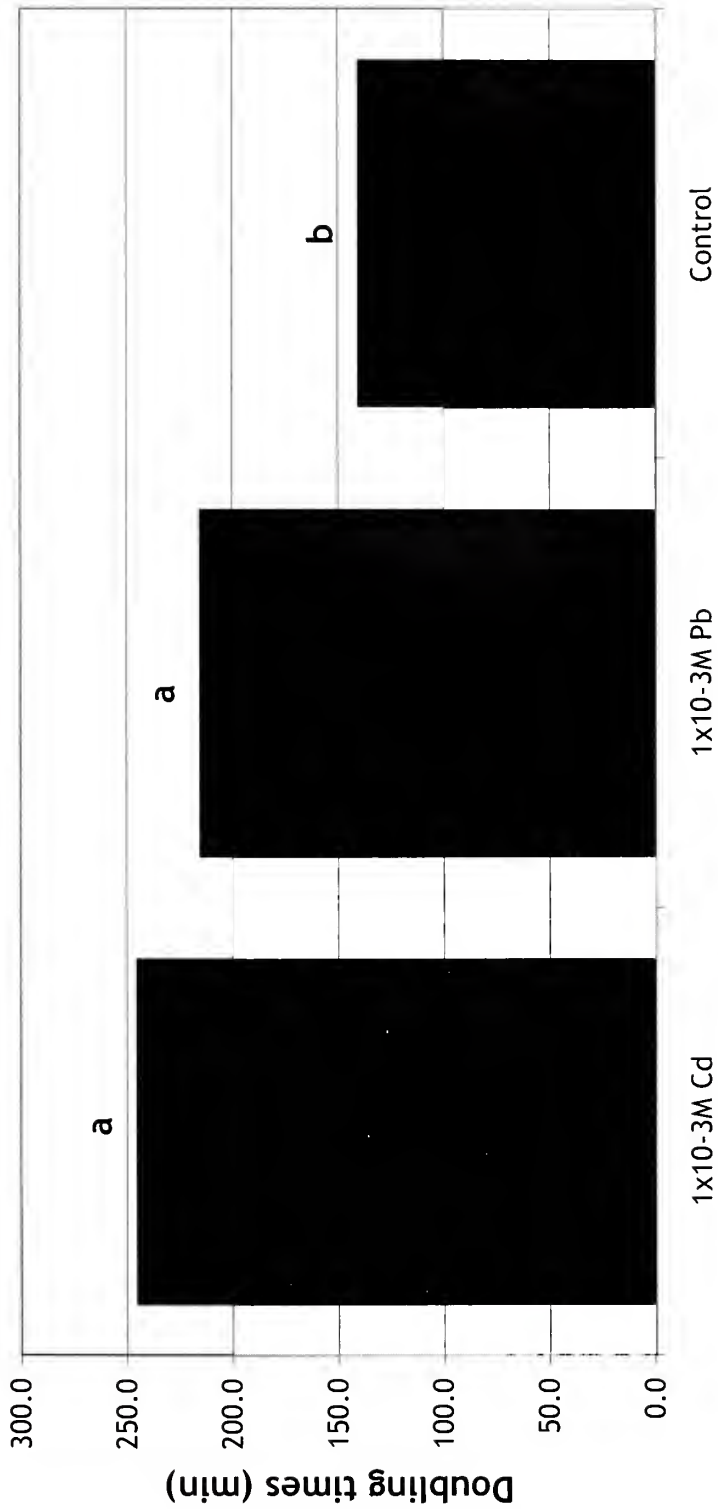
Table 2-1. Summary of doubling times for *M. roseus* absorbance vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

1×10^{-3} M Cd	1×10^{-3} M Pb	Control	
245.0	215.0	140.0	
Mr-glu	200		
Mr-lac	200		
	1×10^{-3} M Cd	1×10^{-3} M Pb	Control
Mr-glu	240	210	150
Mr-lac	250	200	130

Variable 1: Bacteria and nutrients (glucose/lactose)
Mr- *M.roseus*

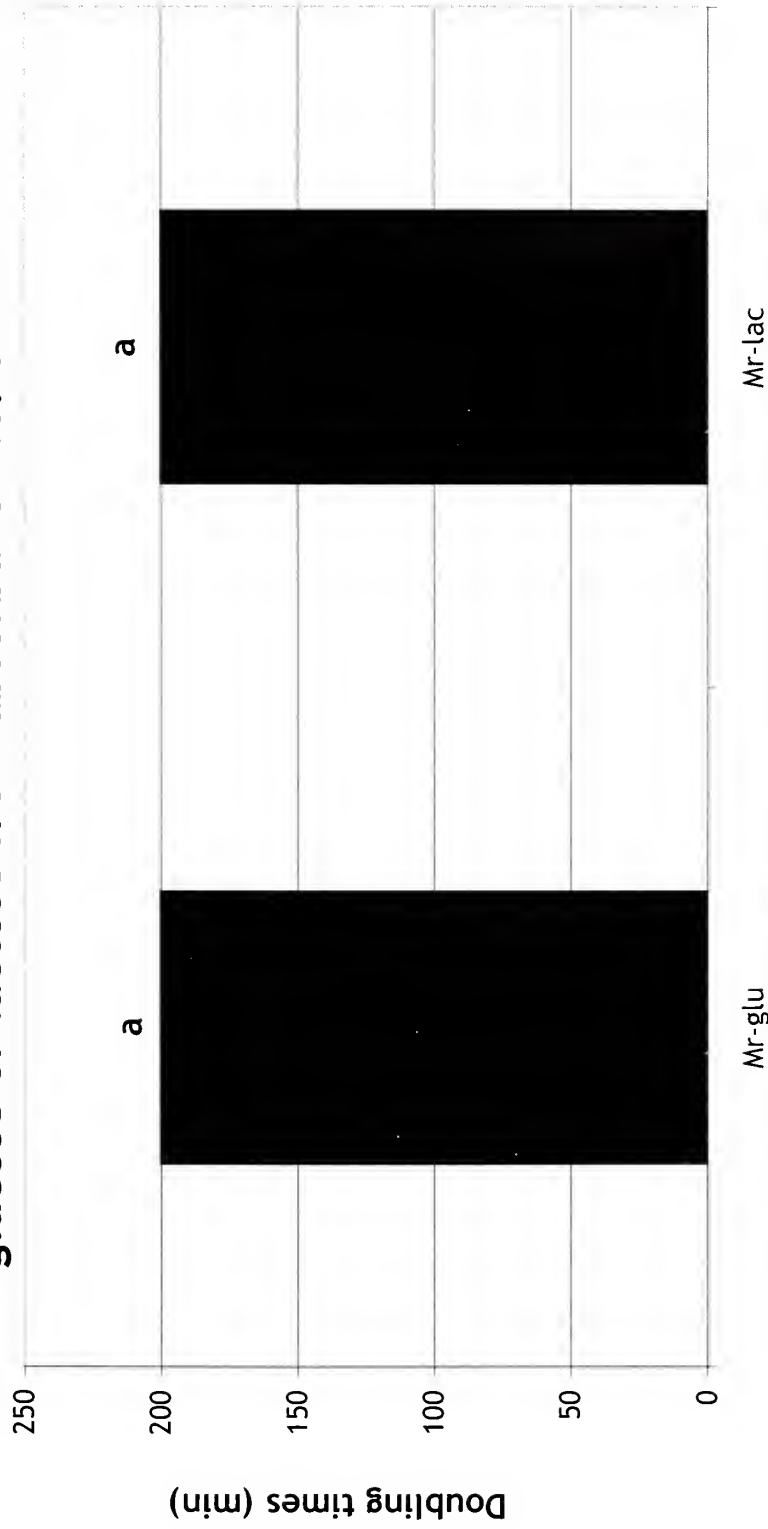
Variable 2: Treatment

Figure 1-4. Average doubling times of *M. roseus* vs. Cd and Pb from absorbance vs. time



M. roseus exposed to 1x10⁻³M Cd and Pb

Figure 1-5. Average doubling times *M. roseus* vs. glucose or lactose from absorbance vs. time



M. roseus exposed to glucose or lactose

Figure 1-6. Summary of average doubling times of *M. roseus* from absorbance vs. time

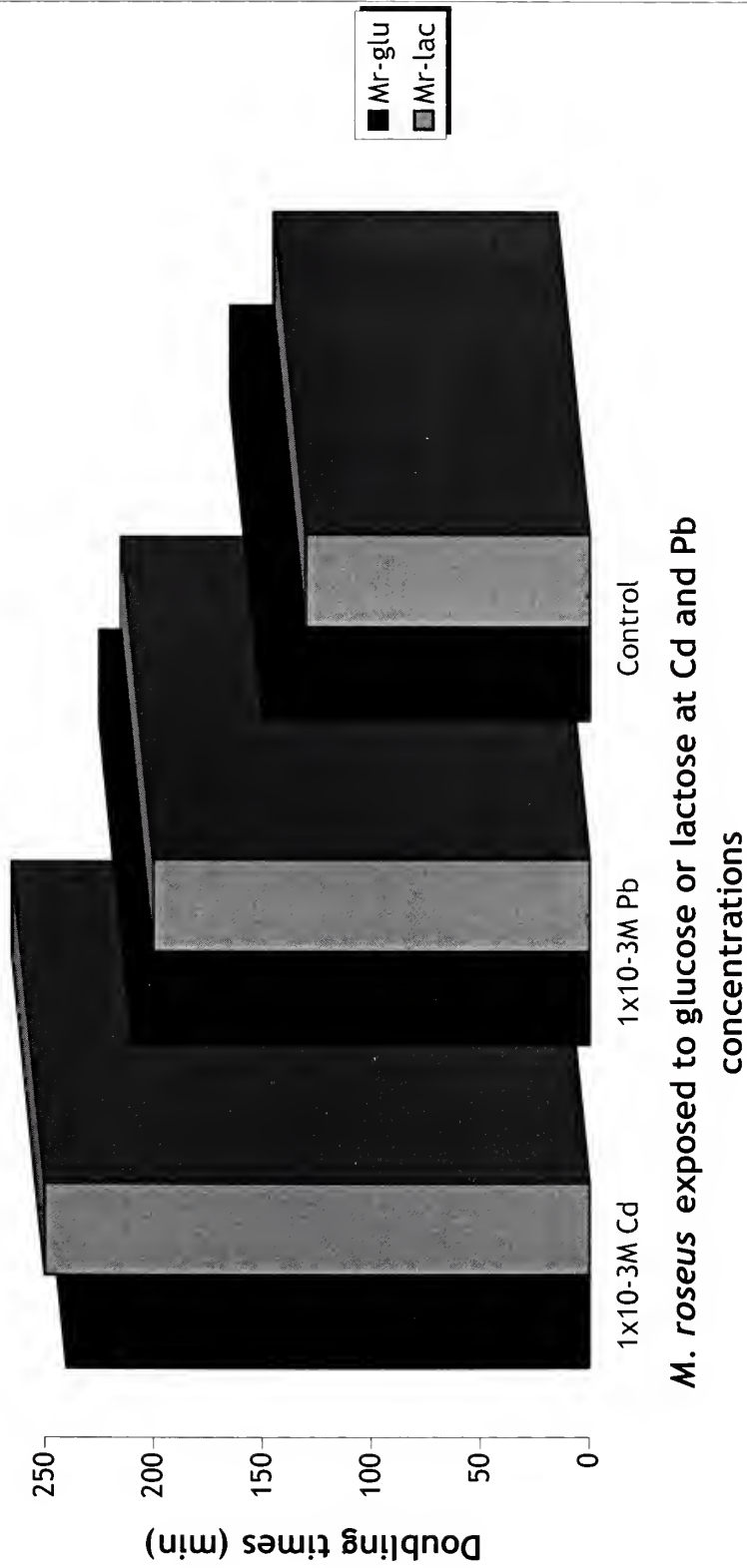


Table 2-2. Doubling times for *E. coli* cell count vs. time in M9 media with glucose or lactose exposed to $1 \times 10^{-3} \text{M}$ Cd and Pb

	Control	$1 \times 10^{-3} \text{M Cd}$	$1 \times 10^{-3} \text{M Pb}$	AVG
Lactose	60	180	120	120.0
	30	210	120	
	30	210	120	
Glucose	90	210	120	173.3
	60	300	210	
	60	300	210	
AVG	55.0	235.0	150.0	

Variable 1: Bacteria and nutrients (glucose/lactose)
Ec- *E.coli*

Variable 2: Treatment

Table 2-3. Two-Factor ANOVA With Replication for *E. coli* cell count vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

Anova: Two-Factor With Replication

SUMMARY	Control	Cd	Pb	Total
<i>Lactose</i>				
Count	3	3	3	9
Sum	120	600	360	1080
Average	40	200	120	120
Variance	300	300	0	4950

<i>Glucose</i>				
Count	3	3	3	9
Sum	210	810	540	1560
Average	70	270	180	173.333
Variance	300	2700	2700	8950

<i>Total</i>				
Count	6	6	6	
Sum	330	1410	900	
Average	55	235	150	
Variance	510	2670	2160	

ANOVA

Source of Variation	SS	df	MS	F	P-value	F crit
Sample	12800	1	12800	12.1905	0.00445	4.74722
Columns	97300	2	48650	46.3333	2.3E-06	3.88529
Interaction	1300	2	650	0.61905	0.5548	3.88529
Within	12600	12	1050			
Total	124000	17				

Table 2-4. Duncan's Multiple Range Test for *E. coli* cell count vs. time grown in M9 media with glucose and lactose exposed to 1×10^{-3} M Cd and Pb

Treatment			Sw	
df=2	Value of p	2	3	1050 EMS
	SSR	6.09	6.09	6 k
	LSR	80.4	80.4	175.0 EMS/k
	Rp=LSR			13.2 Sx

$$LSR = Sx * SSR$$

Nutrient			Sw	
df=1	Value of p	2	9	1050 EMS
	SSR	18.0	116.7	9 k
	LSR	194.4	10.8	116.7 EMS/k
				10.8 Sx

$$Sx = \sqrt{\frac{EMS}{k}}$$

p = number of means for range being tested
 Protection level = 0.05

Table 2-5. Mean Comparisons and Rp Comparisons for *E. coli* cell count vs. time grown in M9 media with glucose and lactose exposed to 1×10^{-3} M Cd and Pb

Cd	Pb	Control
235	150	55

Glucose	Lactose
173.3	120

	Difference	LSR		Difference	LSR	
Cd-Pb	85.0	80.4	4.6	53.3	194.4	-141.1
Pb-Control	95.0	80.4	14.6			not sig
Cd-Control	180.0	80.4	99.6			

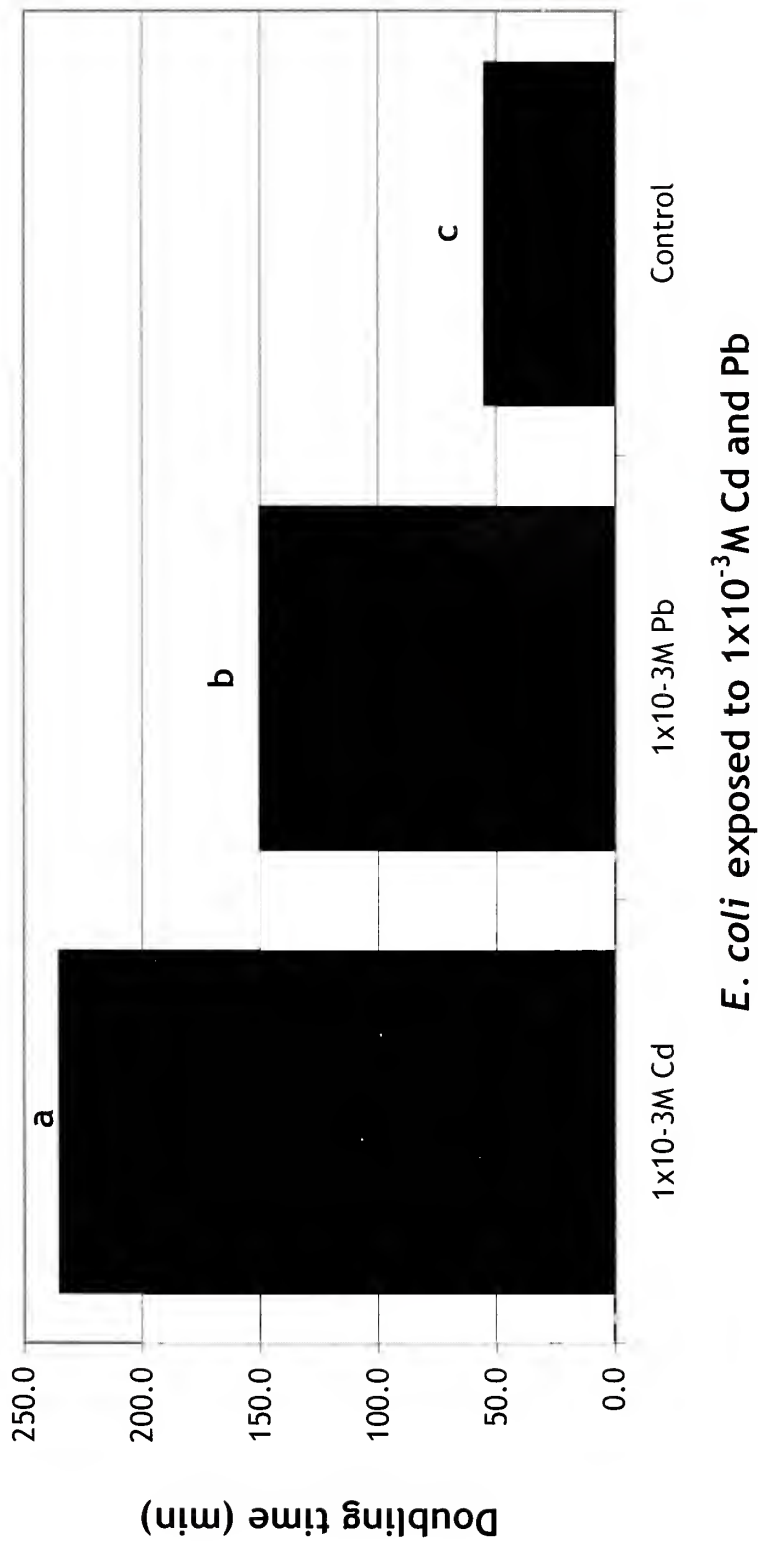
Table 2-6. Summary of doubling times for *E. coli* cell count vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

1×10^{-3} M Cd	1×10^{-3} M Pb	Control	
235.0	150.0	55.0	
Glucose	120		
Lactose	173.3		
	1×10^{-3} M Cd	1×10^{-3} M Pb	Control
Lactose	200	120	40
Glucose	270	180	70

Variable 1: Bacteria and nutrients (glucose/lactose)
Ec- *E.coli*

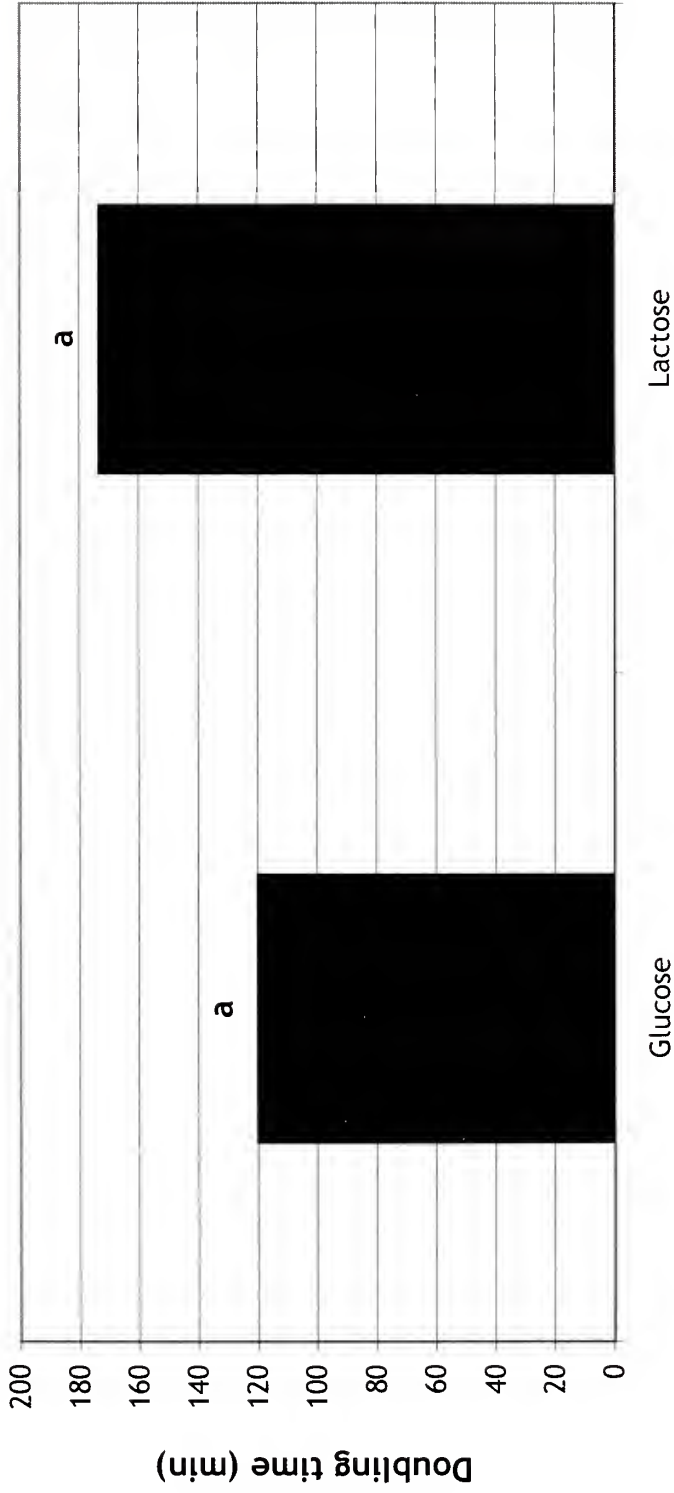
Variable 2: Treatment

Figure 1-7. Average doubling times of *E. coli* vs. Cd and Pb from cell count vs. time



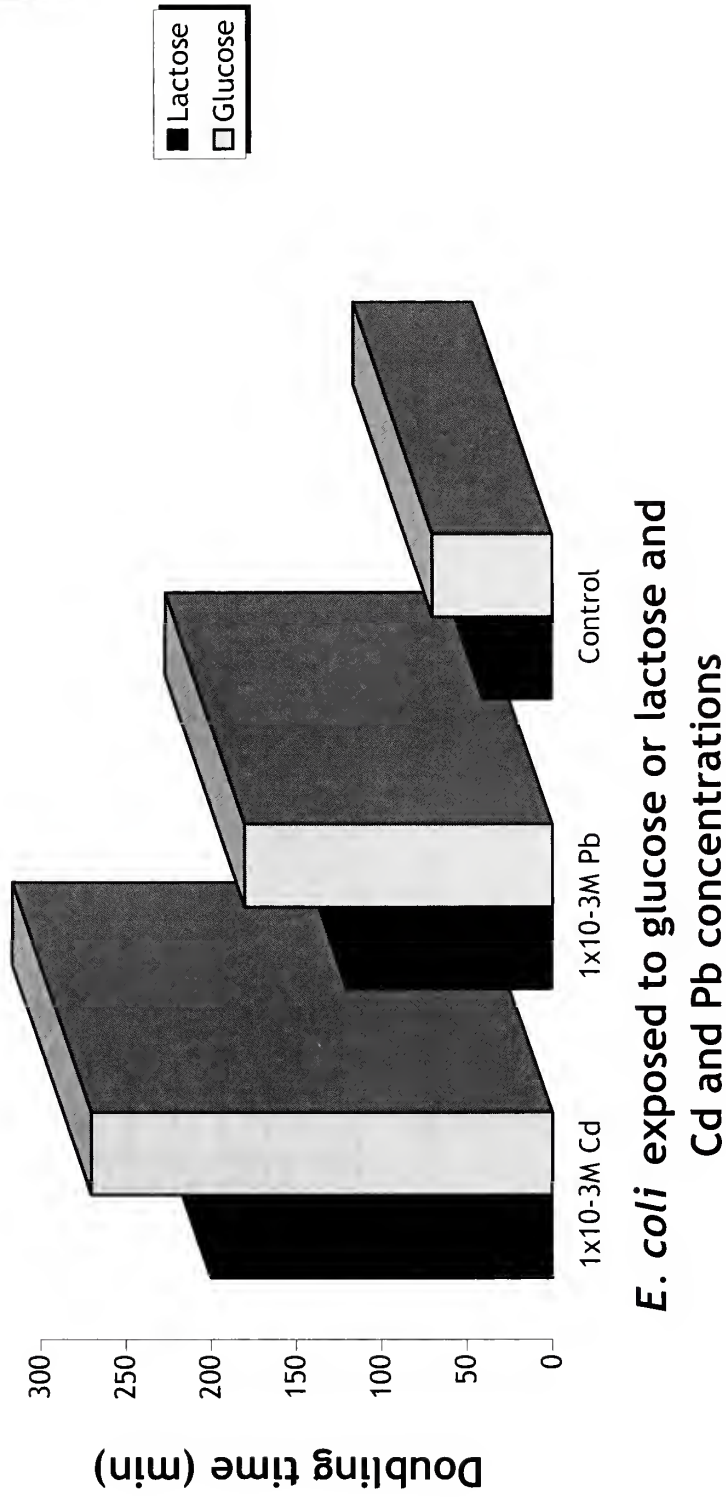
E. coli exposed to 1x10⁻³M Cd and Pb

Figure 1-8. Average doubling times of *E. coli* vs. glucose or lactose from cell count vs. time



E. coli exposed to glucose or lactose

Figure 1-9. Summary of average doubling times of *E. coli* from cell count vs. time



***E. coli* exposed to glucose or lactose and Cd and Pb concentrations**

Table 2-7. Doubling times for *E. coli* cell count vs. absorbance in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

	Control	1×10^{-3} M Cd	1×10^{-3} M Pb	AVG
Lactose	0.077	0.029	0.044	0.050
	0.074	0.022	0.037	
	0.091	0.026	0.046	
Glucose	0.099	0.016	0.048	0.062
	0.109	0.023	0.051	
	0.124	0.027	0.063	
AVG	0.096	0.024	0.048	

Variable 1: Bacteria and nutrients (glucose/lactose)
Ec- *E. coli*

Variable 2: Treatment

Table 2-8. Two-Factor ANOVA With Replication for *E. coli* cell count vs. absorbance in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

Anova: Two-Factor With Replication

SUMMARY	Control	Cd	Pb	Total
<i>Lactose</i>				
Count	3	3	3	9
Sum	0.242	0.077	0.127	0.446
Average	0.08067	0.02567	0.04233	0.04956
Variance	8.2E-05	1.2E-05	2.2E-05	0.00063

<i>Glucose</i>				
Count	3	3	3	9
Sum	0.332	0.066	0.162	0.56
Average	0.11067	0.022	0.054	0.06222
Variance	0.00016	3.1E-05	6.3E-05	0.00158

<i>Total</i>				
Count	6	6	6	
Sum	0.574	0.143	0.289	
Average	0.09567	0.02383	0.04817	
Variance	0.00037	2.1E-05	7.5E-05	

ANOVA

Source of Variation	SS	df	MS	F	P-value	F crit
Sample	0.00072	1	0.00072	11.7292	0.00503	4.74722
Columns	0.01602	2	0.00801	130.1	7.3E-09	3.88529
Interaction	0.00085	2	0.00043	6.92329	0.01002	3.88529
Within	0.00074	12	6.2E-05			
Total	0.01833	17				

Table 2-9. Duncan's Multiple Range Test for *E. coli* cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

Treatment				Sw	
df=2	Value of p	2	3	0.0000616	EMS
	SSR	6.09	6.09	6	k
	LSR	0.01951	0.01951	0.0000103	EMS/k
	Rp=LSR			0.003204	Sx

$$Sx = \frac{EMS}{k}$$

$$LSR = Sx * SSR$$

Nutrient			Sw	
df=1	Value of p	2	0.0000616	EMS
	SSR	18.0	0.0000068	9 k
	LSR	0.0471	0.002616	EMS/k
				Sx

p = number of means for range being tested
 Protection level = 0.05

Table 3-1. Mean Comparisons and Rp Comparisons for *E. coli* cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

Cd	Pb	Control
0.024	0.048	0.096

Glucose	Lactose
0.062	0.05

	Difference	LSR		Difference	LSR	
Cd-Pb	0.02400	0.01951	0.00449	0.0120	0.0471	-0.035 not sig
Pb-Control	0.04800	0.01951	0.02849			
Cd-Control	0.03400	0.01951	0.01449			

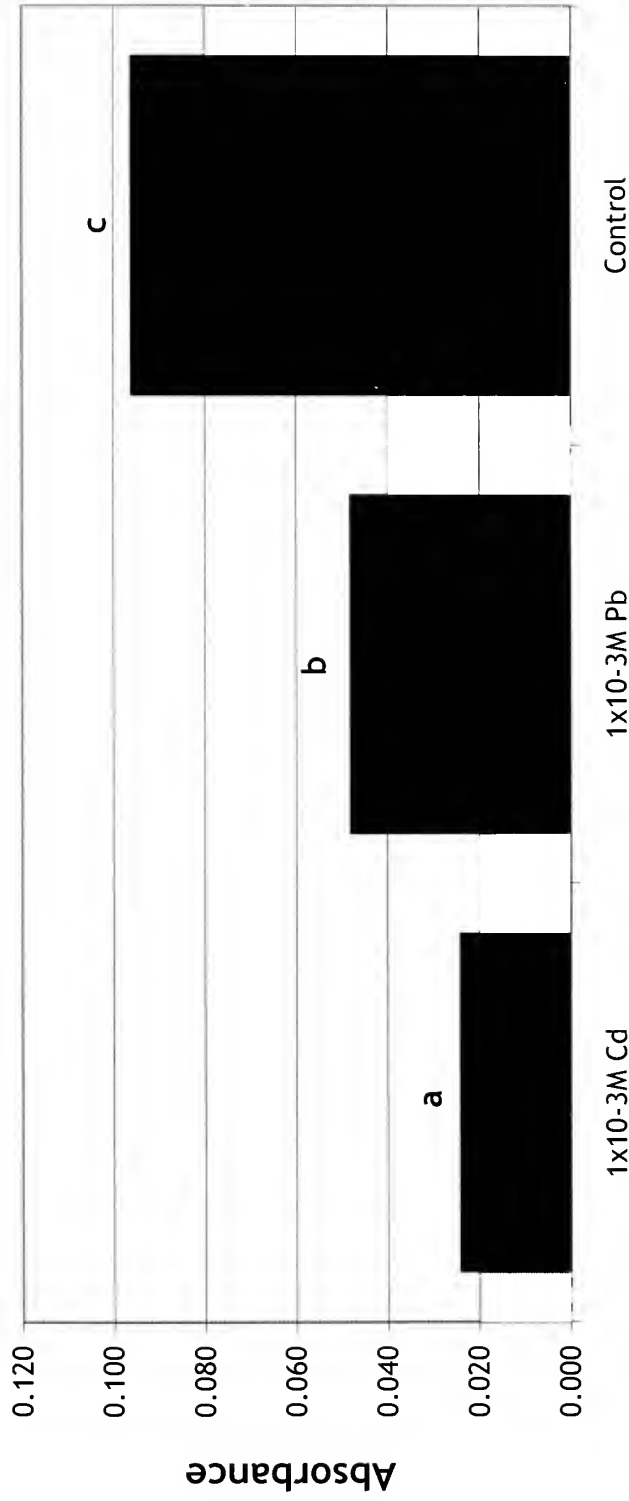
Table 3-2. Summary of doubling times for *E. coli* cell count vs. absorbance in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

1×10^{-3} M Cd	1×10^{-3} M Pb	Control	
0.024	0.048	0.0960	
Glucose	0.062		
Lactose	0.050		
	1×10^{-3} M Cd	1×10^{-3} M Pb	Control
Lactose	0.026	0.042	0.081
Glucose	0.017	0.054	0.111

Variable 1: Bacteria and nutrients (glucose/lactose)
Ec- *E. coli*

Variable 2: Treatment

Figure 2-1. Average absorbance of *E. coli* vs. Cd and Pb from cell count vs. absorbance



E. coli exposed to 1x10⁻³M Cd and Pb

Figure 2-2. Average absorbance of *E. coli* vs. glucose or lactose from cell count vs. absorbance

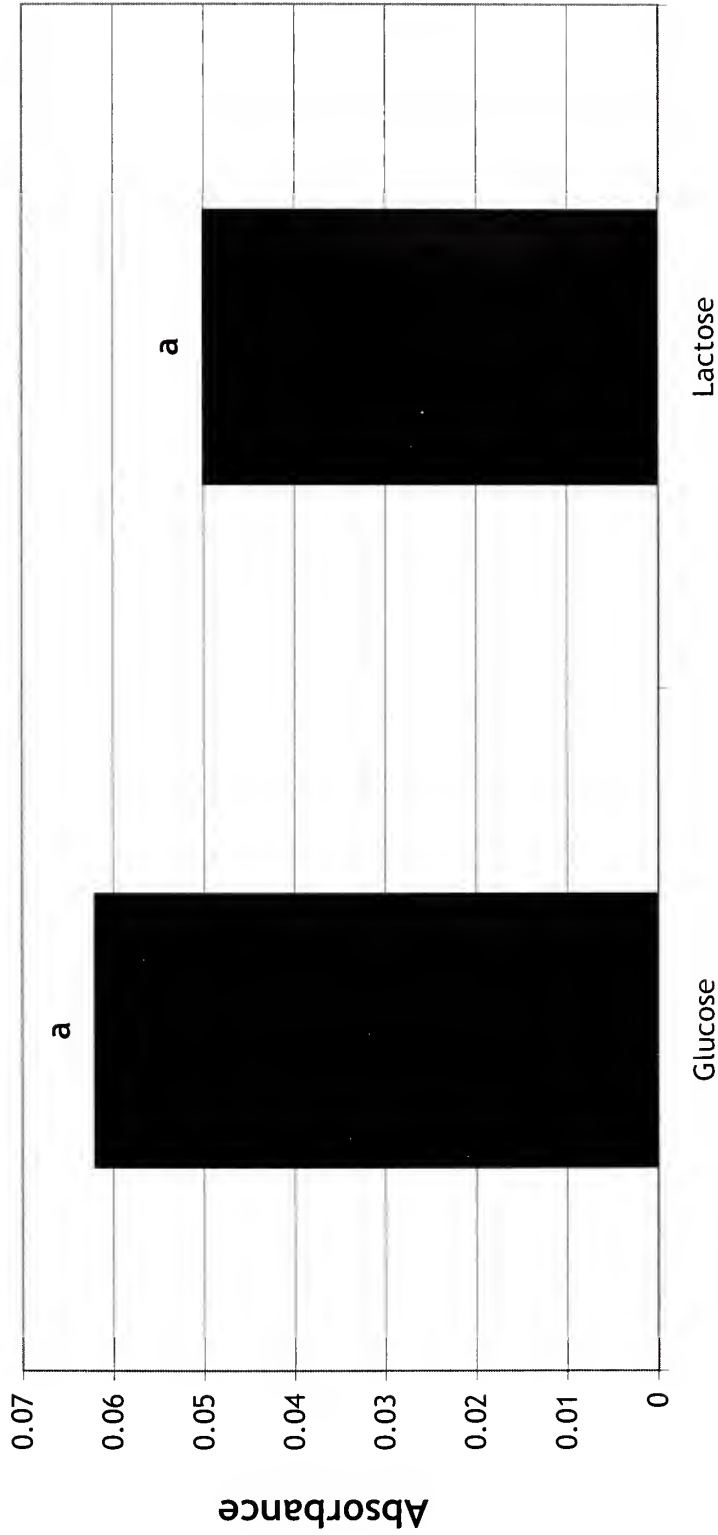


Figure 2-3. Summary of average absorbance's of *E. coli* from cell count vs. absorbance

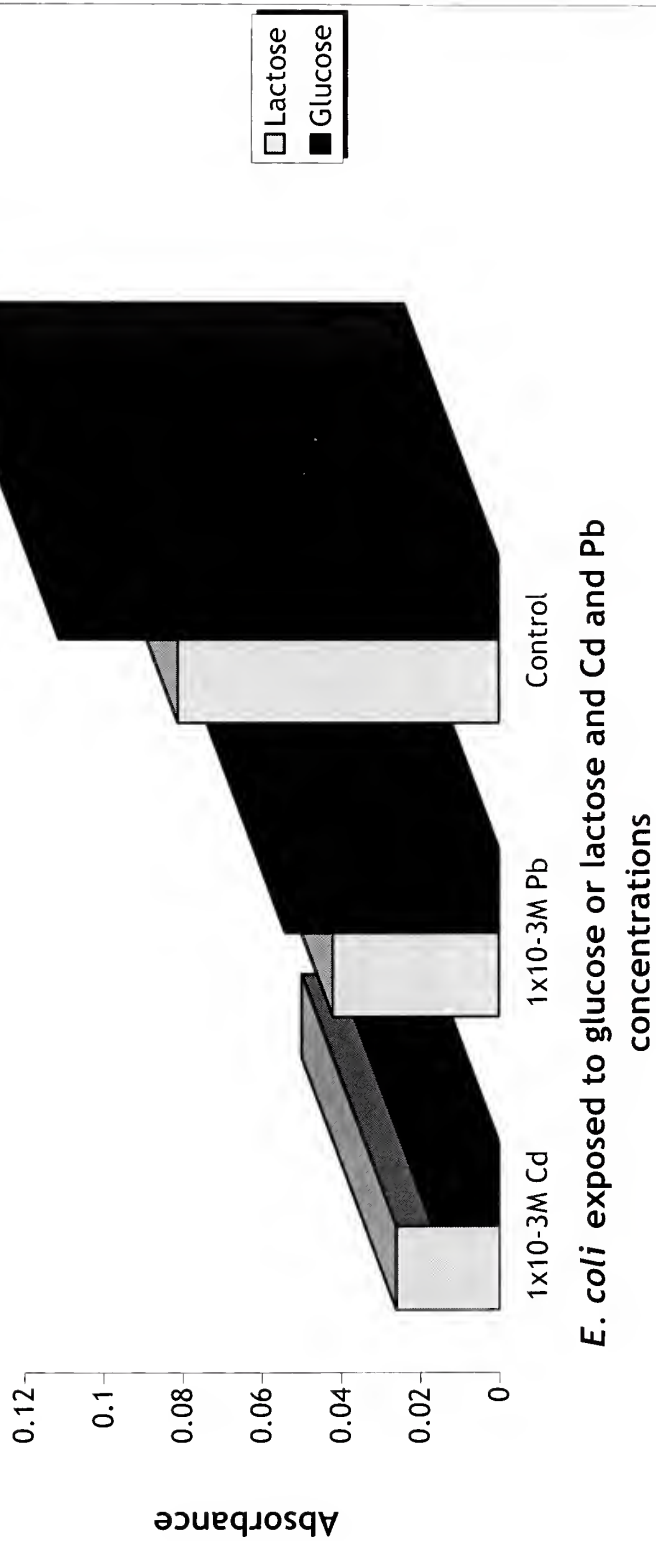


Table 3-3. Doubling times for *M. roseus* cell count vs. time in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

	Control	1×10^{-3} M Cd	1×10^{-3} M Pb	AVG
Lactose	60	300	180	190.0
	60	300	180	
	120	180	180	
	120	420	180	
Glucose	120	660	660	410.0
	60	540	600	
	60	360	300	
	180	720	660	
AVG	97.5	435.0	367.5	

Variable 1: Bacteria and nutrients (glucose/lactose)
 Mr- *M. roseus*

Variable 2: Treatment

Table 3-4. Two-Factor ANOVA With Replication for *M. roseus* cell count vs. time in M9 media with glucose or lactose exposed to $1 \times 10^{-3} \text{M}$ Cd and Pb

Anova: Two-Factor With Replication

SUMMARY	Control	Cd	Pb	Total
<i>Lactose</i>				
Count	4	4	4	12
Sum	360	1200	720	2280
Average	90	300	180	190
Variance	1200	9600	0	11018.2

<i>Glucose</i>				
Count	4	4	4	12
Sum	420	2280	2220	4920
Average	105	570	555	410
Variance	3300	25200	29700	66654.5

<i>Total</i>				
Count	8	8	8	
Sum	780	3480	2940	
Average	97.5	435	367.5	
Variance	1992.857	35742.9	52907.1	

ANOVA

Source of Variation	SS	df	MS	F	P-value	F crit
Sample	290400	1	290400	25.2522	8.8E-05	4.41386
Columns	510300	2	255150	22.187	1.4E-05	3.55456
Interaction	137100	2	68550	5.96087	0.01032	3.55456
Within	207000	18	11500			
Total	1144800	23				

Table 3-5. Duncan's Multiple Range Test for *M. roseus* cell count vs. time grown in M9 media with glucose and lactose exposed to $1 \times 10^{-3} \text{M}$ Cd and Pb

Treatment				Sw
df=2	Value of p	2	3	11500 EMS
	SSR	6.09	6.09	9 k
	LSR	217.4	217.4	1277.8 EMS/k
	Rp=LSR			35.7 Sx

Nutrient			Sw
df=1	Value of p	2	11500 EMS
	SSR	18.0	12 k
	LSR	558.0	958.3 EMS/k
			31.0 Sx

$$Sx = \sqrt{\frac{EMS}{k}}$$

p=number of means for range being tested
 Protection level = 0.05

$$LSR = Sx * SSR$$

Table 3-6. Mean Comparisons and Rp Comparisons for *M. roseus* cell count vs. time grown in M9 media with glucose and lactose exposed to 1×10^{-3} M Cd and Pb

Cd	Pb	Control
435.00	367.50	97.50

Glucose	Lactose
410	190

	Difference	LSR		Difference	LSR	
Cd-Pb	67.5	217.4	-149.9	220	558.0	-338.00 not sig
Pb-Control	270.0	217.4	52.6			sig
Cd-Control	337.5	121.8	215.7			sig

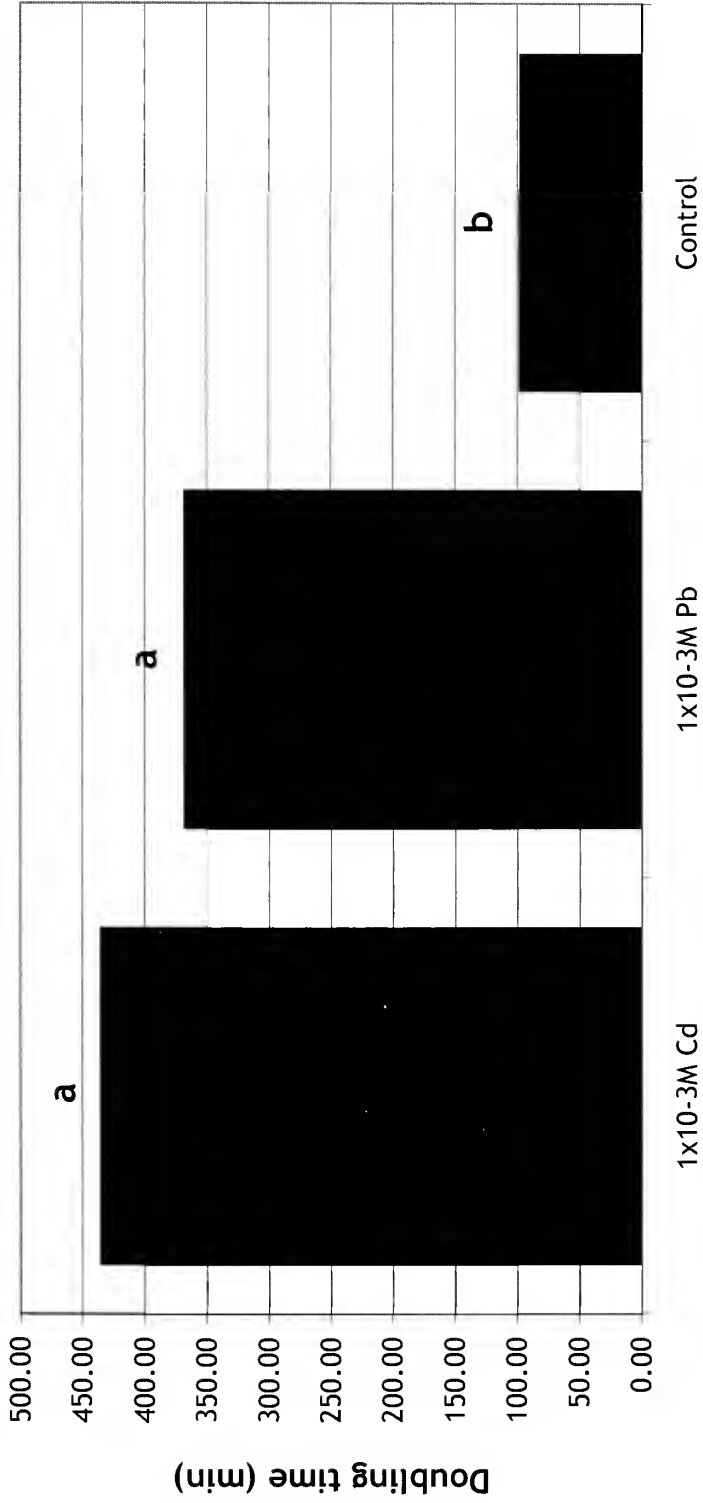
Table 3-7. Summary of doubling times for *M. roseus* cell count vs. time in M9 media with glucose or lactose exposed to $1 \times 10^{-3} \text{M}$ Cd and Pb

$1 \times 10^{-3} \text{M}$ Cd	$1 \times 10^{-3} \text{M}$ Pb	Control	
435.00	367.50	97.50	
Glucose			
	410		
Lactose			
	190		
Variable 1: Bacteria and nutrients (glucose/lactose)			
	$1 \times 10^{-3} \text{M}$ Cd	$1 \times 10^{-3} \text{M}$ Pb	Control
Lactose	300	180	90
Glucose	570	555	105

Variable 1: Bacteria and nutrients (glucose/lactose)
 Mr- *M. roseus*

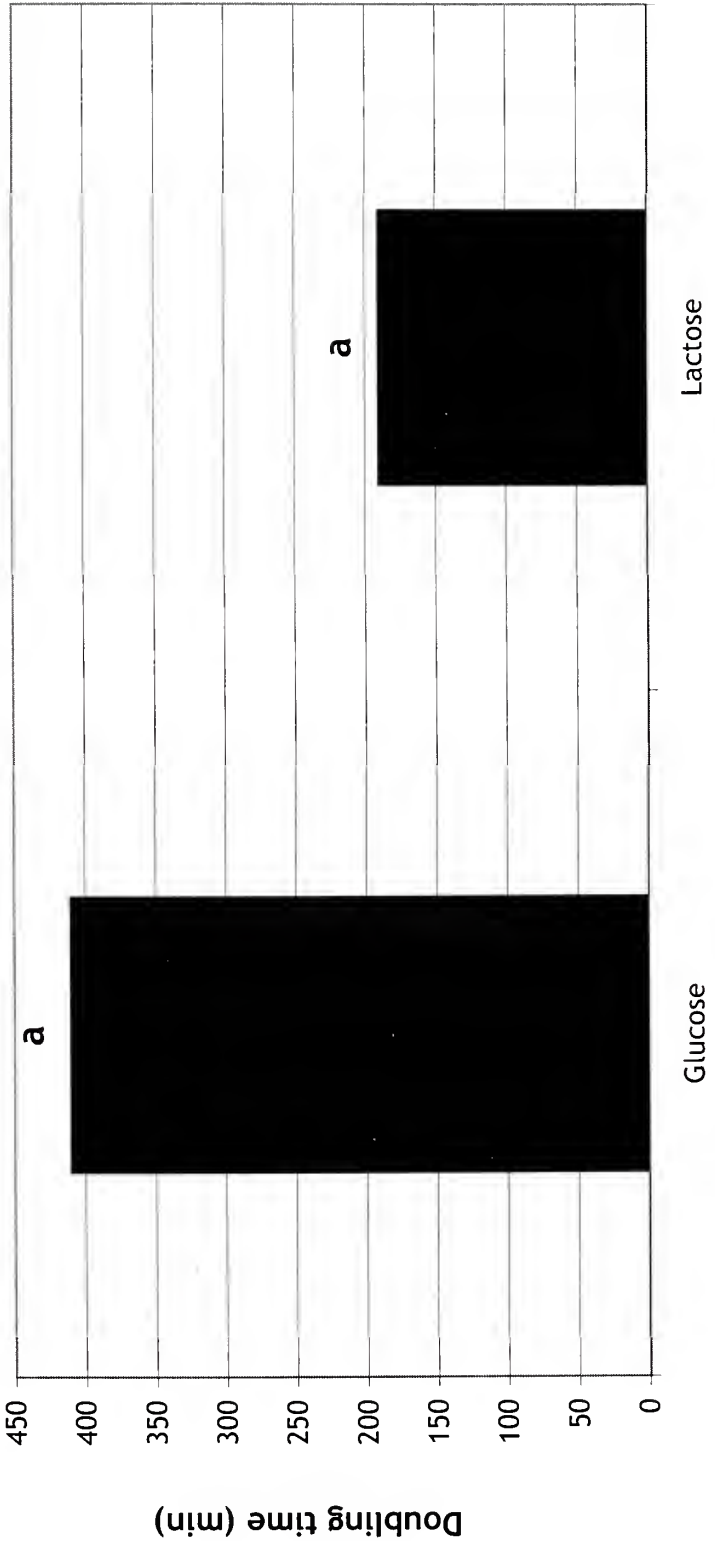
Variable 2: Treatment

Figure 2-4. Average doubling times of *M. roseus* vs. Cd and Pb from cell count vs. time



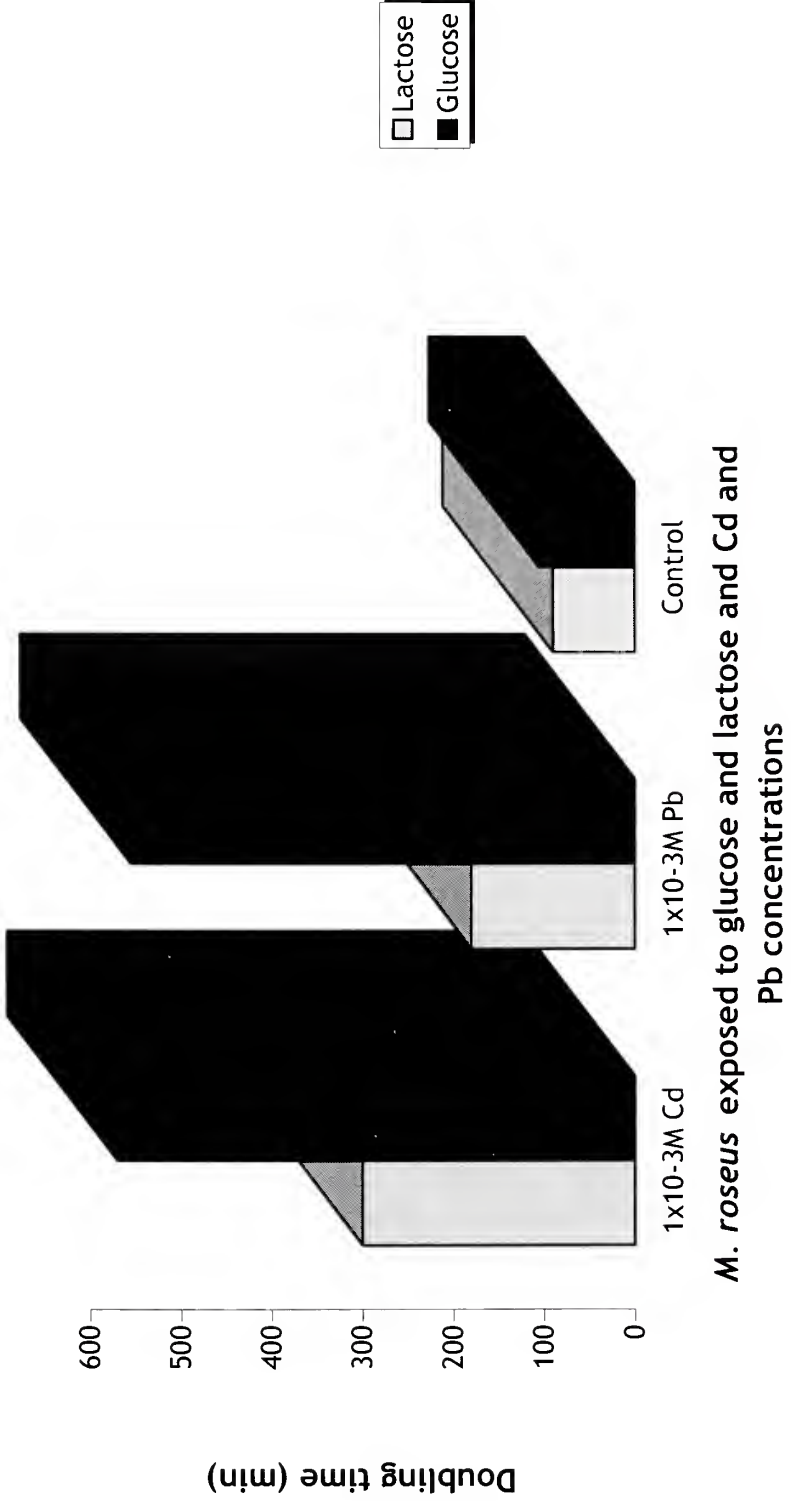
M. roseus exposed to 1x10⁻³M Cd and Pb

Figure 2-5. Average doubling times of *M. roseus* vs. glucose or lactose from cell count vs. time



M. roseus exposed to glucose or lactose

Figure 2-6. Summary of average doubling times of *M. roseus* from cell count vs. time



M. roseus exposed to glucose and lactose and Cd and Pb concentrations

Table 3-8. Doubling times for *M. roseus* cell count vs. absorbance in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

	Control	1×10^{-3} M Cd	1×10^{-3} M Pb	AVG
Lactose	0.138	0.047	0.057	0.080
	0.104	0.05	0.079	
	0.104	0.06	0.077	
Glucose	0.078	0.069	0.053	0.060
	0.077	0.04	0.058	
	0.071	0.038	0.057	
AVG	0.095	0.051	0.064	

Variable 1: Bacteria and nutrients (glucose/lactose)
 Mr- *M. roseus*

Variable 2: Treatment

Table 3-9. Two-Factor ANOVA With Replication for *M. roseus* cell count vs. absorbance in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

Anova: Two-Factor With Replication

SUMMARY	Control	Cd	Pb	Total
<i>Lactose</i>				
Count	3	3	3	9
Sum	0.346	0.157	0.213	0.716
Average	0.11533	0.05233	0.071	0.07956
Variance	0.00039	4.6E-05	0.00015	0.00093

<i>Glucose</i>				
Count	3	3	3	9
Sum	0.226	0.147	0.168	0.541
Average	0.07533	0.049	0.056	0.06011
Variance	1.4E-05	0.0003	7E-06	0.00022

<i>Total</i>				
Count	6	6	6	
Sum	0.572	0.304	0.381	
Average	0.09533	0.05067	0.0635	
Variance	0.00064	0.00014	0.00013	

ANOVA

Source of Variation	SS	df	MS	F	P-value	F crit
Sample	0.0017	1	0.0017	11.3174	0.005631	4.74722
Columns	0.00635	2	0.00317	21.1075	0.000156	3.88529
Interaction	0.00105	2	0.00053	3.50148	0.06341	3.88529
Within	0.0018	12	0.00015			
Total	0.0109	17				

Table 4-1. Duncan's Multiple Range Test for *M. roseus* cell count vs. absorbance grown in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

Treatment				Sw
df=2	Value of p	2	3	0.00015 EMS
	SSR	6.09	6.09	6 k
	LSR	0.03048	0.03048	0.00002505 EMS/k
	Rp=LSR			0.005005 Sx

Nutrient			Sw
df=1	Value of p	2	0.00015 EMS
	SSR	18.0	9 k
	LSR	0.0736	0.00002 EMS/k
			0.00409 Sx

$$Sx = \frac{\overline{EMS}}{k}$$

$$LSR = Sx * SSR$$

p=number of means for range being tested
 Protection level = 0.05

Table 4-2. Mean Comparisons and Rp Comparisons for *M. roseus* cell count vs. absorbance grown in M9 media with glucose or lactose exposed to $1 \times 10^{-3} \text{M}$ Cd and Pb

Cd	Pb	Control
0.051	0.064	0.0950

Glucose	Lactose
0.06	0.08

	Difference	LSR		Difference	LSR	
Cd-Pb	0.01300	0.03048	-0.01748	0.0200	0.0736	-0.054 not sig
Pb-Control	0.03100	0.03048	0.00052	Glucose-Lactose		
Cd-Control	0.04400	0.03048	0.01352			

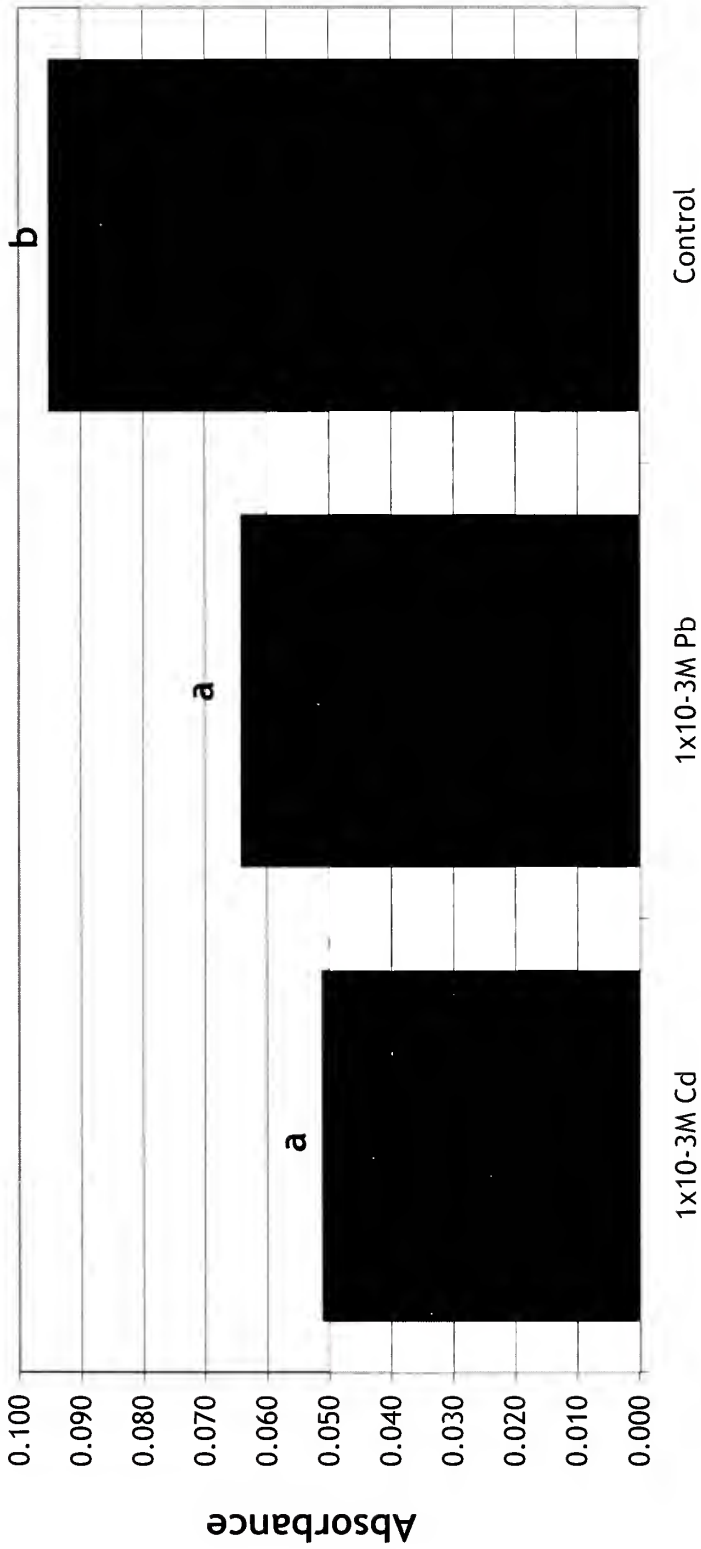
Table 4-3. Summary of doubling times for *M. roseus* cell count vs. absorbance in M9 media with glucose or lactose exposed to 1×10^{-3} M Cd and Pb

1×10^{-3} M Cd	1×10^{-3} M Pb	Control
0.051	0.064	0.0950
Glucose		
	0.06	
Lactose		
	0.080	
Control		
	1×10^{-3} M Cd	1×10^{-3} M Pb
Lactose	0.052	0.071
Glucose	0.049	0.056
		0.115
		0.075

Variable 1: Bacteria and nutrients (glucose/lactose)
 Mr- *M. roseus*

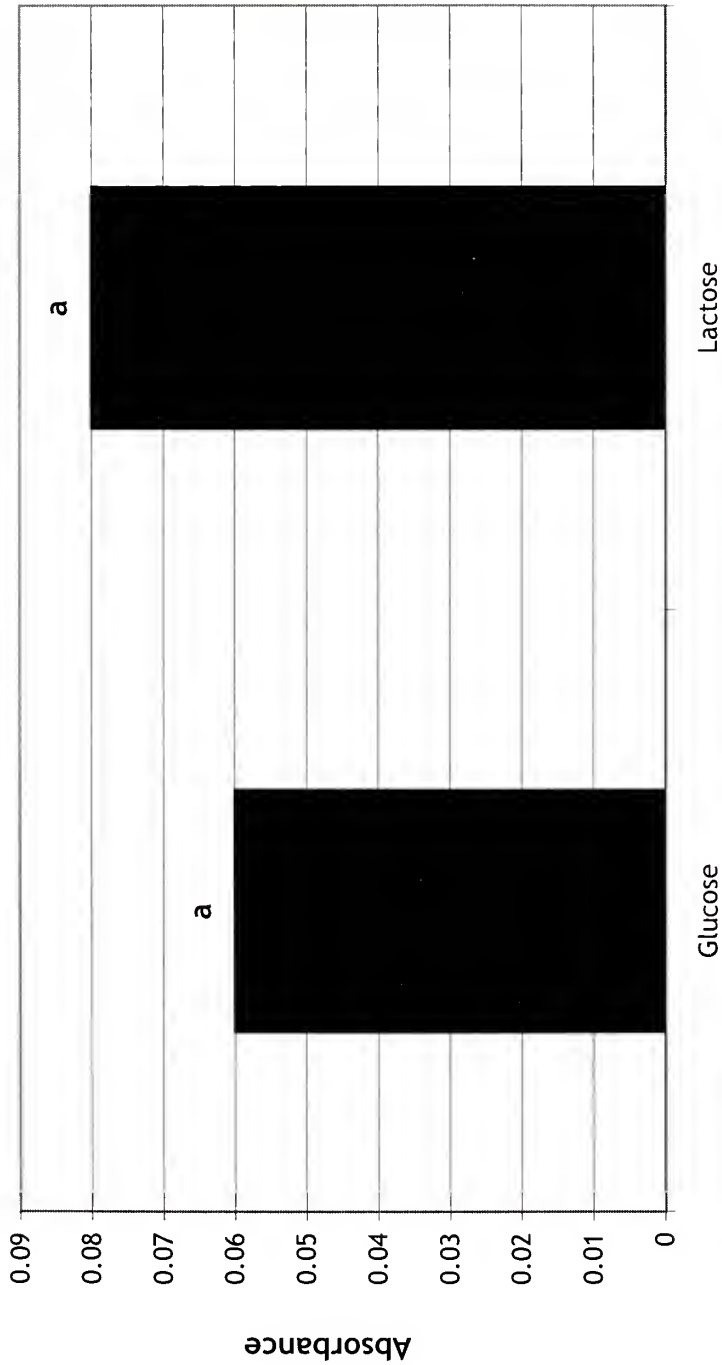
Variable 2: Treatment

Figure 2-7. Average absorbances of *M. roseus* vs. Cd and Pb from cell count vs. absorbance



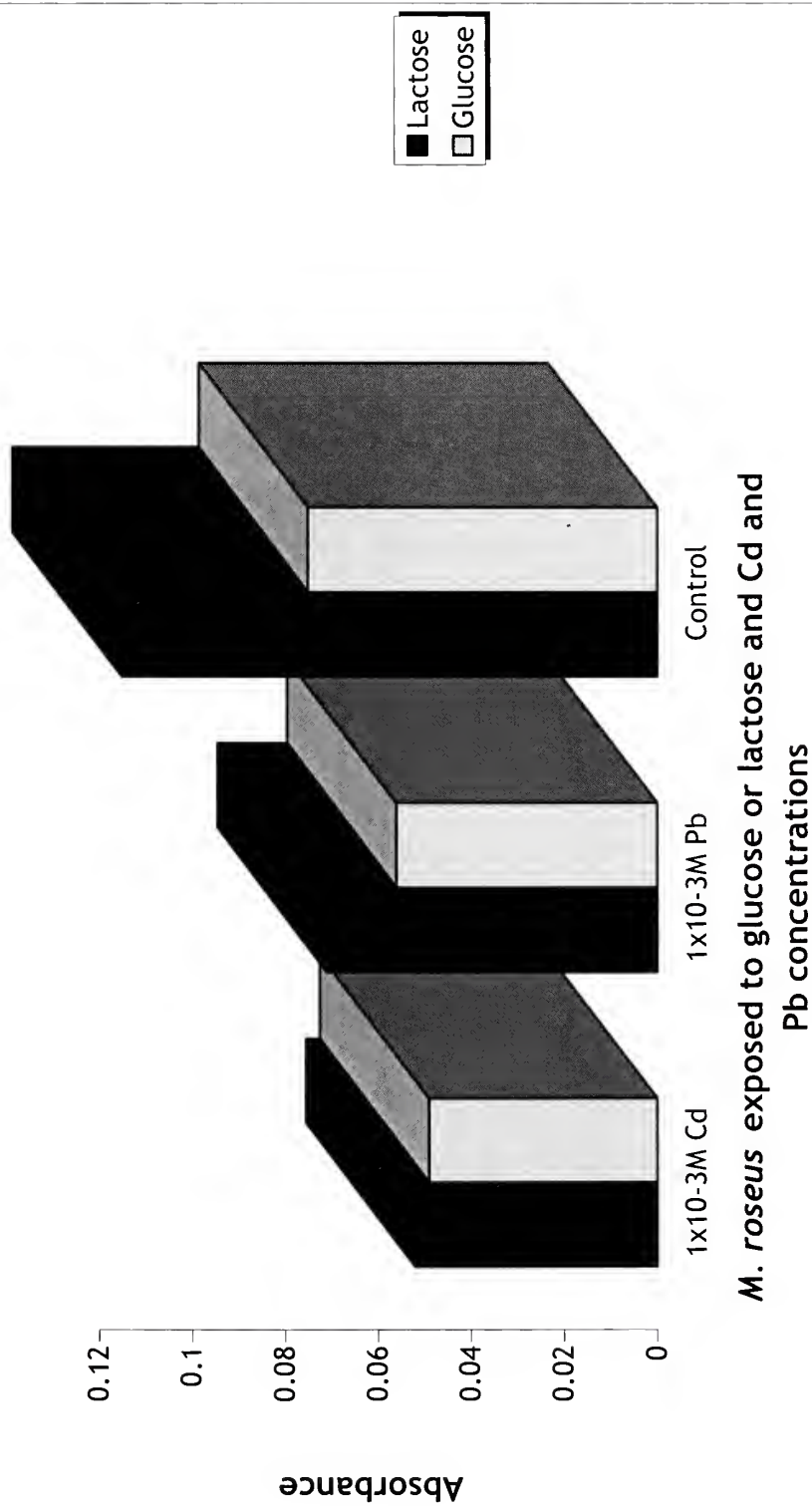
***M. roseus* exposed to 1x10⁻³M Cd and Pb**

Figure 2-8. Average absorbances of *M. roseus* vs. glucose or lactose from cell count vs. absorbance



M. roseus exposed to glucose or lactose

Figure 2-9. Summary of average absorbance's of *M.roseus* from cell count vs. absorbance



M. roseus exposed to glucose or lactose and Cd and Pb concentrations

Table 4-4. Doubling times for *M. roseus* absorbance vs. time in M9 and LB media exposed to $1 \times 10^{-3} \text{M}$ and $1 \times 10^{-5} \text{M}$ Cd and Pb

	$1 \times 10^{-3} \text{M}$ Cd	$1 \times 10^{-3} \text{M}$ Pb	$1 \times 10^{-5} \text{M}$ Cd	$1 \times 10^{-5} \text{M}$ Pb	Control	Avg
M1-M1	240	210	180	150	150	186
	240	210	180	150	150	186
	240	210	210	180	150	198
M1-M2	240	240	180	120	120	180
	240	180	180	120	120	168
	300	240	210	180	120	210
Avg	250	215	190	150	135	

Variable 1: Bacteria and Media

M1= Control medium M9 media

M2= Control rich LB media

Variable 2: Treatment

Table 4-5. Anova: Two-Factor With Replication for *M. roseus* absorbance vs. time in M9 and LB media exposed to $1 \times 10^{-3} \text{M}$ and $1 \times 10^{-5} \text{M}$ Cd and Pb

SUMMARY	$1 \times 10^{-3} \text{M}$	$1 \times 10^{-3} \text{M}$	$1 \times 10^{-5} \text{M}$	$1 \times 10^{-5} \text{M}$	Control	Total
<i>Mr-M1</i>						
Count	3	3	3	3	3	15
Sum	720	630	570	480	450	2850
Average	240	210	190	160	150	190
Variance	0	0	300	300	0	1242.86
<i>Mr-M2</i>						
Count	3	3	3	3	3	15
Sum	780	660	570	420	360	2790
Average	260	220	190	140	120	186
Variance	1200	1200	300	1200	0	3368.57
<i>Total</i>						
Count	6	6	6	6	6	
Sum	1500	1290	1140	900	810	
Average	250	215	190	150	135	
Variance	600	510	240	720	270	

ANOVA						
Source of Variation	SS	df	MS	F	P-value	F crit
Sample	120	1	120	0.2667	0.6112	4.3513
Columns	52980	4	13245	29.433	4E-08	2.8661
Interaction	2580	4	645	1.4333	0.2596	2.8661
Within	9000	20	450			
Total	64680	29				

Table 4-6. Duncan's Multiple Range Test for *M. roseus* absorbance vs. time grown in M9 and LB media exposed to $1 \times 10^{-3} \text{M}$ and $1 \times 10^{-5} \text{M}$ Cd and Pb

Treatment					Sw	
df=4	Value of p	2	3	4	5	450 EMS
	SSR	3.9	4.0	4.0	4.0	6 k
	LSR	34.0	34.7	34.8	34.8	75.0 EMS/k
	Rp=LSR					8.66 Sx

$$Sx = \frac{EMS}{k}$$

Bacteria Type and Media		Sw
df=1	Value of p	450 EMS
	SSR	15 k
	LSR	30.0 EMS/k
		5.5 Sx

p=number of means for range being tested
 Protection level = 0.05

$$LSR = Sx * SSR$$

Table 4-7. Mean Comparisons and Rp Comparisons for *M. roseus* absorbance vs. time grown in M9 and LB media exposed to 1×10^{-3} M and 1×10^{-5} M Cd and Pb

High Cd	High Pb	Low Cd	Low Pb	Control
250.0	215.0	190.0	150.0	135.0
Mr-M1	Mr-M2			
190	190			

	Difference	LSR			Difference	LSR		
HC-HP	35.0	34.0	1.0	sig	Mr-M1-Mr-M2	98.6	-98.6	not sig
HC-LC	60.0	34.7	25.3	sig				
HC-LP	100.0	34.8	65.2	sig				
HC-Co	115.0	34.8	80.2	sig				
HP-LC	25.0	34.0	-9.0	not sig				
HP-LP	65.0	34.7	30.3	sig				
HP-Co	80.0	34.8	45.2	sig				
LC-LP	40.0	34.0	6.0	sig				
LC-Co	55.0	34.7	20.3	sig				
LP-Co	15.0	34.0	-19.0	not sig				

Table 4-8. Summary of doubling times for *M. roseus* absorbance vs. time in M9 and LB media exposed to $1 \times 10^{-3} \text{M}$ and $1 \times 10^{-5} \text{M}$ Cd and Pb

$1 \times 10^{-3} \text{M}$ Cd	$1 \times 10^{-3} \text{M}$ Pb	$1 \times 10^{-5} \text{M}$ Cd	$1 \times 10^{-5} \text{M}$ Pb	Control				
250.0	215.0	190.0	150.0	135.0				
<table border="1"> <tbody> <tr> <td>Mr-M1</td> <td>190</td> </tr> <tr> <td>Mr-M2</td> <td>190</td> </tr> </tbody> </table>					Mr-M1	190	Mr-M2	190
Mr-M1	190							
Mr-M2	190							
	$1 \times 10^{-3} \text{M}$ Cd	$1 \times 10^{-3} \text{M}$ Pb	$1 \times 10^{-5} \text{M}$ Cd	$1 \times 10^{-5} \text{M}$ Pb	Control			
Mr-M1	240	210	190	160	150			
Mr-M2	260	220	190	160	120			

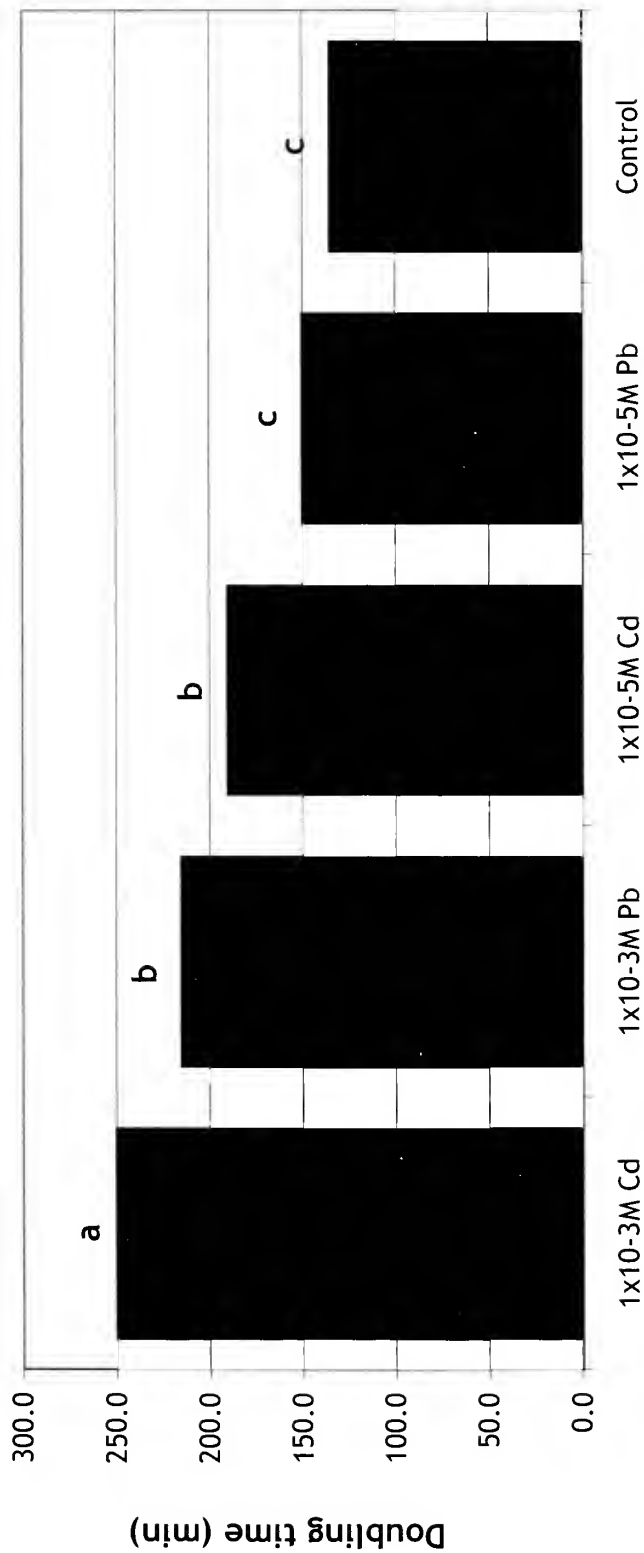
Variable 1: Bacteria and Media

M1= Control medium M9 media

M2= Control rich LB media

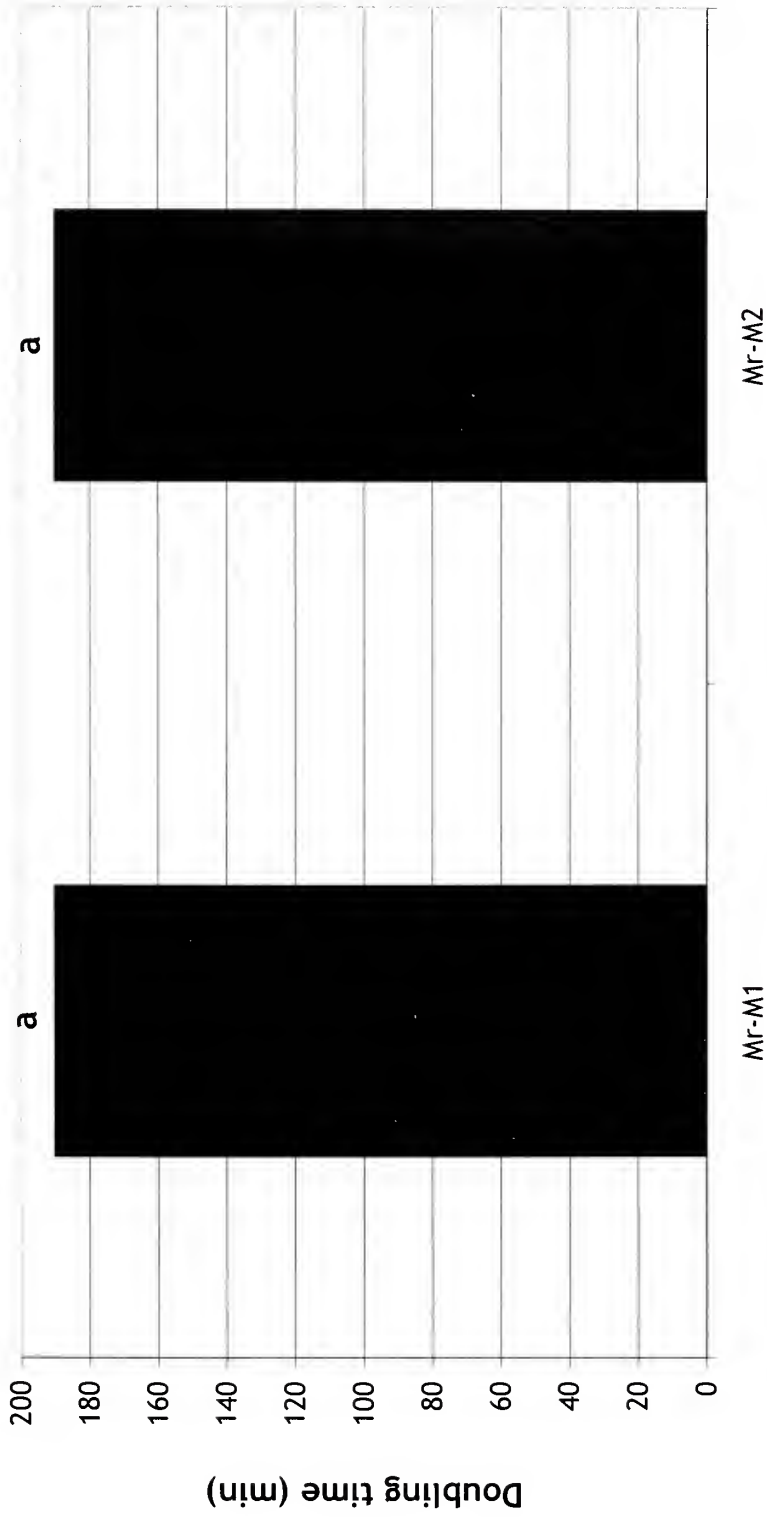
Variable 2: Treatment

Figure 3-1. Average doubling times of *M. roseus* vs. Cd and Pb from absorbance vs. time



M. roseus exposed to 1x10⁻³M and 1x10⁻⁵M Cd and Pb

Figure 3-2. Average doubling times of *M. roseus* vs. different media from absorbance vs. time



M. roseus exposed to different media

Figure 3-3. Summary of average doubling times of *M. roseus* from absorbance vs. time

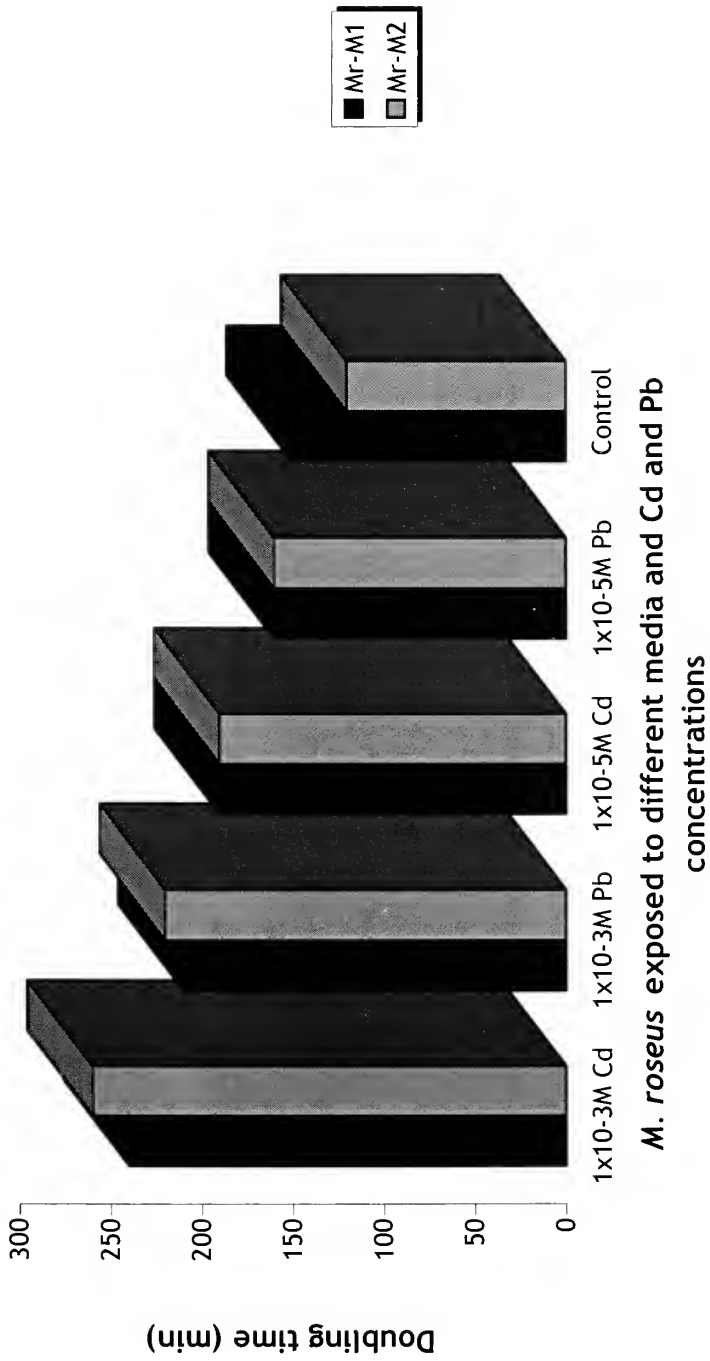


Table 4-9. Doubling times for *E. coli* absorbance vs. time in M9 with and without casamino acids and LB media exposed to $1 \times 10^{-3} \text{M}$ and $1 \times 10^{-5} \text{M}$ Cd and Pb

	Control	$1 \times 10^{-5} \text{M}$ Cd	$1 \times 10^{-3} \text{M}$ Cd	$1 \times 10^{-5} \text{M}$ Pb	$1 \times 10^{-3} \text{M}$ Pb	AVG
M1	120	180	210	150	210	180
	150	180	210	150	210	
	150	180	240	150	210	
M2	60	90	150	60	90	90
	60	90	150	60	90	
	60	90	150	60	90	
M3	30	60	90	30	60	56
	30	60	90	30	60	
	30	60	90	30	90	
AVG	76.7	110	153.3	80.0	123.3	

Variable = *E. coli* (only *E. coli* since *M. roseus* didn't grow in low medium)

M1= Control low media (M9 with no casamino acids)

M2= Control medium M9 media

M3= Control rich LB media

Table 5-1. Two-Factor ANOVA With Replication for *E. coli* absorbance vs. time in M9 with and without casamino acids and LB media exposed to $1 \times 10^{-3} \text{M}$ and $1 \times 10^{-5} \text{M}$ Cd and Pb

Anova: Two-Factor With Replication

SUMMARY	Control	Low Cd	High Cd	Low Pb	High Pb	Total
<i>M1</i>						
Count	3	3	3	3	3	15
Sum	420	540	660	450	630	2700
Average	140	180	220	150	210	180
Variance	300	0	300	0	0	1157.143
<i>M2</i>						
Count	3	3	3	3	3	15
Sum	180	270	450	180	270	1350
Average	60	90	150	60	90	90
Variance	0	0	0	0	0	1157.143
<i>M3</i>						
Count	3	3	3	3	3	15
Sum	90	180	270	90	210	840
Average	30	60	90	30	70	56
Variance	0	0	0	0	300	625.7143
<i>Total</i>						
Count	9	9	9	9	9	9
Sum	690	990	1380	720	1110	
Average	76.6667	110	153.333	80	123.333	
Variance	2500	2925	3250	2925	4375	

ANOVA

Source of Variation	SS	df	MS	F	P-value	F crit
Sample	123160	2	61580	1026.33	2.4E-28	3.315833
Columns	36520	4	9130	152.167	1.8E-19	2.689632
Interaction	2840	8	355	5.91667	0.00014	2.266162
Within	1800	30	60			
Total	164320	44				

Table 5-2. Duncan's Multiple Range Test for *E. coli* absorbance vs. time grown in M9 with and without casamino acids and LB media at 1×10^{-3} M and 1×10^{-5} M Cd and Pb

Treatment					Sw	
df=4	Value of p	2	3	4	5	60 EMS
	SSR	3.9	4.0	4.0	4.0	9 k
	LSR	10.2	10.5	10.5	10.5	6.7 EMS/k
Rp=LSR						2.6 Sx

Media				Sw
df=2	Value of p	2	3	60 EMS
	SSR	6.1	6.1	15 k
	LSR	12.2	12.2	4.0 EMS/k
				2.0 Sx

$$Sx = \frac{EMS}{k}$$

$$LSR = Sx * SSR$$

p=number of means for range being tested
 Protection level = 0.05

Table 5-3. Mean Comparisons and Rp Comparisons for *E. coli* absorbance vs. time grown in M9 with and without casamino acids and LB media at 1×10^{-3} M and 1×10^{-5} M Cd and Pb

High Cd	High Pb	Low Cd	Low Pb	Control
153.3	123.3	110	80	76.7

M1	M2	M3
180	90	54

	Difference	LSR		Difference	LSR	
HC-HP	30.0	10.2	19.8	M1-M2	90	77.8 sig
HC-LC	43.3	10.5	32.8	M1-M3	126	113.8 sig
HC-LP	73.3	10.5	62.8			
HC-Co	76.6	10.5	66.1	M2-M3	36	23.8 sig
HP-LC	13.3	10.2	3.1			
HP-LP	43.3	10.5	32.8			
HP-Co	46.6	10.5	36.1			
LC-LP	30.0	10.2	19.8			
LC-Co	33.3	10.5	22.8			
LP-Co	3.3	10.2	-6.9			not sig

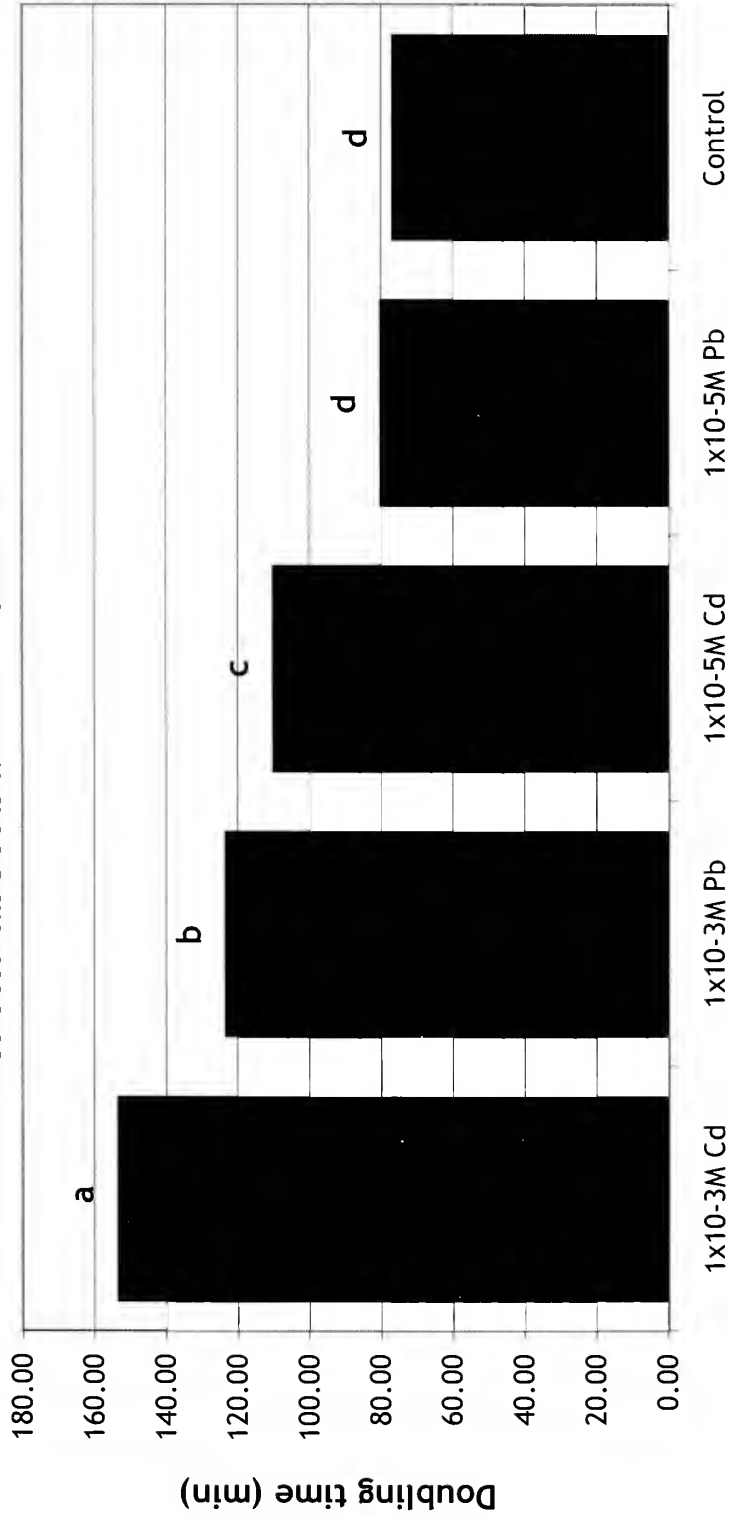
Table 5-4. Summary of doubling times for *E. coli* absorbance vs. time in M9 with and without casamino acids and LB media exposed to 1×10^{-3} M and 1×10^{-5} M Cd and Pb

1×10^{-3} M Cd	1×10^{-3} M Pb	1×10^{-5} M Cd	1×10^{-5} M Pb	Control
153.30	123.30	110.00	80.00	76.67
M1	180			
M2	90			
M3	56			
1×10^{-3} M Cd	1×10^{-3} M Pb	1×10^{-5} M Cd	1×10^{-5} M Pb	Control
M1	220	180	150	140
M2	150	90	60	60
M3	90	60	30	30

Variable = *E. coli* (only *E. coli* since *M. roseus* didn't grow in low medium)

- M1= Control low media (M9 with no casamino acids)
- M2= Control medium M9 media
- M3= Control rich LB media

Figure 3-4. Average doubling times of *E. coli* vs. from absorbance vs. time



E. coli exposed to 1x10⁻³M and 1x10⁻⁵M Cd and Pb

Figures 3-5. Average doubling times of *E. coli* vs. different media from absorbance vs. time

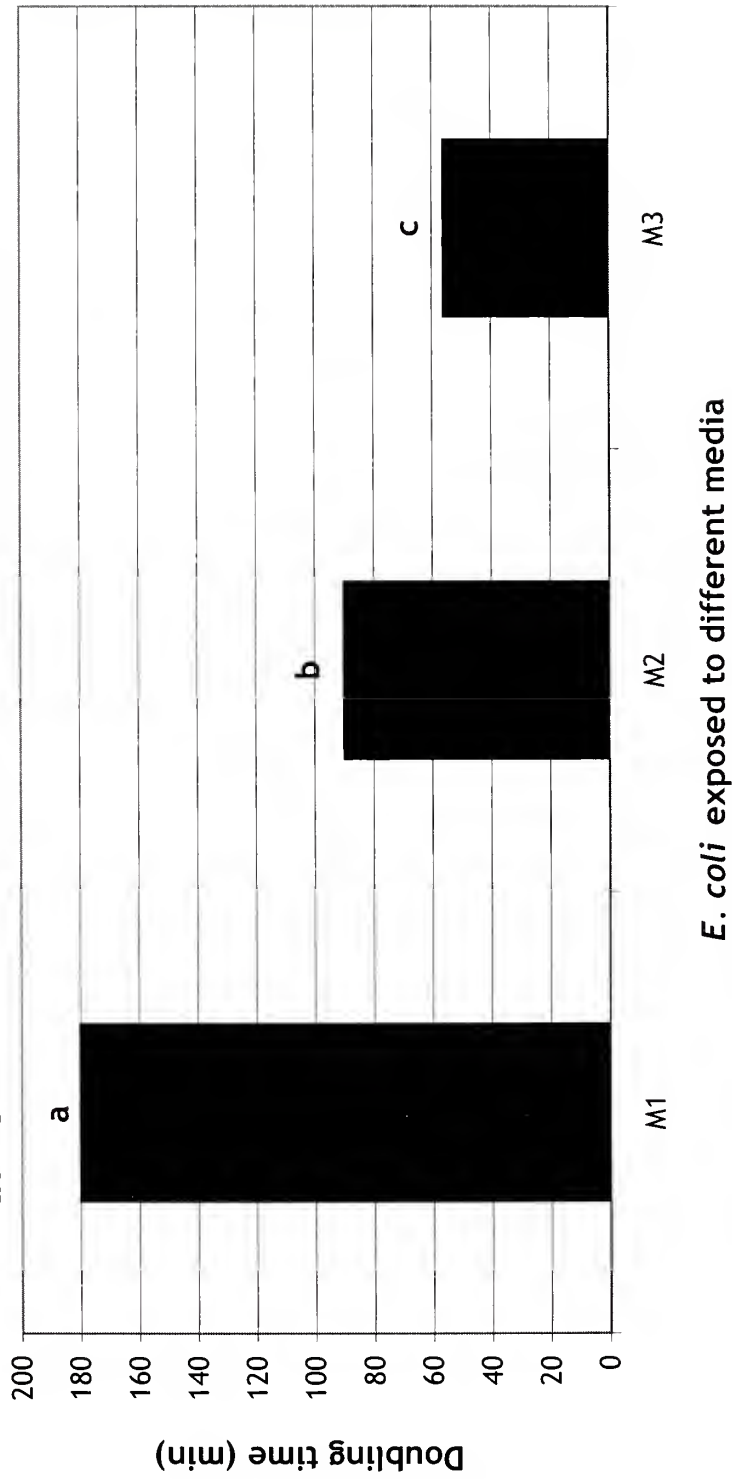


Figure 3-6. Summary of average doubling times of *E. coli* from absorbance vs. time

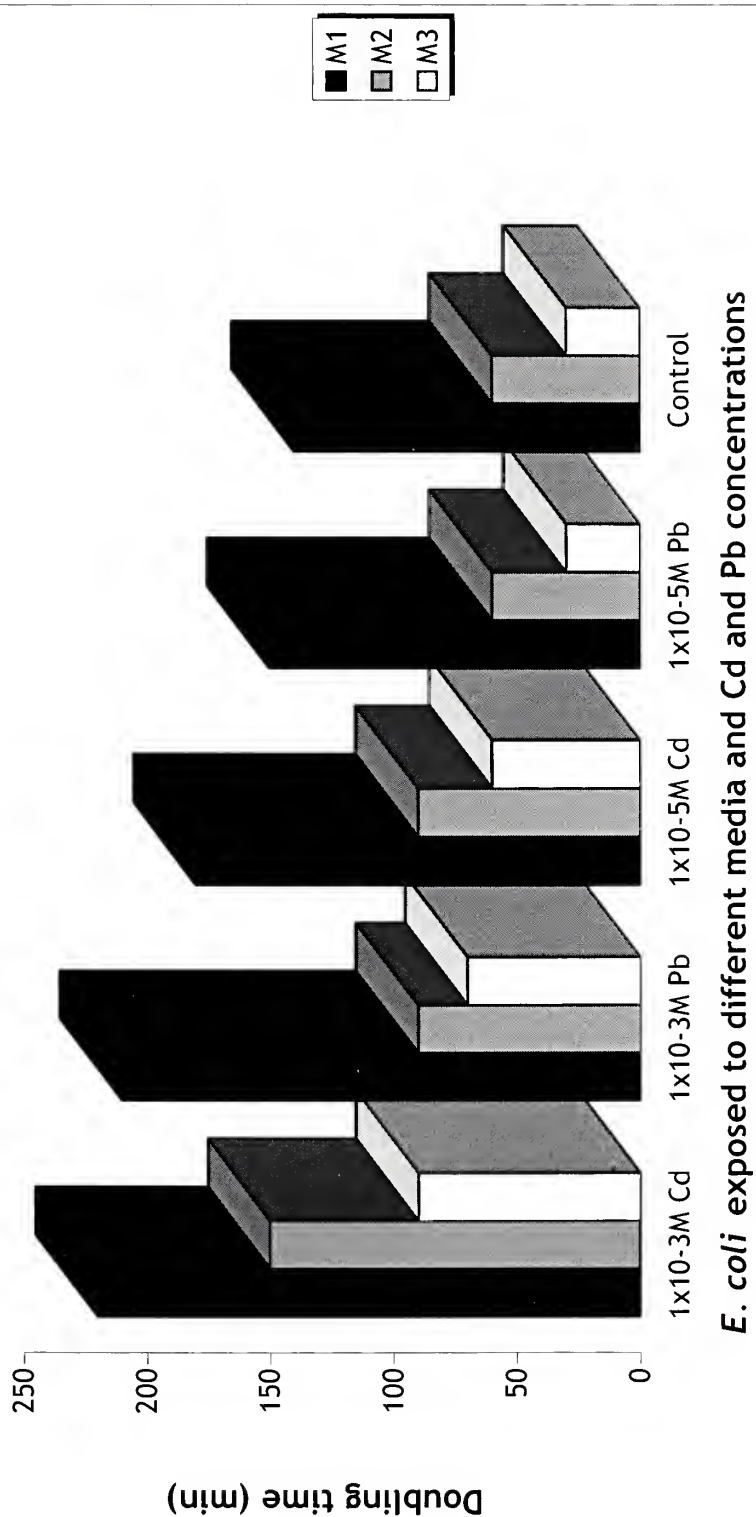


Table 5-5. Exposure of Rf^r wild type, Tc^r and Rf^r donor strain, and mutant to UV-light

	cell number before the UV-light exposure	cell number after the UV-light exposure
wild type	352000000	264000000
	289000000	251000000
	334000000	248000000
Tc ^r donor	2260000	296000
	1880000	283000
	2160000	264000
Rf ^r donor	1960000	128000
	1790000	116000
	2190000	123000
mutant	1730000	85000
	1890000	71000
	1840000	69000
AVG	82725000	63702917

Table 5-6. % Survival of wild type, Tc^r donor, Rf^r donor and mutant strain

	wild type	Tc ^r donor	Rf ^r donor	mutant
% Survival Run 1	75.0	13.1	6.5	4.9
% Survival Run 1	86.9	15.1	6.5	3.8
% Survival Run 1	74.3	12.2	5.6	3.8
AVG % Survival	78.73	13.47	6.20	4.17

Table 5-7. Single Factor Anova for % survival of wild type, Tc^r donor, Rf^r donor and mutant strain

SUMMARY

Groups	Count	Sum	Average	Variance
Wild	3	236.2	78.733	50.143
Tc	3	40.4	13.467	2.203
Rf	3	18.6	6.200	0.270
mutant	3	12.5	4.167	0.403

ANOVA

Source of Variation	SS	df	MS	F	P-value	F crit
Between Groups	11418.3292	3	3806.109722	287.145207	1.76E-08	4.06618
Within Groups	106.04	8	13.255			
Total	11524.3692	11				

Table 5-8. Analysis of Variance for % survival of wild type, Tc^r donor, Rf^r donor and mutant strain

$$S = \sqrt{\text{Variance}}$$

t=4.3

	s	se	ci	ll	ul
wild type	7.081	4.088	17.580	61.154	96.313 a
Tc ^r donor	1.484	0.857	3.685	9.782	17.152 b
Rf ^r donor	0.520	0.300	1.290	4.910	7.490 c
mutant	0.635	0.367	1.577	2.590	5.743 c

$$SE = S / \sqrt{\text{count}}$$

$$CI = t * SE$$

Figure 3-7. Average % survival for wild Type, Tc^r donor, Rf^r donor, and mutant

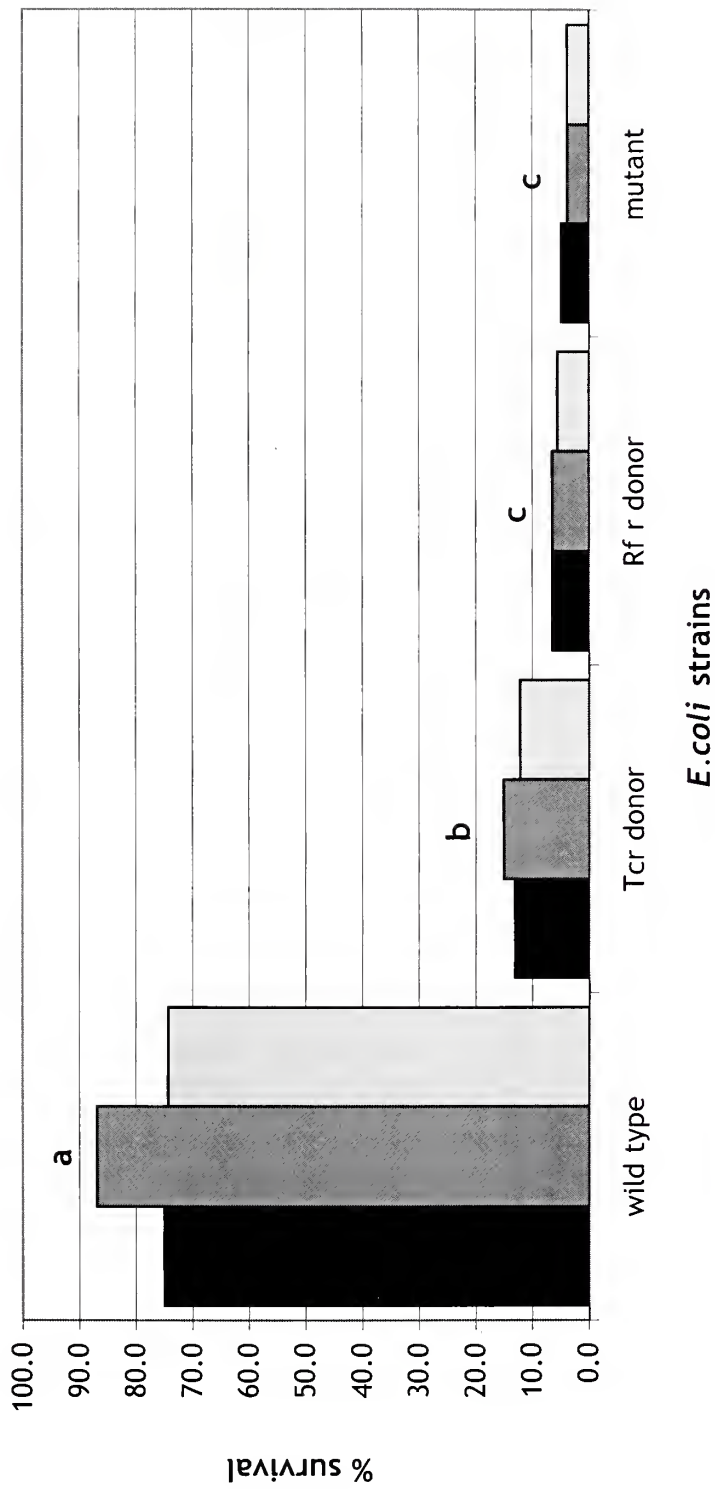


Table 5-9. Duncan's Multiple Range Test for % survival of wild type, Tc^r donor, Rf^r donor, and mutant strain

Media				SW	EMS
df=3	Value of p	2	3	4	12 k
	SSR	4.5	4.5	4.5	1.1 EMS/k
	LSR	4.7	4.7	4.7	1.1 Sx

$$Sx = \sqrt{\frac{EMS}{k}}$$

p=number of means for range being tested
 Protection level = 0.05

$$LSR = Sx * SSR$$

Table 6-1. Mean Comparisons and Rp Comparisons for % survival of wild type, Tc^r donor, Rf^r donor, and mutant strain

wild type	Tc ^r donor	Rf ^r donor	mutant
78.2	13.4	6.2	4.1

	Difference	LSR	
W-Tc	64.8	4.7	60.1 sig
W-Rf	72.0	4.7	67.3 sig
W-Mutant	74.1	4.7	69.4 sig
Tc-Rf	7.2	4.7	2.5 sig
Tc-Mutant	9.3	4.7	4.6 sig
Rf-Mutant	2.1	4.7	-2.6 not sig

DISCUSSION AND CONCLUSION

Injury to *E. coli* strains may result from environmental stress factors such as the presence of sublethal doses of heavy metals (Singh et al., 1986). Heavy metals have been introduced into the environment from industrial processes and during the mining and refining of metal ores. Some metals are essential for microbes, but at the certain level they are toxic. The exposed cells would not be necessarily killed, but physiologically damaged with the lower ability to adapt to an adverse environment (Cenci et al., 1985). The culmination of this study noted that Cd and Pb had an effect on *E. coli* and *M. roseus* doubling times. An *E. coli* strain was also identified that was more resistant to Cd.

Toxic metals Cd⁺² and Pb⁺² have demonstrated a significant effect on *E. coli* and *M. roseus* growth rates. Cd⁺², at 10 μ M and 1mM concentrations as well as Pb⁺² at 1mM concentration, had a significant effect on *E. coli* in all media types. Results suggest that *E. coli* did not show any significant difference between control and growth rates exposed to 10 μ M Pb. However, 10 μ M and 1mM concentrations of both metals had a major effect on growth rates in *M. roseus* medium and rich media. Insignificant effect was found in *M. roseus* growth rates when exposed to 10 μ M Cd and 1mM Pb, as well as 10 μ M Pb and the control. Also, there was no significant difference discovered between *M. roseus* medium and rich media. The responses of *E. coli* and *M. roseus* to cadmium salts were, in general, increased in comparison to lead salts, suggesting slightly different biological effect of these heavy metals. The key to

understanding distinctions may lie in the analysis of different gene induction by the two metals if they are significantly different.

Results by Mariscal et al. showed similar effects of Cd and Pb salts on *E. coli* growth rate (Mariscal et al., 1995). Cenci et al. (1985) previously discussed the complexity of metal-bacteria interaction. Their results emphasize that the binding of heavy metals to the bacterial envelope may play an important role in removing metals from environments polluted with heavy metals (Cenci et al., 1985). Higher concentrations of metals are toxic because of their ability to cause breakage in DNA (Shapiro and Keasling, 1996). Since metals play an important role in bacterial metabolism and the concentration in natural settings limits growth, bacteria have developed numerous high affinity transporters to import essential nutrients. However at high concentrations many metals ions can precipitate or bind inappropriately to macromolecules leading to toxic effects (Gaballa and Helmann, 2003).

In a study by Pazirandeh (1996), expression of the *Neurospora crassa* metallothionein gene (NCP) in *E. coli* was reported, and this recombinant *E. coli* (NCP) was able to sequester cadmium from solution rapidly and with high selectivity. It has been previously shown that NCP was capable of sequestering more Cd from solutions than control bacteria, which did not contain the metallothionein gene. The results in this study indicate that the NCP has properties that are desirable and necessary for a heavy metal biosorbent. These properties include affinity for a wide range of heavy metals, reusability

and durability, it's potential to be used as non-viable biomass, and its ability to be immobilized into a matrix (Pazirandeh, 1996).

Microbes can be used to assay the toxicity and mutagenicity of heavy metals. In this study *E. coli* strains and cadmium were used to some extent to show effects in mutagenesis. The isolation of mutants resistant to specific toxic agents such as Tc and Rif antibiotics was not a very useful genetic approach in this study. It was unfortunate that it cannot be proven that real mutants were created. Instead, the donor was already cadmium sensitive, and the Rif^r donor appeared to be even more sensitive to cadmium and therefore behaved like a mutant. UV-light exposure to wild type (Rif^r recipient), Rif^r donor, Tc^r donor, and suspected mutant, and % survival among above mentioned bacteria confirmed exactly the same behavior of Rif resistant donor and suspected mutant. Growth rates of Rif resistant donor and mutant also supported our theory. Minimal inhibitory Cd concentration of 300µM was established by growing suspected mutants in LB. Cadmium sensitivity of 300µM was then tested on Tc and Rif resistant donor strain and compared with suspected mutant. Absence of growth was observed on all three strains. Therefore it seemed like our donor strain was already a "mutant" that has very high cadmium sensitivity. Further mating of the donor and recipient will be required to identify true mutant. PCR screening of these cadmium sensitive donors will be necessary to assure that the suspected mutants are valid.

In order to identify *E. coli* genes involved with cadmium resistance, wild type *E. coli* was exposed to different concentrations of Cd. Suspected mutant

grew in LB with and without 4 mM of Cd and *E. coli* grew only without Cd in LB. At this time it is not known if these *E. coli* cadmium resistant strains are real mutants. They may increase the expression of some protein that allows them to grow at higher concentrations of Cd. Bacteria carrying cadmium resistance mechanisms have a selective advantage for survival in the environment. Unlike mercury and arsenic resistance systems, which are found in all bacteria studied, cadmium resistance seems to have evolved at least three times, having given rise to the ATP-dependant cadmium efflux transporters present in gram positive bacteria, the unrelated chemi-osmotic cation-proton antiporters found in gram negative bacteria, and the metallothionein system used by cyanobacteria (Schirawski et al., 2002).

Bacterial accommodation to moderate concentrations of cadmium was accompanied by transient activation of general stress proteins (Puskarova et al., 2002). Previously identified protein was identified as the product of the *E. coli yodA* ORF. Three of the four crystal forms were obtained in the presence of zinc, nickel and cadmium, suggesting that YodA may be a metal-binding protein. The increased synthesis of YodA protein during cadmium stress was found probably to be a result of transcriptional activation from one single promoter upstream of the structural *yodA* gene. Analysis of a transcriptional gene fusion, P_{yodA} -*lacZ*, demonstrated that basal expression of *yodA* is low during exponential growth, and increased 50-fold by addition of cadmium to growing cells (Puskarova et al., 2002). However, challenging cells with additional metals such as zinc, copper, cobalt and nickel did not increase the

level of *yodA* expression. YodA protein was primarily detected in the cytoplasmic fraction after 45 minutes of cadmium exposure. After 150 minutes YodA protein was found in both the cytoplasmic and periplasmic compartments (Puskarova et al., 2002). The function of this protein has been identified only under conditions of bacterial stress (David et al., 2002).

P-type adenosine-triphosphatase (ATPases) is polyprotic membrane protein that catalyzes the ATP-dependent transport of cations across membranes. ATPases transporting cations has been identified in many recent studies. P-type ATPases are large and are in easily polarized form. Their physiological roles are to maintain homeostasis of the essential soft metals such as Cu (I) and to mediate resistance to toxic concentrations of Pb (II), Cd (II), Cu (I), and Ag (I) (Sharma et al., 1999). Putative soft metal P-type ATPases have been found in bacteria and can be further subdivided into two classes. The class of soft metal ATPases that are of interest to us have been shown to transport the divalent soft metals, Zn (II) including ZntA from *E. coli*, and Cd (II) including CadA from *Staphylococcus aureus*. ATPase activity was displayed by purified ZntA and specifically stimulated by Pb (II) first, then Cd (II), Zn (II), and Hg (II). This suggests that the physiological role of ZntA is to mediate resistance of these toxic metals (Sharma et al., 1999). The metal specificity study by Okkeri and Haltia shows the substrate ions Zn^{+2} and Cd^{+2} differ in their relative capability to stimulate the ATPase activity and phosphorylation. In comparison with Zn^{+2} , Cd^{+2} is more effective in prompting phosphorylation and less effective in stimulating ATP hydrolysis. These results may suggest that the

enzyme has at least two different metal binding sites with slightly different metal specificities. One that activates the phosphoryl transfer from ATP and another to which the ion to be translocated ion binds (Okkeri and Haltia, 1999).

Zn (II) is a structural component of a large number of proteins in all-living systems and plays an important catalytic role in numerous enzymes. When internal concentrations of Zn (II) are high, a Zn (II) transport system ZntA is switched on. ZntA is a cation-translocating ATPase that exports Cd (II), Pb (II) and Zn (II). Purified ZntA showed the metal dependent ATP hydrolysis activity for Pb (II), Cd (II) and Zn (II) (Binet and Poole, 2000). In a study by Binet and Poole (2000) it was demonstrated that substrates of ZntA encoded exporter Cd (II) and Pb (II) are much more effective inducers of the expressions of this system than the previously considered major inducer Zn (II). From the observation in this study that a ZntA mutant exhibits a higher basal level of ZntA expression than does a wild type it can be concluded that ZntA is functional in metal ion export (Binet and Poole, 2000).

Two *Streptococcus thermophilus* genes *cadC* and *cadA* located on the chromosome of *S. thermophilus* 4134 were shown to comprise a cadmium/zinc cassette. The product of *cadA* is highly similar to P-type cadmium efflux ATPases. Electrophoretic mobility shift assays were used to demonstrate that *cadC* is a DNA binding protein that binds specifically to its own promoter region, and that this *cadC*-DNA interaction is lost in the presence of cadmium (Schirawski et al., 2002).

The metal binding sites in ZntA and CadA are yet to be discovered. It can be hypothesized that certain residues confer specificity for certain metals. Identification of the metal binding sites on ZntA and CadA may lead to understanding the basis of metal recognition and perhaps engineering of the protein to modify metal specificity (Sharma et al., 1999).

Future studies involving the identification of *E. coli* genes that are Cd resistant can provide better insight into Cd resistant mechanisms in bacteria. Additional research is required where two different bacteria should be used as donor and recipient instead of the two different strains of the same bacteria. Further potential improvements may be achieved by modification of the present study procedure for discovering mutants. Characterization of genes for wild type and donor may provide insight into the cellular targets, and the mechanisms of sensitivity to metal exposure. Therefore, it is hoped that the results described in this research thesis will be useful for further studies based on the use of this environmentally important scientific approach.

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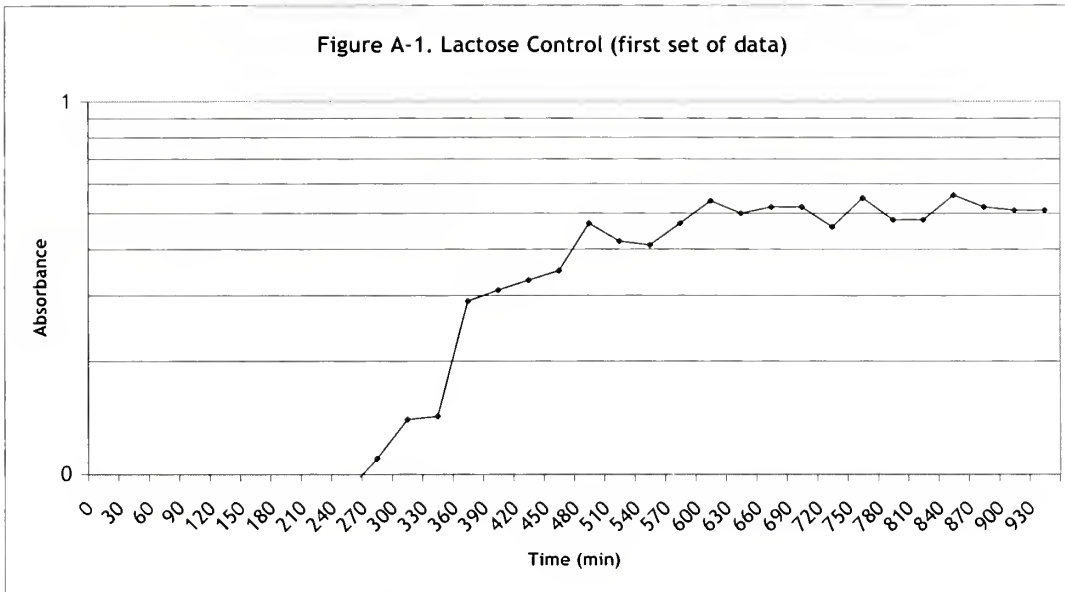
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APPENDIX A

All of the doubling times for *E.coli* are presented in APPENDIX A

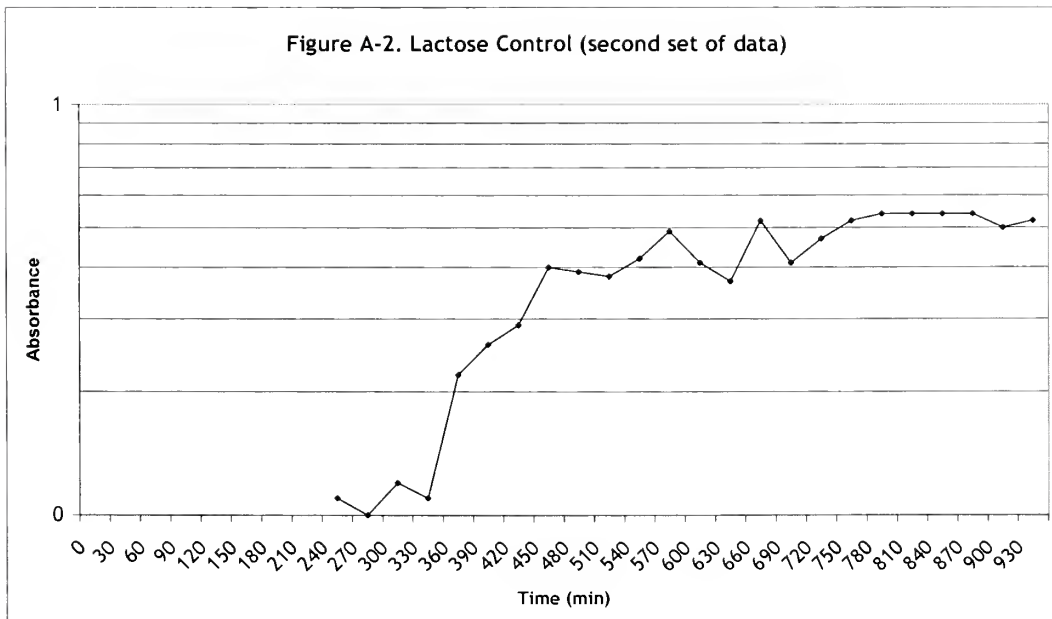
Data for *E.coli* from absorbance vs. time

Lactose control				
Time	Run1	Run2	Run 3	Average
0	0	0	0	0
30	0	0	0	0
60	0	0	0	0
90	0	0	0	0
120	0	0	0	0
150	0	0	0	0
180	0	0	0	0
210	0	0	0	0
240	0.09	0.11	0.10	0.10
270	0.11	0.10	0.12	0.11
300	0.14	0.12	0.14	0.13
330	0.14	0.11	0.15	0.13
360	0.29	0.22	0.29	0.27
390	0.31	0.26	0.30	0.29
420	0.33	0.29	0.31	0.31
450	0.35	0.40	0.34	0.36
480	0.47	0.39	0.45	0.44
510	0.42	0.38	0.46	0.42
540	0.41	0.42	0.47	0.43
570	0.47	0.49	0.47	0.48
600	0.54	0.41	0.54	0.50
630	0.50	0.37	0.54	0.47
660	0.52	0.52	0.55	0.53
690	0.52	0.41	0.55	0.49
720	0.46	0.47	0.54	0.49
750	0.55	0.52	0.55	0.54
780	0.48	0.54	0.53	0.52
810	0.48	0.54	0.54	0.52
840	0.56	0.54	0.54	0.55
870	0.52	0.54	0.53	0.53
900	0.51	0.50	0.52	0.51
930	0.51	0.52	0.52	0.52



Absorbance increases from 0.15 to 0.29

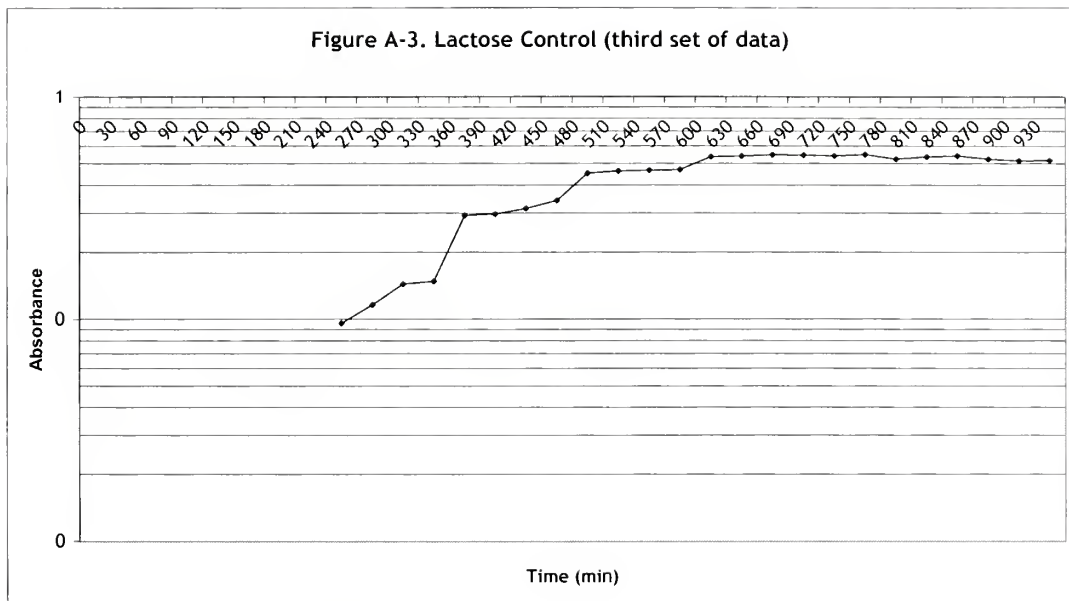
Actual grow rate (360 min - 330 min) = 30 min



Absorbance increases from 0.11 to 0.22

Actual grow rate (360 min - 330 min) = 30 min

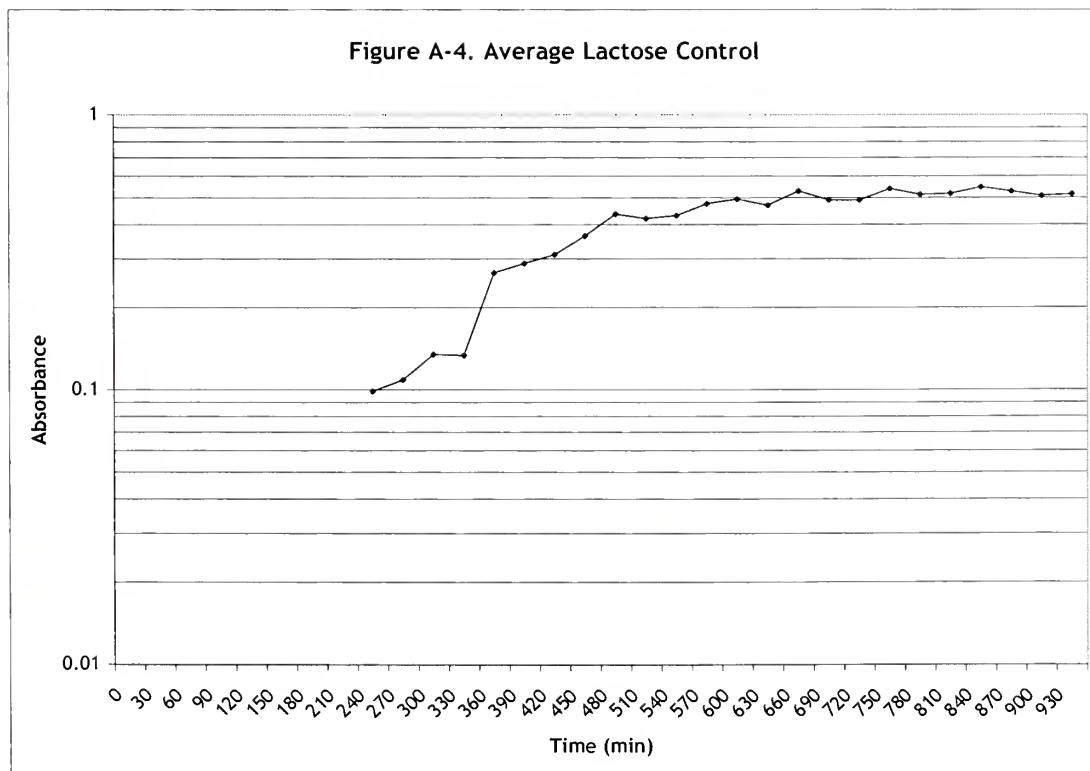
Figure A-3. Lactose Control (third set of data)



Absorbance increases from 0.148 to 0.297

Actual grow rate (390 min - 330 min) = 60 min

Figure A-4. Average Lactose Control

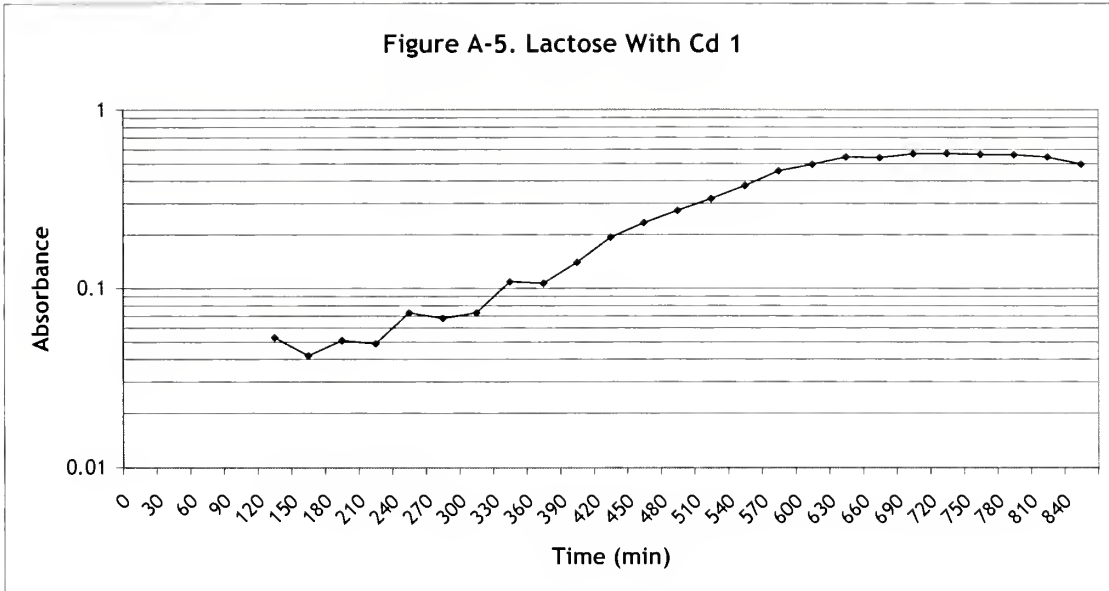


Absorbance increases from 0.134 to 0.268

Actual grow rate (360 min - 330 min) = 30 min

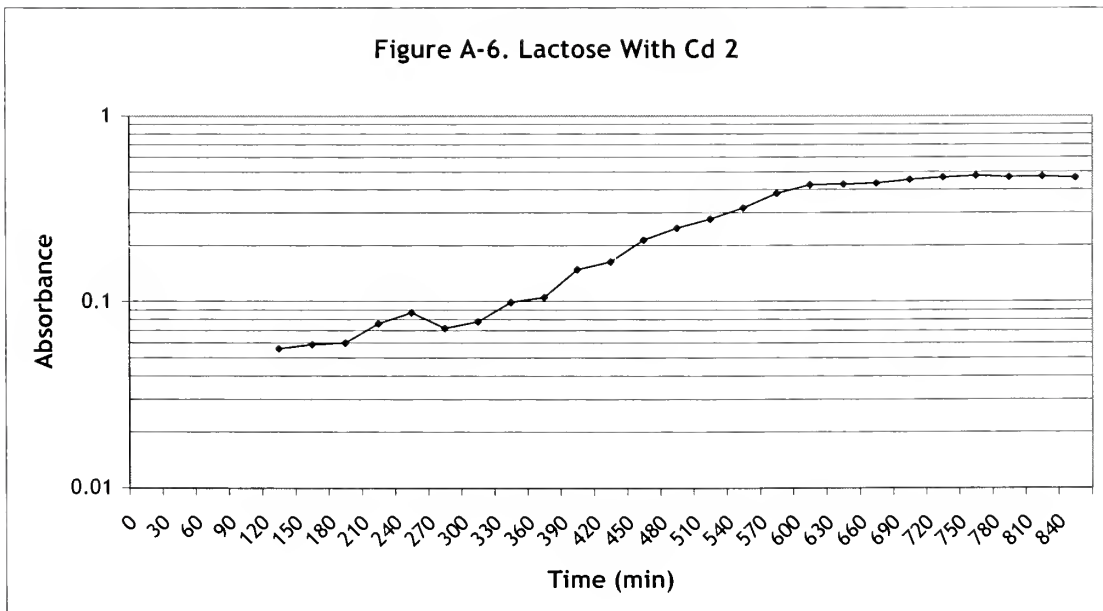
Data for *E.coli* from absorbance vs. time

Lactose only Cadmium at $1 \cdot 10^{-3} \text{ M}$					
Time (min)	Run 1	Run 2	Run 3	Run 4	Average
0	0	0	0	0	0
30	0	0	0	0	0
60	0	0	0	0	0
90	0	0	0	0	0
120	0.05	0.06	0.05	0.05	0.05
150	0.04	0.06	0.06	0.04	0.05
180	0.05	0.06	0.05	0.05	0.05
210	0.05	0.08	0.06	0.07	0.06
240	0.07	0.09	0.08	0.07	0.08
270	0.07	0.07	0.06	0.07	0.07
300	0.07	0.08	0.08	0.08	0.08
330	0.11	0.10	0.07	0.06	0.08
360	0.11	0.11	0.07	0.09	0.09
390	0.14	0.15	0.08	0.06	0.11
420	0.19	0.16	0.07	0.09	0.13
450	0.23	0.21	0.09	0.07	0.15
480	0.27	0.25	0.09	0.11	0.18
510	0.32	0.28	0.13	0.11	0.21
540	0.38	0.32	0.15	0.12	0.24
570	0.46	0.38	0.20	0.16	0.30
600	0.50	0.43	0.23	0.19	0.34
630	0.54	0.43	0.24	0.25	0.37
660	0.54	0.43	0.32	0.27	0.39
690	0.57	0.45	0.37	0.29	0.42
720	0.57	0.47	0.42	0.35	0.45
750	0.56	0.48	0.47	0.38	0.47
780	0.56	0.47	0.50	0.41	0.49
810	0.55	0.47	0.52	0.44	0.49
840	0.50	0.46	0.53	0.46	0.49



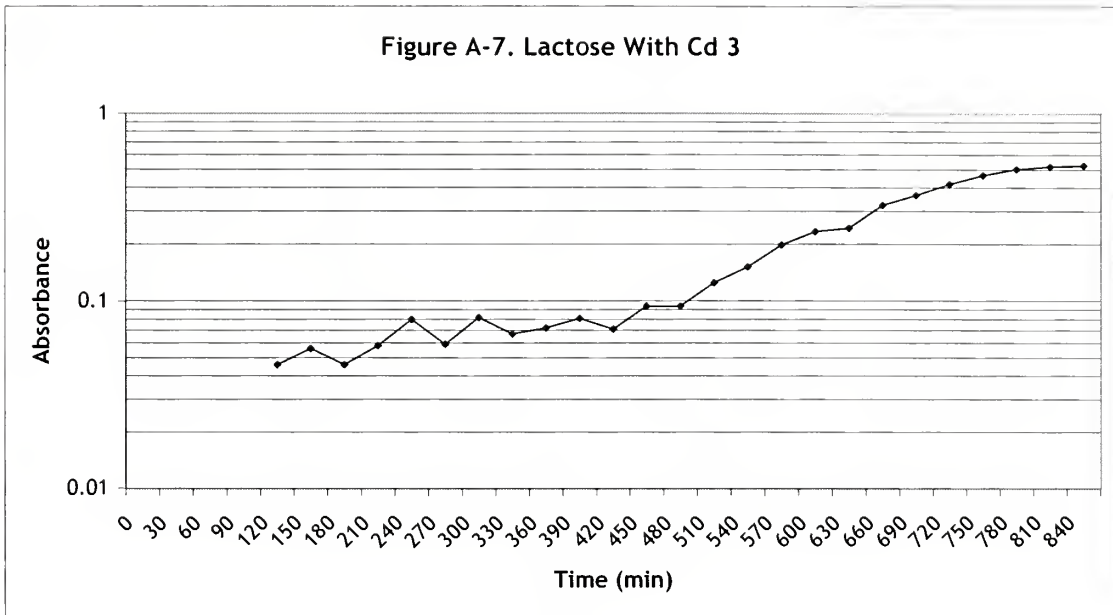
Absorbance increases from 0.273 to 0.544

Actual growth rate = (630 min - 480 min) = 150 min



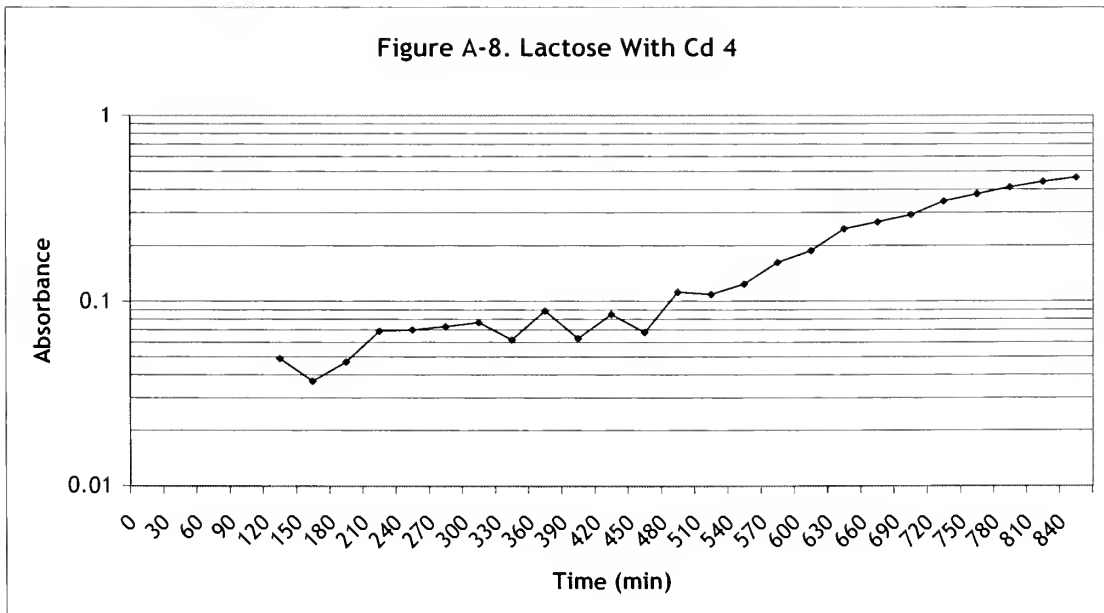
Absorbance increases from 0.214 to 0.426

Actual growth rate = (600 min - 450 min) = 150 min



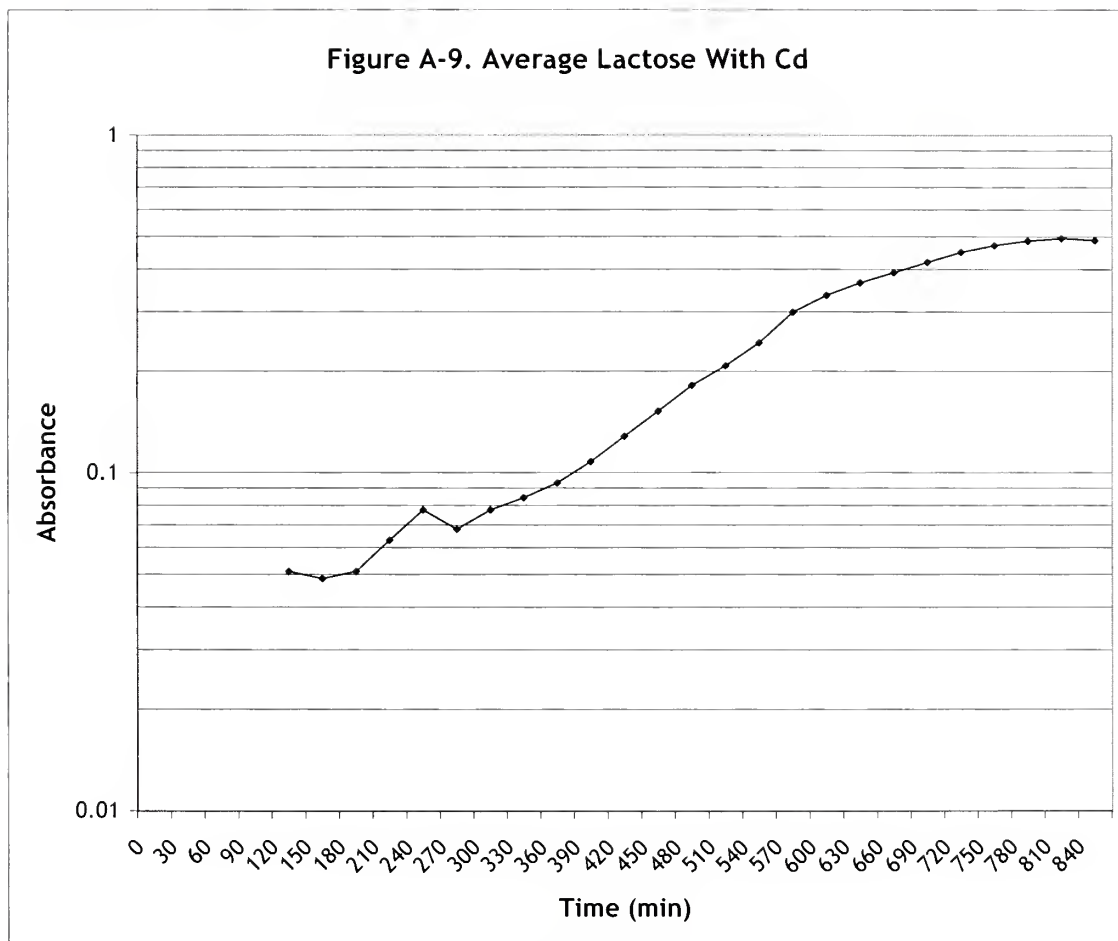
Absorbance increases from 0.234 to 0.467

Actual growth rate = (750 min - 600 min) = 150 min



Absorbance increases from 0.187 to 0.378

Actual growth rate = (750 min - 600 min) = 150 min

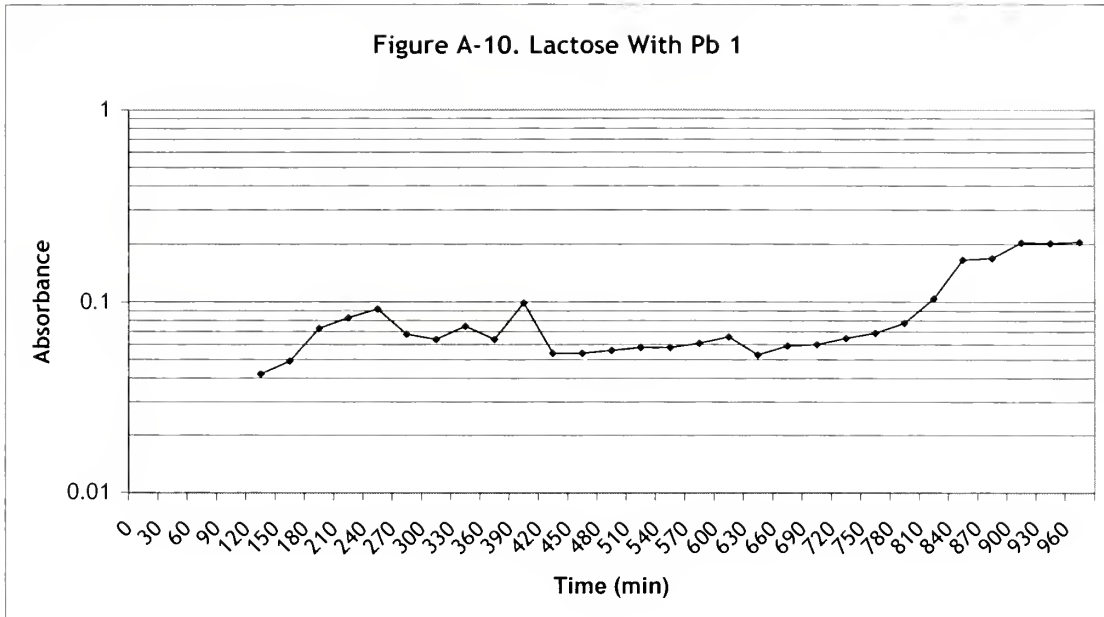


Absorbance increases from 0.182 to 0.366

Actual growth rate = (630 min - 480 min) = 150 min

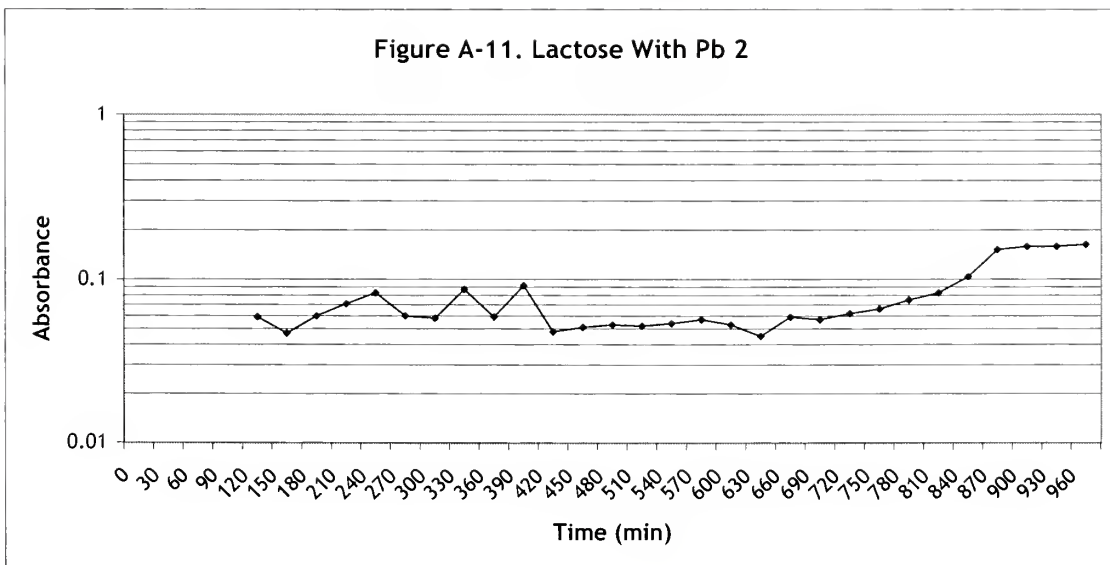
Data for *E.coli* from absorbance vs. time

Lactose only Lead at $1 \cdot 10^{-3} M$				
Time (min)	Run 1	Run 2	Run 3	Average
0	0	0	0	0
30	0	0	0	0
60	0	0	0	0
90	0	0	0	0
120	0.042	0.059	0.062	0.054
150	0.049	0.047	0.041	0.046
180	0.073	0.060	0.054	0.062
210	0.083	0.071	0.076	0.077
240	0.092	0.083	0.088	0.088
270	0.068	0.060	0.072	0.067
300	0.064	0.058	0.055	0.059
330	0.075	0.087	0.070	0.077
360	0.064	0.059	0.065	0.063
390	0.099	0.092	0.095	0.095
420	0.054	0.048	0.040	0.047
450	0.054	0.051	0.054	0.053
480	0.056	0.053	0.057	0.055
510	0.058	0.052	0.058	0.056
540	0.058	0.054	0.055	0.056
570	0.061	0.057	0.060	0.059
600	0.066	0.053	0.060	0.060
630	0.053	0.045	0.051	0.050
660	0.059	0.059	0.066	0.061
690	0.060	0.057	0.065	0.061
720	0.065	0.062	0.076	0.068
750	0.069	0.066	0.087	0.074
780	0.078	0.075	0.097	0.083
810	0.104	0.083	0.121	0.103
840	0.166	0.104	0.171	0.147
870	0.169	0.152	0.188	0.170
900	0.204	0.159	0.198	0.187
930	0.202	0.159	0.203	0.188
960	0.205	0.163	0.205	0.191



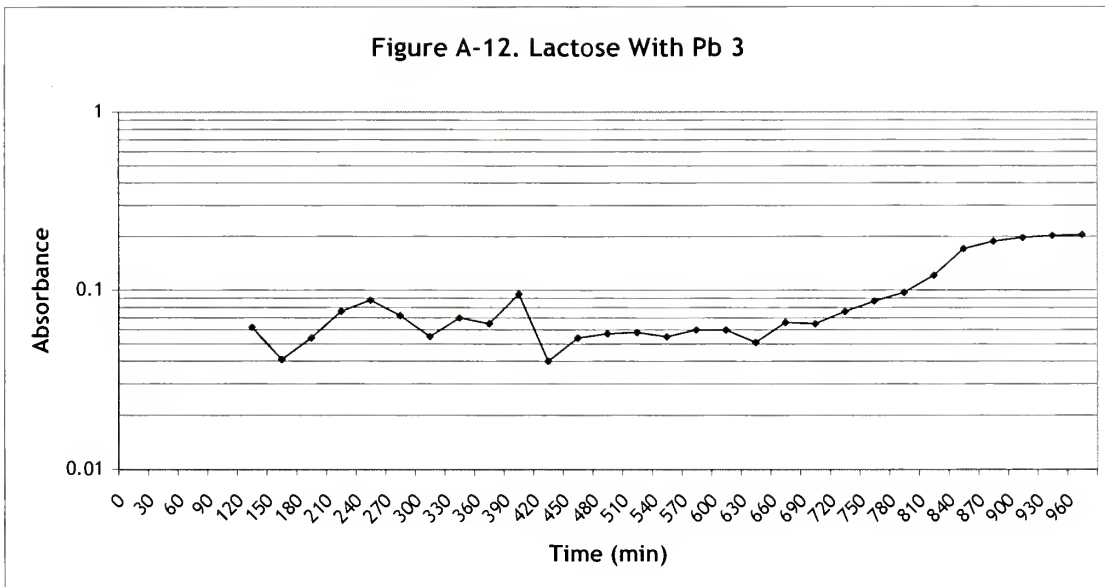
Absorbance increases from 0.104 to 0.204

Actual growth rate = (900 min - 810 min) = 90 min



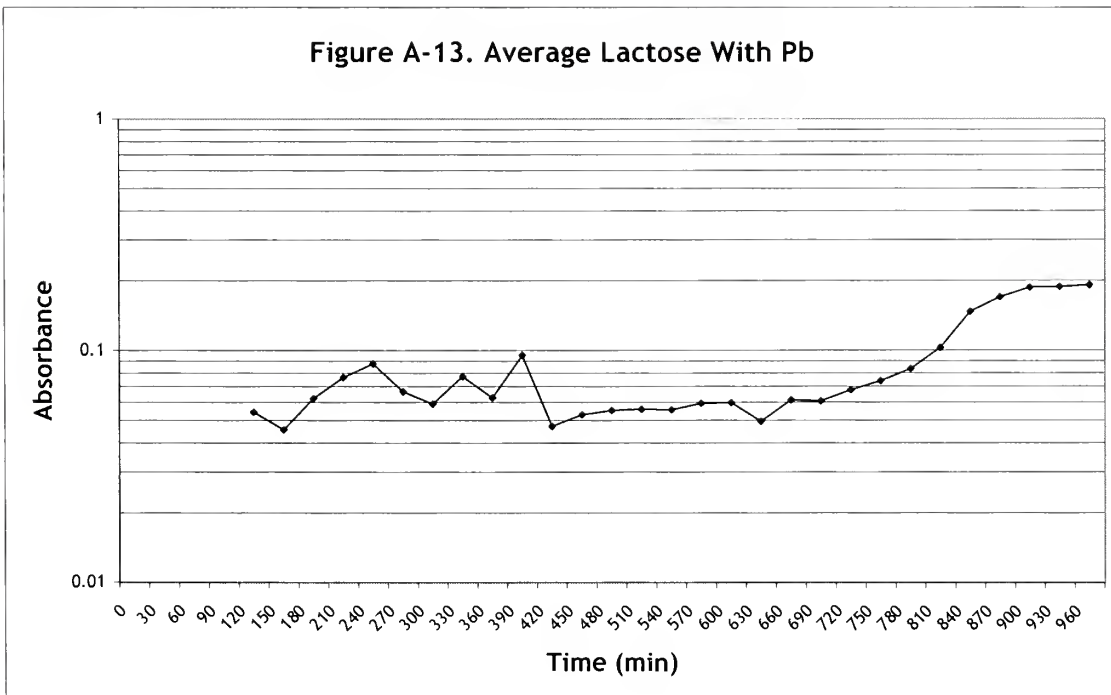
Absorbance increases from 0.075 to 0.152

Actual growth rate = (870 min - 780 min) = 90 min



Absorbance increases from 0.087 to 0.171

Actual growth rate = (840 min - 750 min) = 90 min

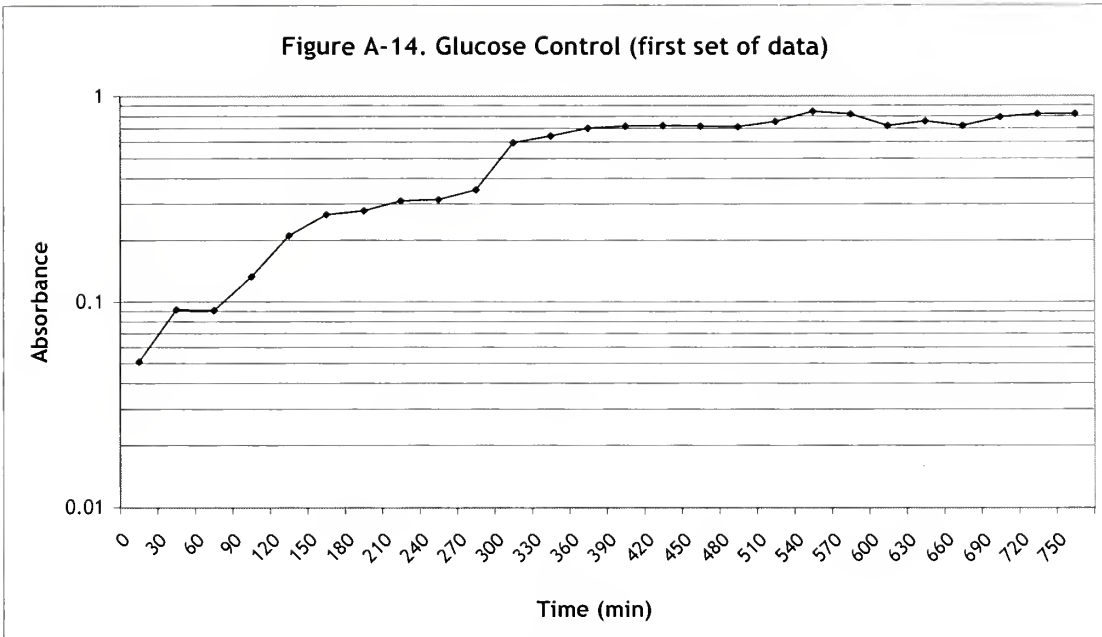


Absorbance increases from 0.074 to 0.147

Actual growth rate = (840 min - 750 min) = 90 min

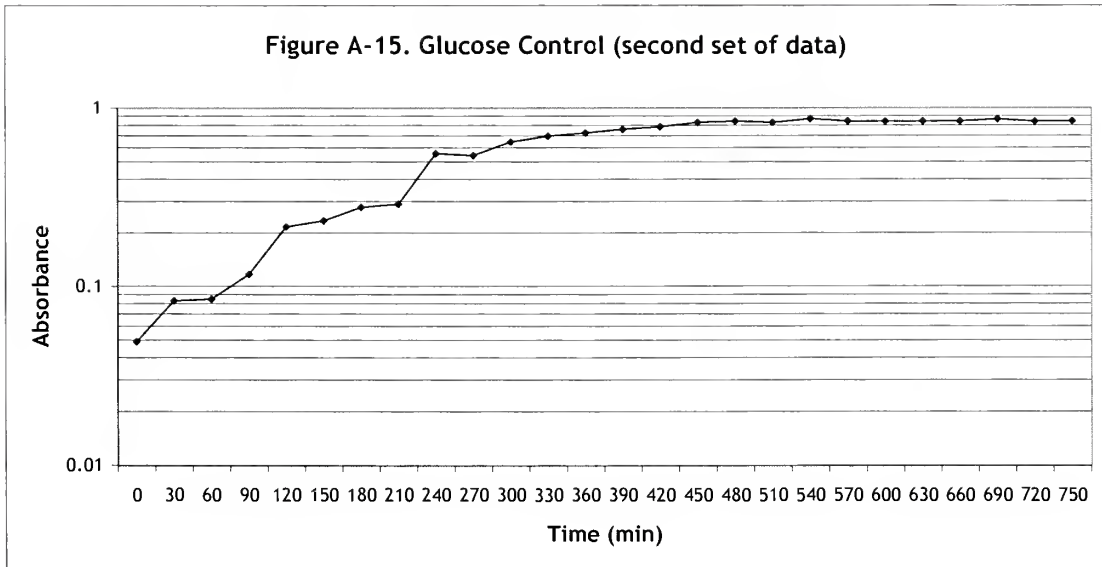
Data for *E.coli* from absorbance vs. time in M9 media

Glucose control				
Time(min)	Run1	Run2	Run3	Average
0	0	0	0	0
0	0.051	0.049	0.051	0.050
30	0.092	0.083	0.089	0.088
60	0.091	0.085	0.09	0.088
90	0.133	0.117	0.129	0.125
120	0.211	0.216	0.213	0.214
150	0.267	0.234	0.259	0.251
180	0.278	0.278	0.272	0.278
210	0.311	0.290	0.308	0.301
240	0.316	0.556	0.411	0.436
270	0.352	0.544	0.482	0.448
300	0.596	0.646	0.546	0.621
330	0.644	0.698	0.612	0.671
360	0.702	0.726	0.698	0.714
390	0.718	0.760	0.712	0.739
420	0.722	0.786	0.736	0.754
450	0.716	0.830	0.758	0.773
480	0.712	0.845	0.763	0.779
510	0.754	0.830	0.782	0.792
540	0.845	0.870	0.843	0.858
570	0.821	0.845	0.841	0.833
600	0.720	0.840	0.836	0.780
630	0.758	0.845	0.841	0.802
660	0.720	0.845	0.838	0.783
690	0.792	0.865	0.835	0.829
720	0.820	0.840	0.838	0.830
750	0.820	0.845	0.837	0.833



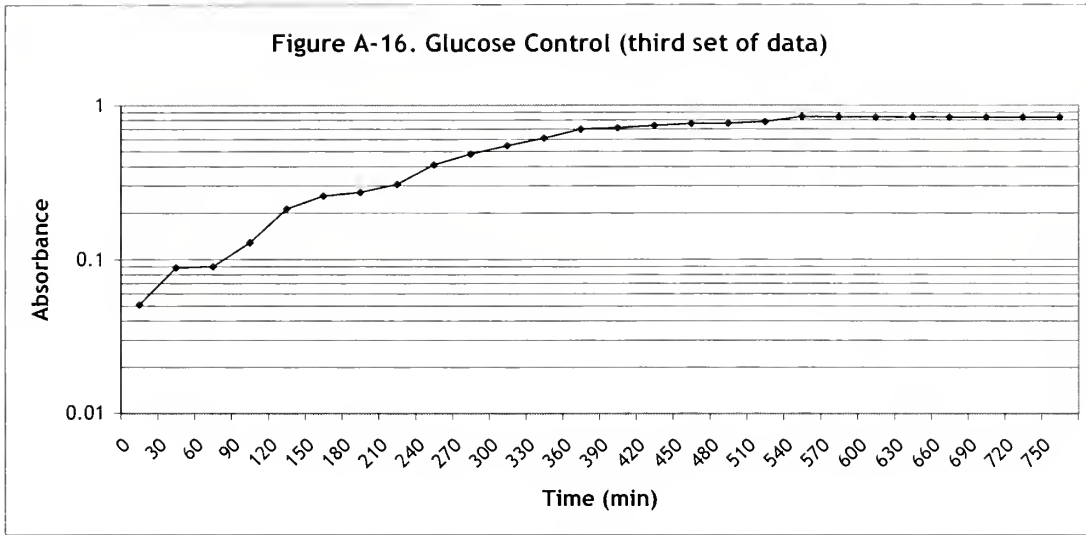
Absorbance increases from 0.133 to 0.267

Actual growth rate = (150 min - 90 min) = 60 min



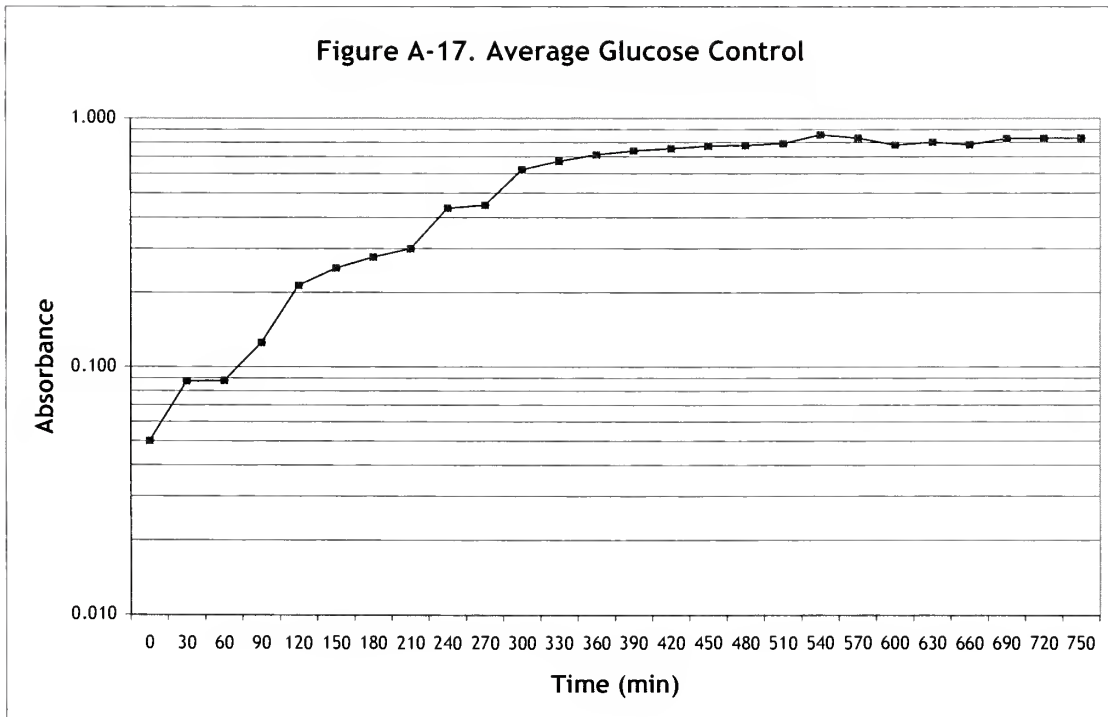
Absorbance increases from 0.278 to 0.556

Actual growth rate = (240 min - 180 min) = 60 min



Absorbance increases from 0.129 to 0.259

Actual growth rate = (150 min - 90 min) = 60 min



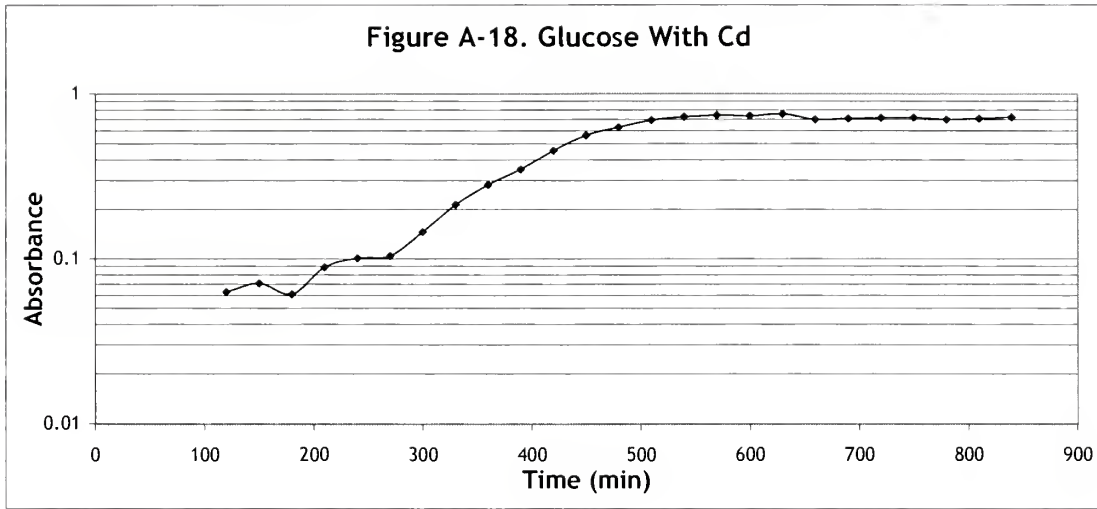
Absorbance increases from 0.125 to .0.251

Actual growth rate = (150 min - 90 min) = 60 min

Data for *E.coli* from absorbance vs. time in M9 media

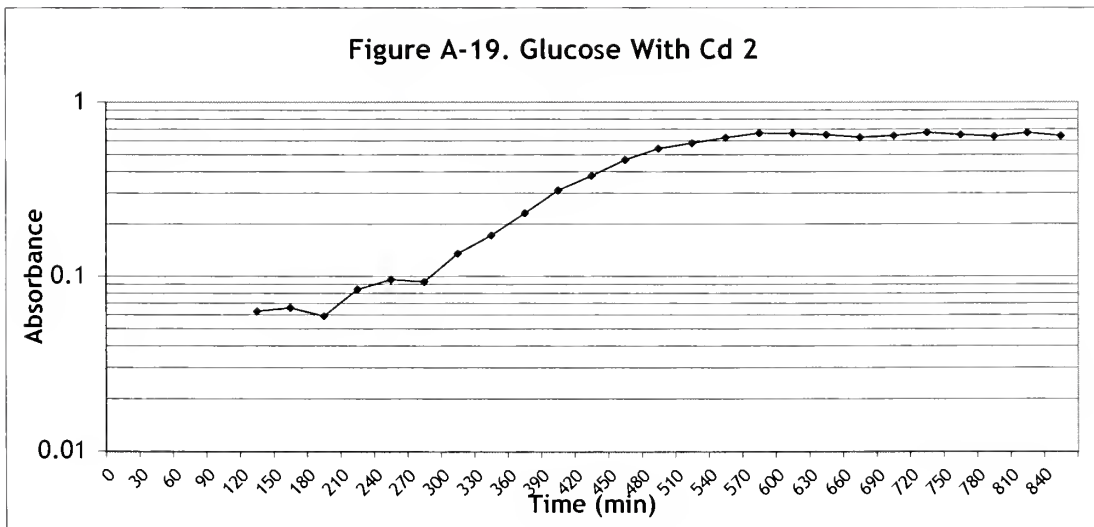
Glucose and Cd at $1 \cdot 10^{-3}M$

Time (min)	Run 1	Run 2	Run 3	Run 4	Average
0	0	0	0	0	0
30	0	0	0	0	0
60	0	0	0	0	0
90	0	0	0	0	0
120	0.063	0.063	0.036	0.075	0.059
150	0.071	0.066	0.079	0.079	0.074
180	0.061	0.059	0.063	0.095	0.070
210	0.089	0.084	0.075	0.083	0.083
240	0.101	0.096	0.086	0.094	0.094
270	0.104	0.093	0.082	0.094	0.093
300	0.146	0.135	0.089	0.120	0.123
330	0.214	0.172	0.136	0.159	0.170
360	0.283	0.231	0.169	0.221	0.226
390	0.351	0.312	0.275	0.291	0.307
420	0.456	0.378	0.280	0.340	0.364
450	0.562	0.468	0.365	0.400	0.449
480	0.628	0.542	0.424	0.474	0.517
510	0.696	0.584	0.466	0.504	0.563
540	0.728	0.628	0.549	0.546	0.613
570	0.742	0.666	0.572	0.580	0.640
600	0.736	0.662	0.600	0.575	0.643
630	0.756	0.650	0.606	0.572	0.646
660	0.702	0.630	0.584	0.562	0.620
690	0.712	0.644	0.576	0.556	0.622
720	0.718	0.672	0.564	0.520	0.619
750	0.720	0.652	0.560	0.560	0.623
780	0.702	0.638	0.554	0.565	0.615
810	0.712	0.668	0.550	0.543	0.618
840	0.722	0.640	0.546	0.546	0.614



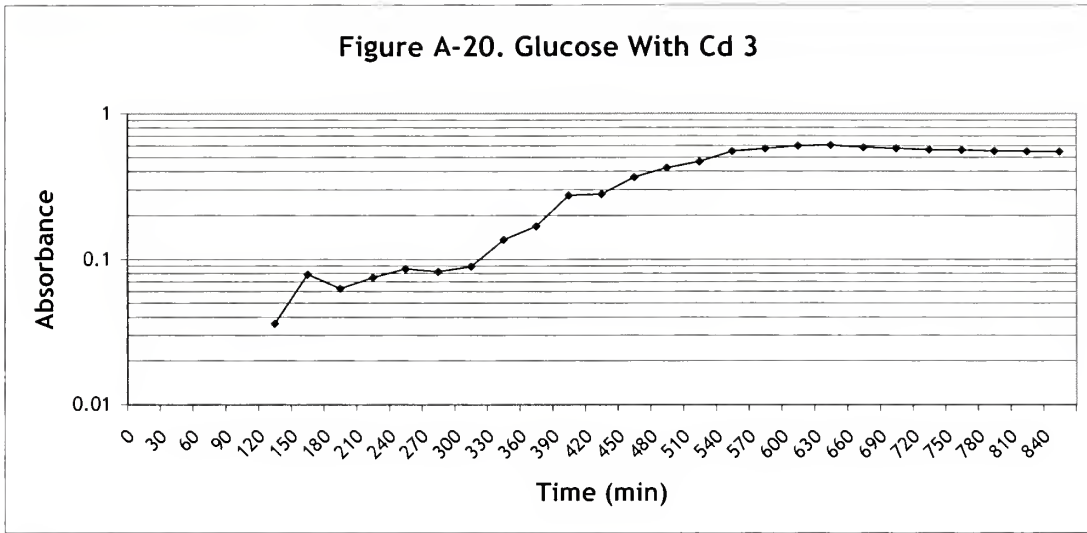
Absorbance increases from 0.365 to 0.728

Actual growth rate = (540 min - 390 min) = 150 min



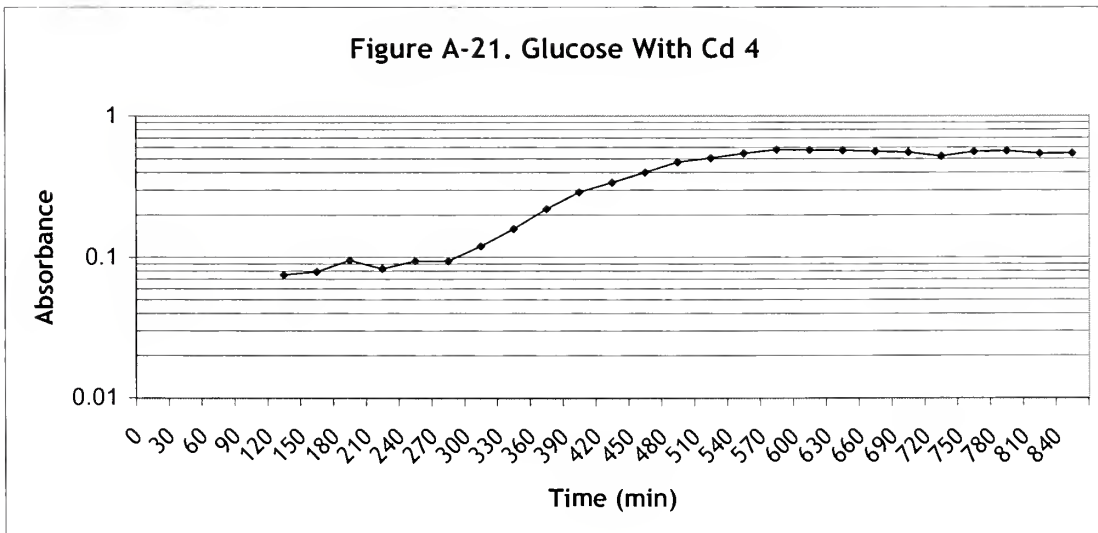
Absorbance increases from 0.312 to 0.628

Actual growth rate = (540 min - 390 min) = 150 min



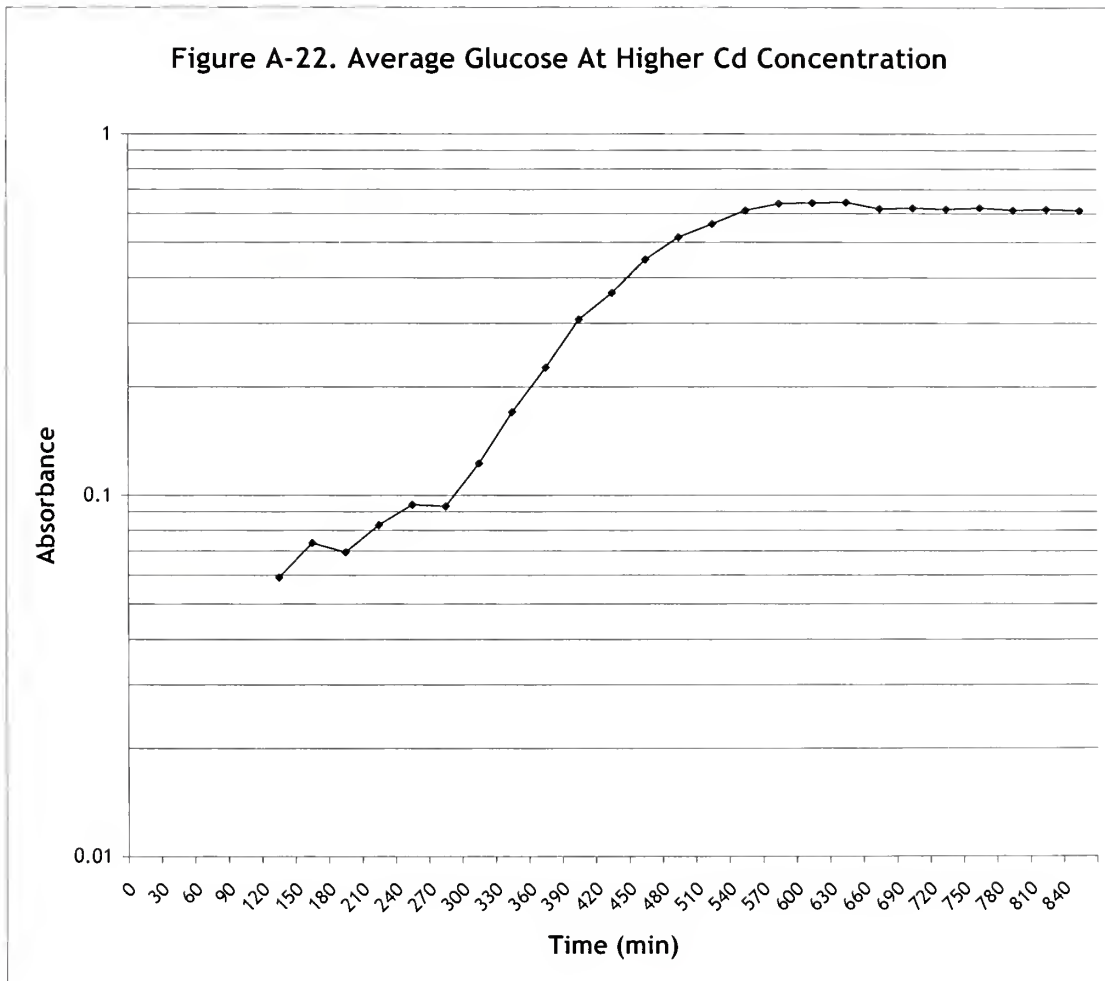
Absorbance increases from 0.275 to 0.549

Actual growth rate = (540 min - 390 min) = 150 min



Absorbance increases from 0.291 to 0.580

Actual growth rate = (540 min - 390 min) = 150 min



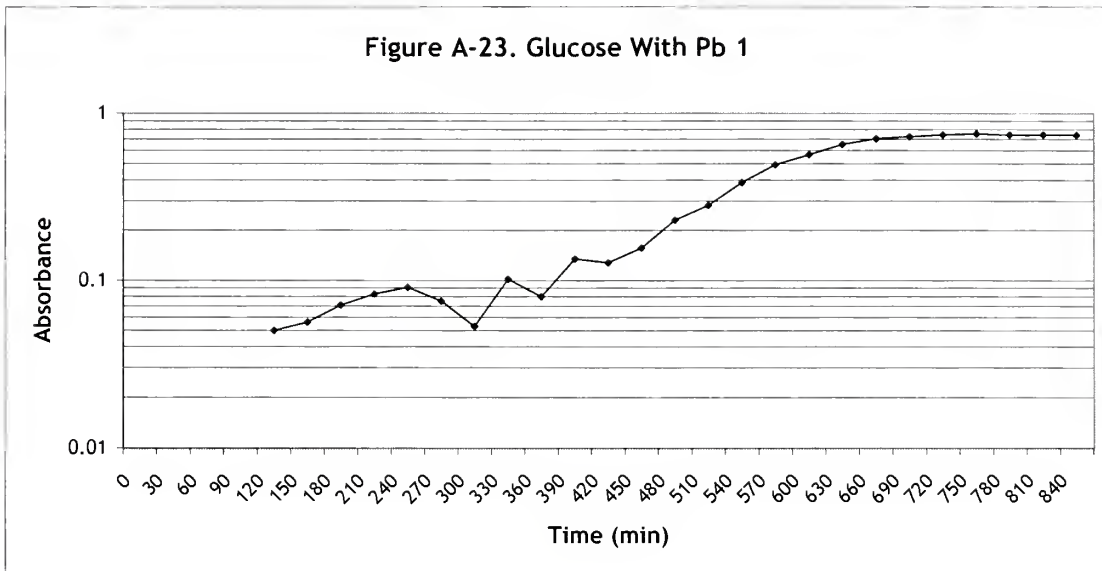
Absorbance increases from 0.307 to 0.613

Actual growth rate = (540 min - 390 min) = 150 min

Data for *E. coli* from absorbance vs. time in M9 media

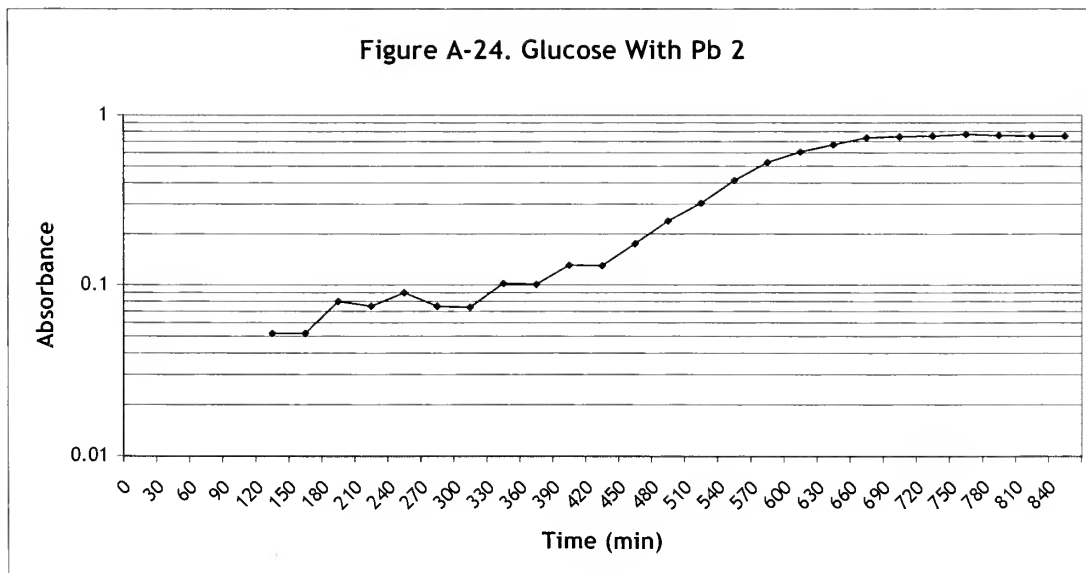
Glucose and Pb at 1×10^{-3} M

Time (min)	Run 1	Run 2	Run 3	Run 4	Average
0	0	0	0	0	0
30	0	0	0	0	0
60	0	0	0	0	0
90	0	0	0	0	0
120	0.050	0.052	0.048	0.069	0.055
150	0.056	0.052	0.050	0.067	0.056
180	0.071	0.080	0.073	0.087	0.078
210	0.083	0.075	0.092	0.093	0.086
240	0.091	0.090	0.099	0.097	0.094
270	0.075	0.075	0.079	0.075	0.076
300	0.053	0.074	0.074	0.076	0.069
330	0.102	0.102	0.091	0.092	0.097
360	0.080	0.101	0.087	0.066	0.084
390	0.135	0.131	0.130	0.143	0.135
420	0.128	0.130	0.109	0.126	0.123
450	0.157	0.175	0.121	0.163	0.154
480	0.231	0.239	0.184	0.215	0.217
510	0.283	0.303	0.272	0.290	0.287
540	0.388	0.414	0.325	0.404	0.383
570	0.494	0.528	0.440	0.516	0.495
600	0.568	0.608	0.538	0.584	0.575
630	0.652	0.670	0.648	0.682	0.663
660	0.708	0.738	0.694	0.726	0.717
690	0.728	0.748	0.722	0.744	0.736
720	0.746	0.754	0.742	0.754	0.749
750	0.756	0.770	0.750	0.756	0.758
780	0.748	0.760	0.758	0.754	0.755
810	0.748	0.754	0.754	0.754	0.753
840	0.744	0.752	0.746	0.744	0.747



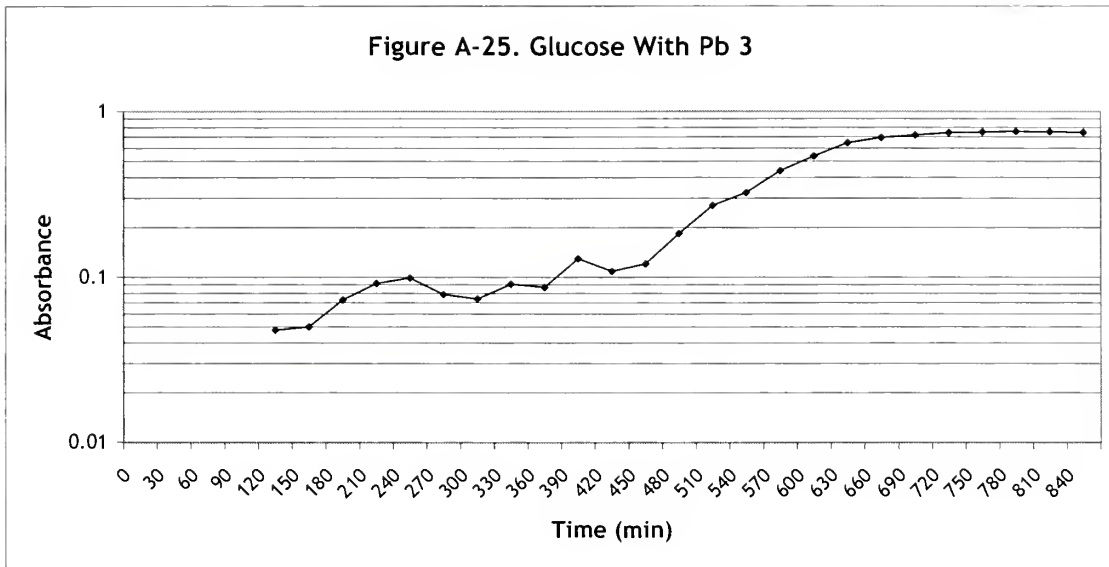
Absorbance increases from 0.283 to 0.568

Actual growth rate = (600 min - 510 min) = 90 min



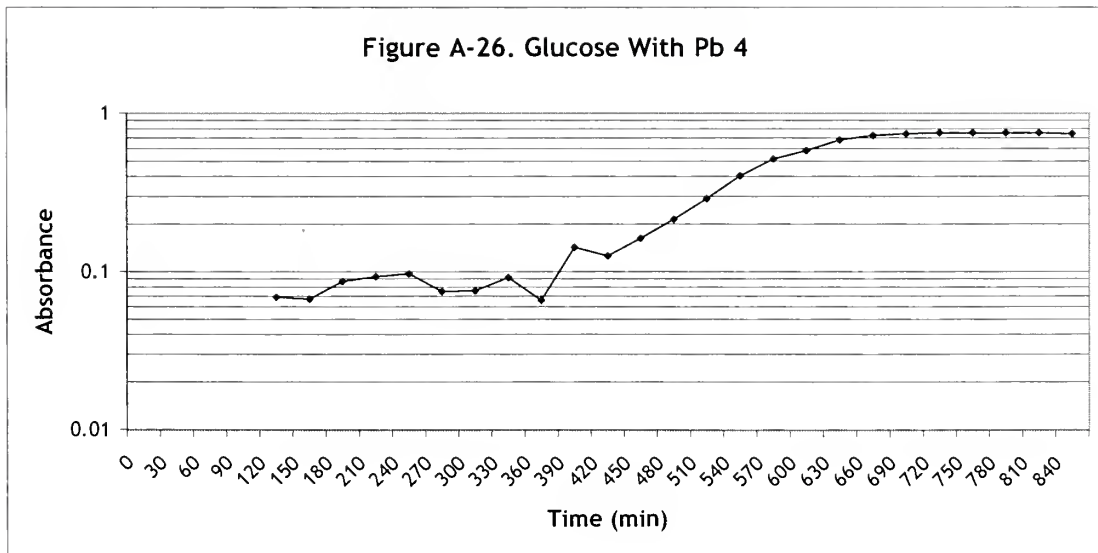
Absorbance increases from 0.303 to 0.608

Actual growth rate = (600 min - 510 min) = 90 min



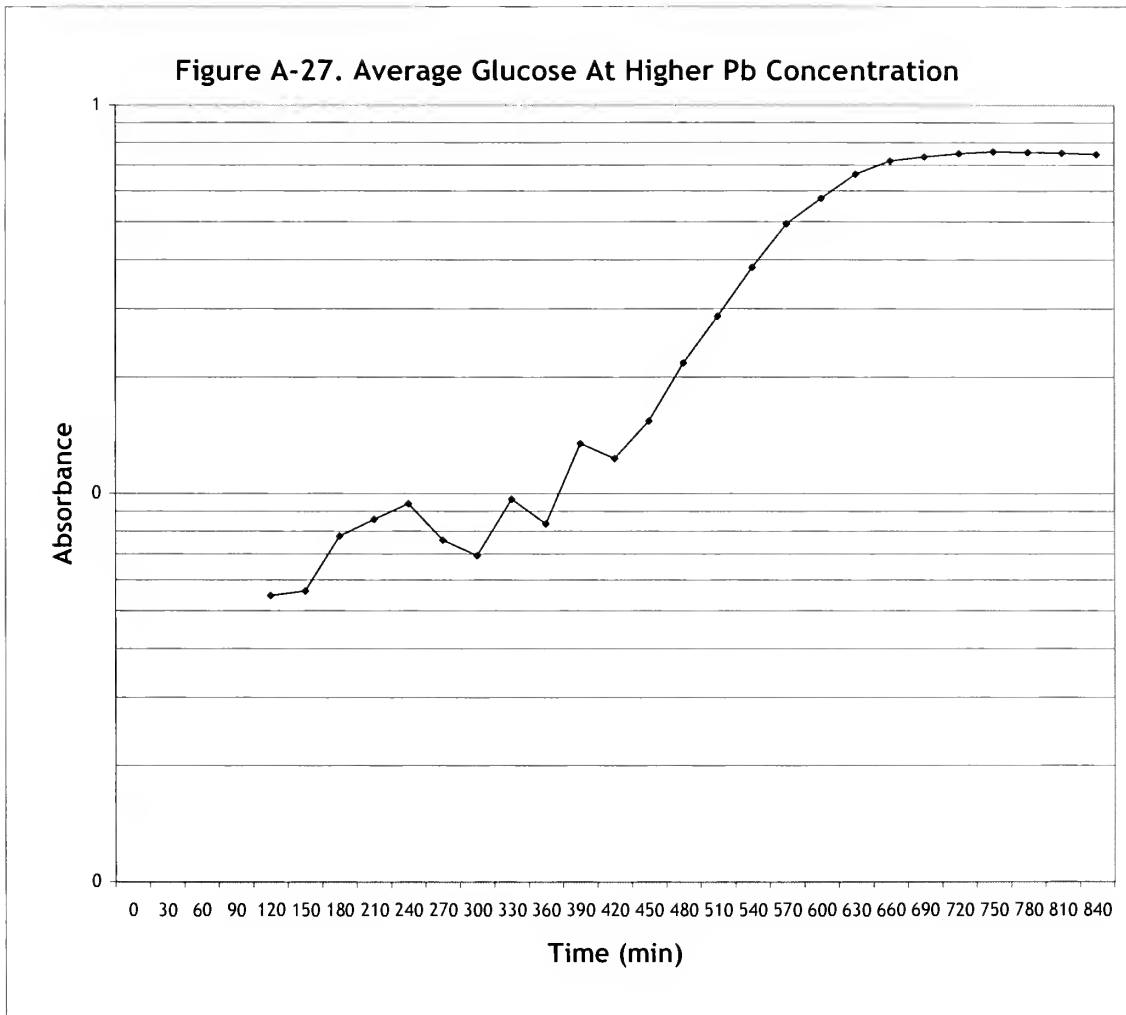
Absorbance increases from 0.325 to 0.648

Actual growth rate = (630 min - 540 min) = 90 min



Absorbance increases from 0.290 to 0.584

Actual growth rate = (600 min - 510 min) = 90 min



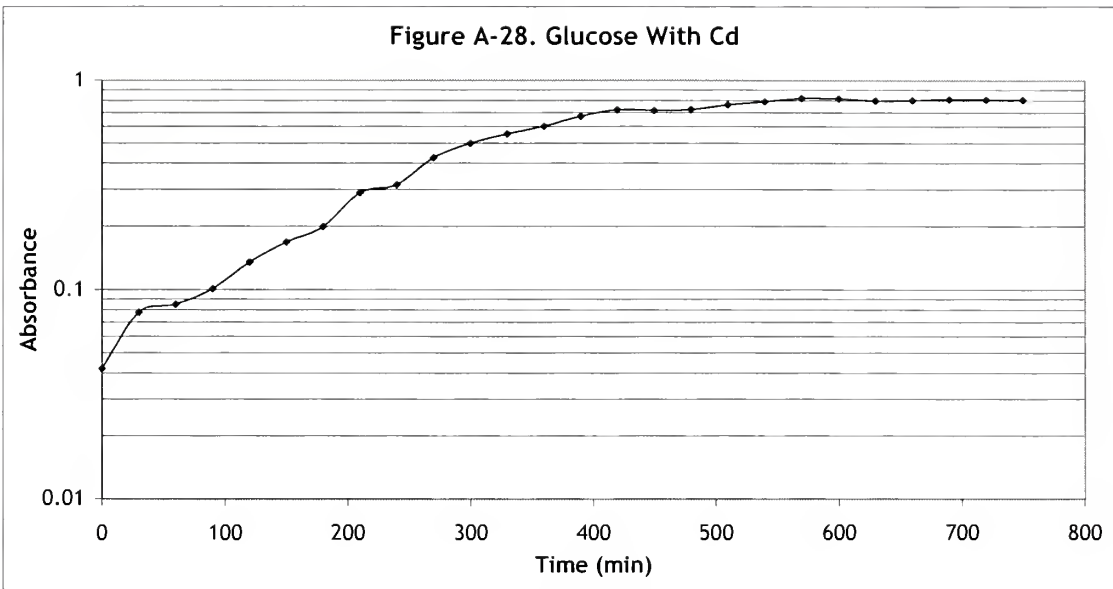
Absorbance increases from 0.287 to 0.575

Actual growth rate = (600 min - 510 min) = 90 min

Data for *E.coli* from absorbance vs. time in M9 media

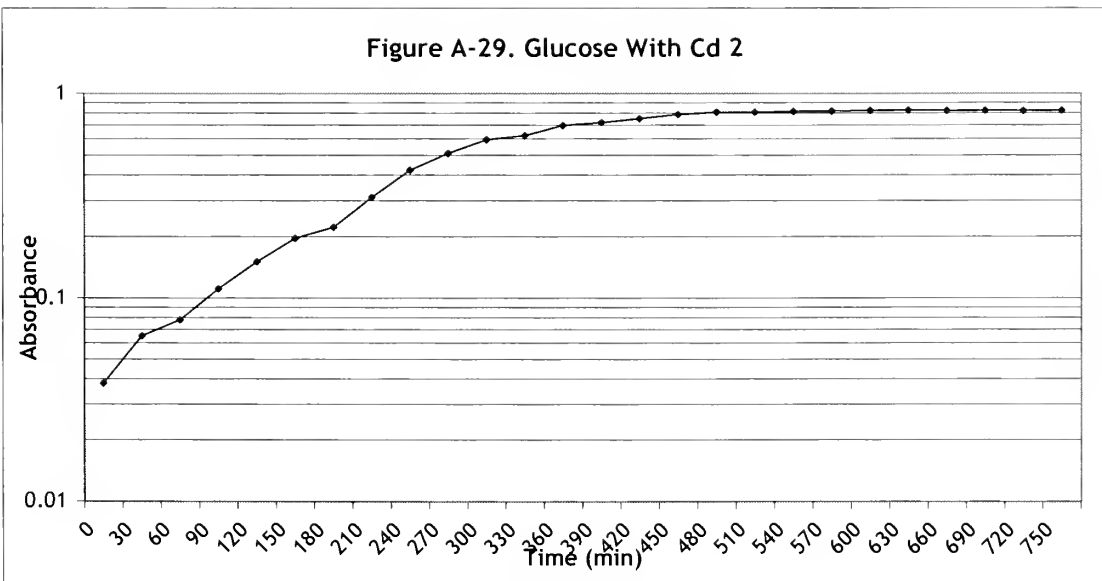
Glucose and Cd at $1 \times 10^{-5} \text{M}$

Time(min)	Run1	Run2	Run3	Average
0	0	0	0	0
0	0.042	0.038	0.041	0.040
30	0.078	0.065	0.072	0.072
60	0.085	0.078	0.082	0.082
90	0.101	0.111	0.108	0.107
120	0.135	0.151	0.142	0.143
150	0.168	0.197	0.183	0.183
180	0.199	0.223	0.217	0.213
210	0.289	0.311	0.302	0.301
240	0.316	0.423	0.374	0.371
270	0.426	0.511	0.464	0.467
300	0.499	0.594	0.537	0.543
330	0.553	0.623	0.583	0.586
360	0.602	0.698	0.652	0.651
390	0.672	0.720	0.691	0.694
420	0.722	0.754	0.733	0.736
450	0.716	0.788	0.754	0.753
480	0.725	0.811	0.769	0.768
510	0.763	0.810	0.785	0.786
540	0.792	0.815	0.801	0.803
570	0.821	0.819	0.812	0.817
600	0.816	0.826	0.818	0.820
630	0.799	0.828	0.821	0.816
660	0.802	0.824	0.819	0.815
690	0.812	0.826	0.823	0.820
720	0.808	0.822	0.825	0.818
750	0.806	0.824	0.824	0.818



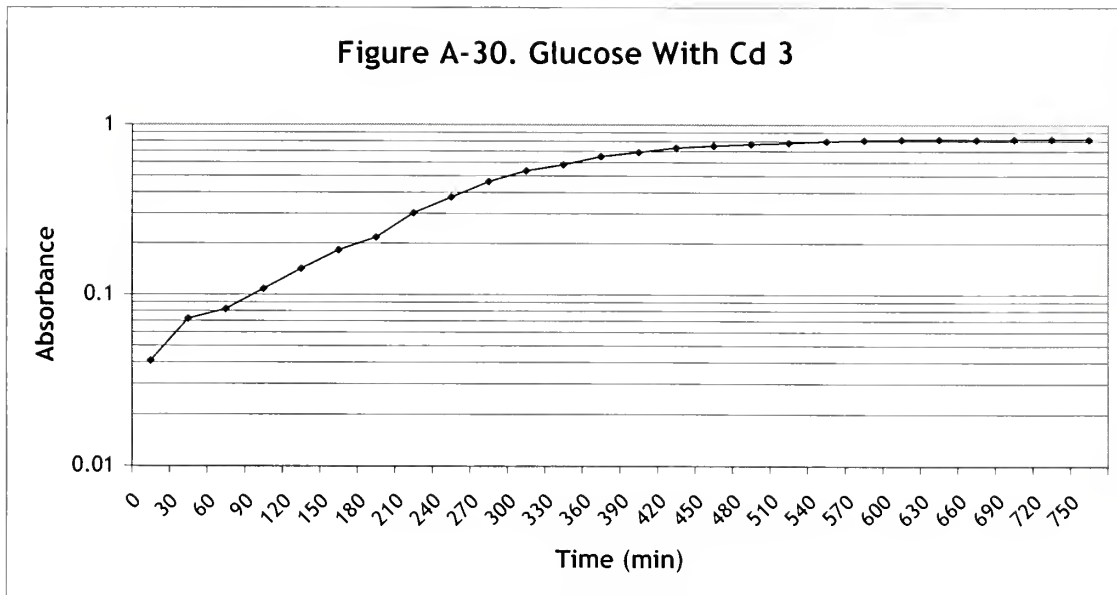
Absorbance increases from 0.101 to 0.199

Actual growth rate = (180 min - 90 min) = 90 min



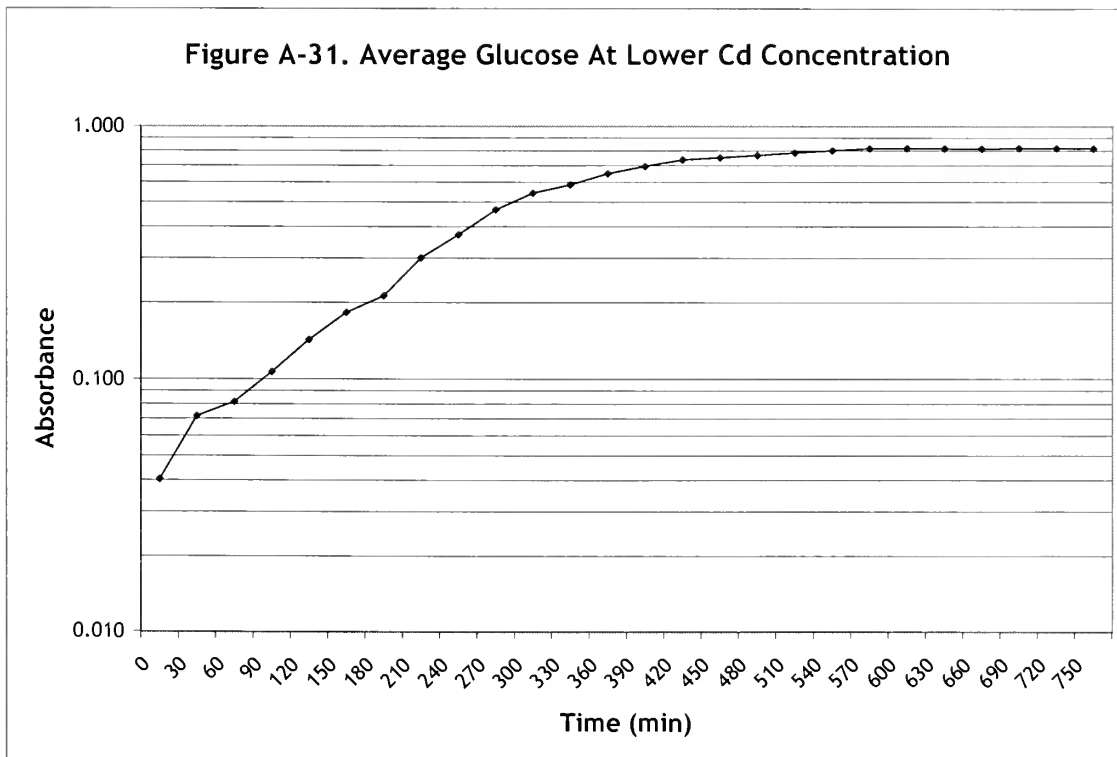
Absorbance increases from 0.111 to .0.223

Actual growth rate = (180 min - 90 min) = 90 min



Absorbance increases from 0.108 to .0.217

Actual growth rate = (180 min - 90 min) = 90 min



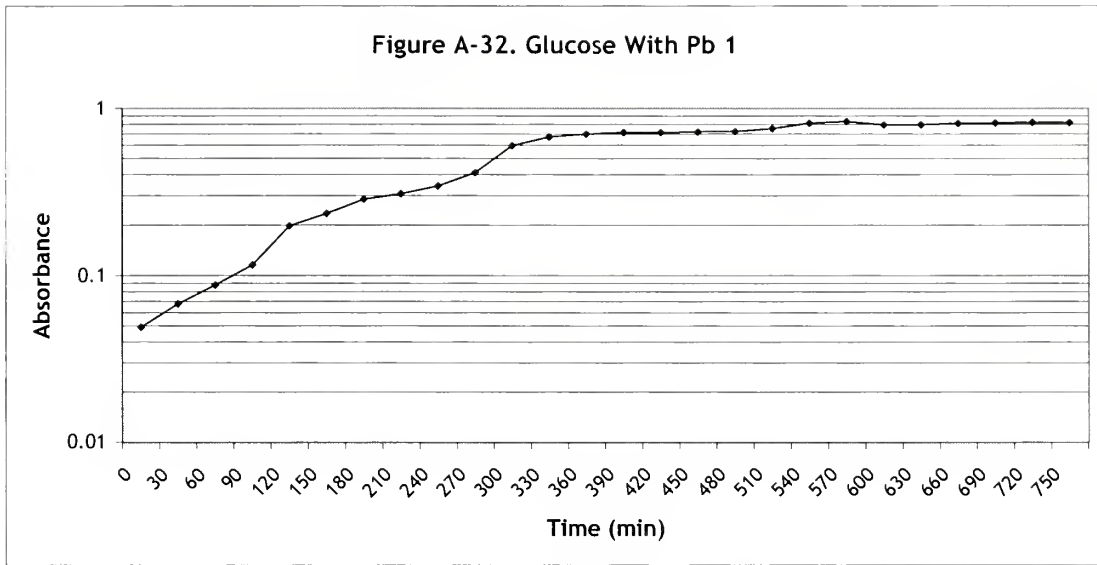
Absorbance increases from 0.107 to .0.213

Actual growth rate = (180 min - 90 min) = 90 min

Data for *E. coli* from absorbance vs. time in M9 media

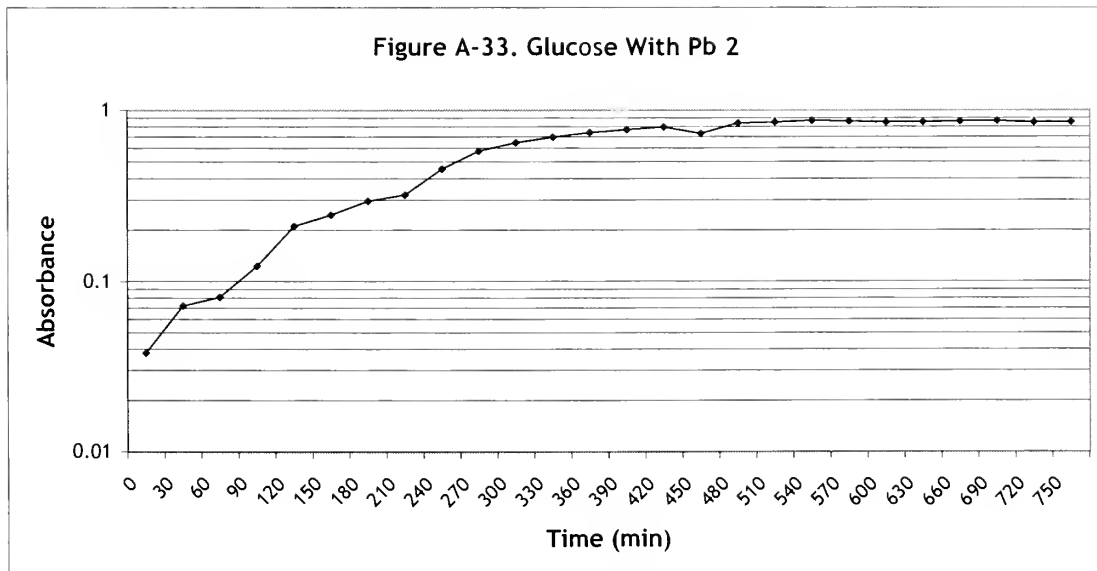
Glucose and Pb at $1 \times 10^{-5} M$

Time(min)	Run1	Run2	Run3	Average
0	0	0	0	0
0	0.049	0.038	0.042	0.043
30	0.068	0.072	0.067	0.069
60	0.088	0.081	0.084	0.084
90	0.116	0.123	0.118	0.119
120	0.198	0.211	0.202	0.204
150	0.235	0.245	0.237	0.239
180	0.285	0.296	0.288	0.290
210	0.308	0.322	0.314	0.315
240	0.342	0.456	0.386	0.395
270	0.411	0.578	0.491	0.493
300	0.596	0.646	0.617	0.620
330	0.672	0.698	0.689	0.686
360	0.702	0.741	0.717	0.720
390	0.718	0.771	0.739	0.743
420	0.716	0.796	0.746	0.753
450	0.722	0.732	0.752	0.735
480	0.725	0.842	0.764	0.777
510	0.756	0.852	0.798	0.802
540	0.814	0.870	0.835	0.840
570	0.834	0.861	0.847	0.847
600	0.795	0.852	0.851	0.833
630	0.798	0.855	0.839	0.831
660	0.812	0.862	0.842	0.839
690	0.815	0.865	0.845	0.842
720	0.822	0.852	0.847	0.840
750	0.818	0.855	0.842	0.838



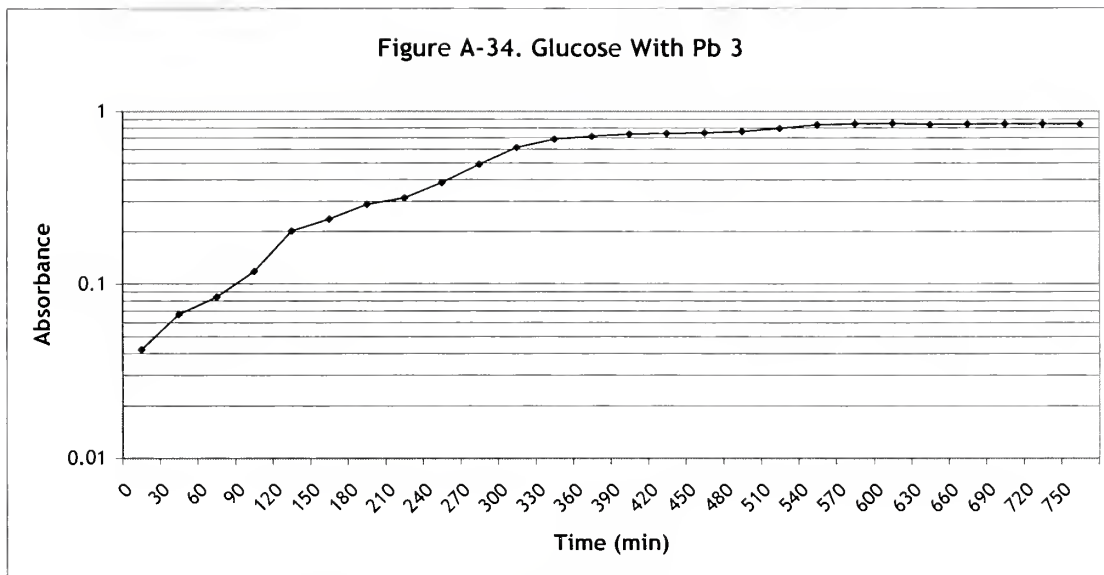
Absorbance increases from 0.116 to 0.235

Actual growth rate = (150 min - 90 min) = 60 min



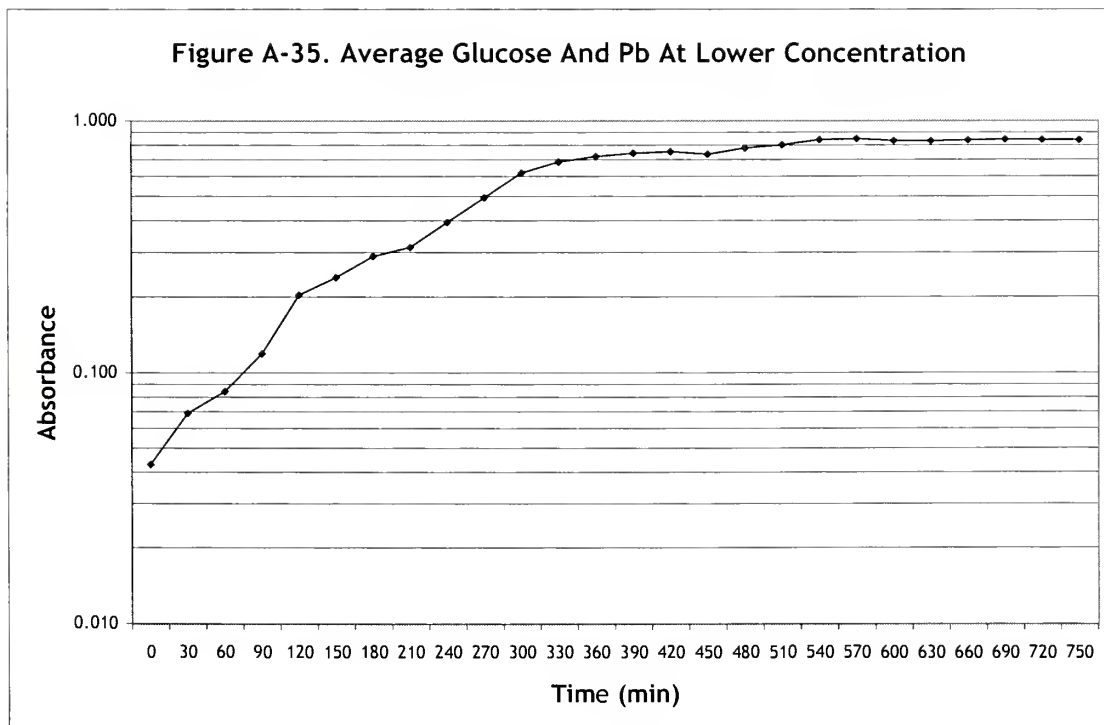
Absorbance increases from 0.123 to 0.245

Actual growth rate = (150 min - 90 min) = 60 min



Absorbance increases from 0.118 to 0.237

Actual growth rate = (150 min - 90 min) = 60 min

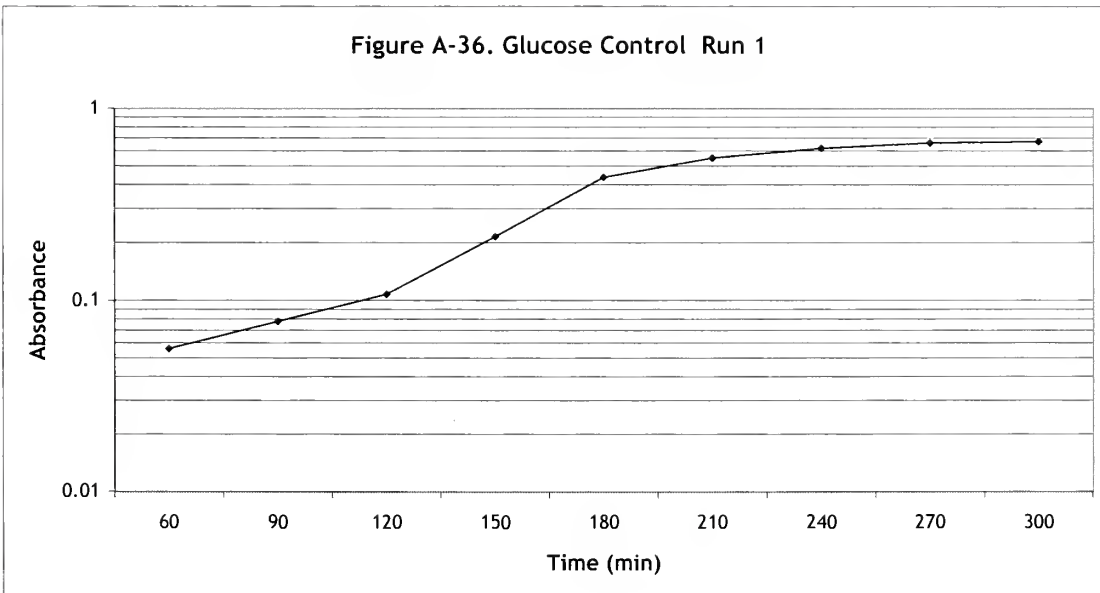


Absorbance increases from 0.119 to .0.239

Actual growth rate = (150 min - 90 min) = 60 min

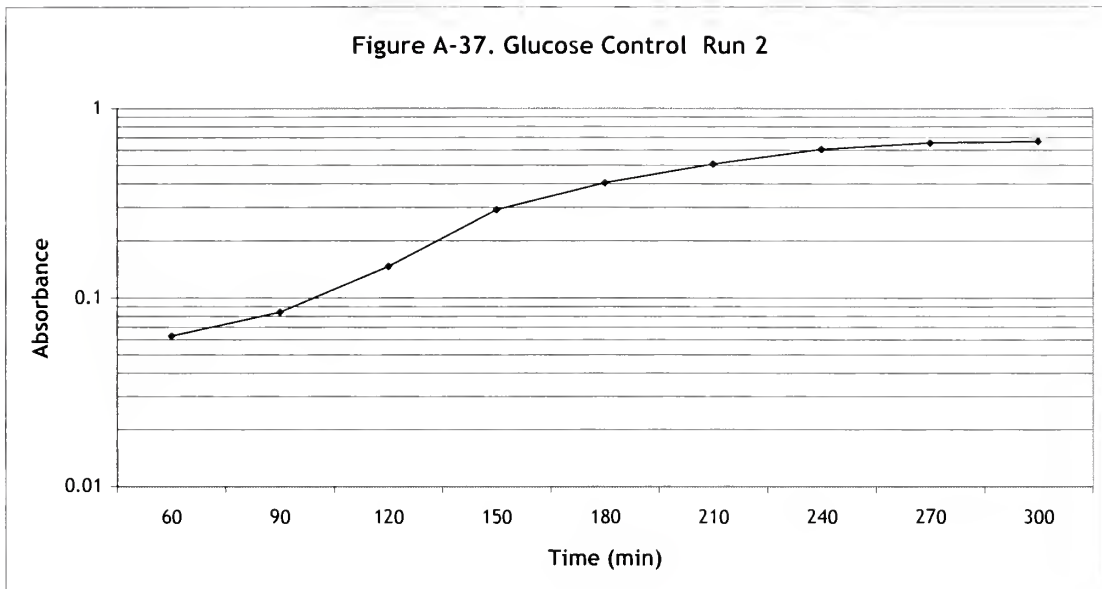
Data for *E.coli* from absorbance vs. time in LB Broth- Rich Media

Glucose control					
Time(min)	Run1	Run2	Run3	Run4	Average
0					
30					
60	0.056	0.063	0.058	0.057	0.059
90	0.078	0.084	0.081	0.077	0.080
120	0.108	0.147	0.116	0.132	0.126
150	0.215	0.292	0.234	0.261	0.251
180	0.436	0.404	0.421	0.412	0.418
210	0.550	0.508	0.521	0.524	0.526
240	0.618	0.608	0.612	0.615	0.613
270	0.660	0.658	0.657	0.653	0.657
300	0.672	0.668	0.681	0.669	0.673



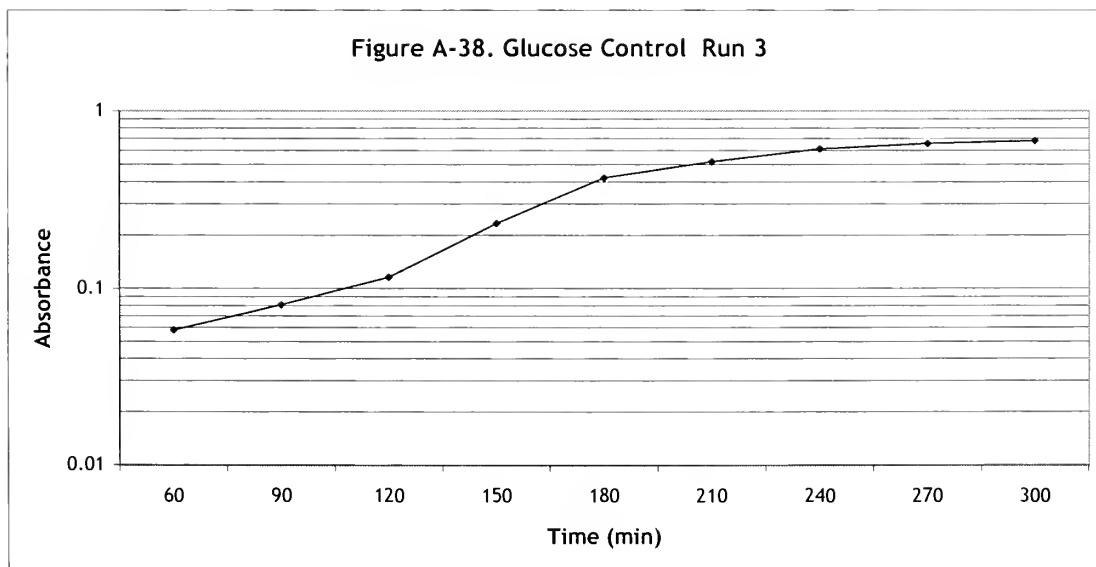
Absorbance increases from 0.108 to .0.215

Actual growth rate = (150 min -120 min) = 30 min



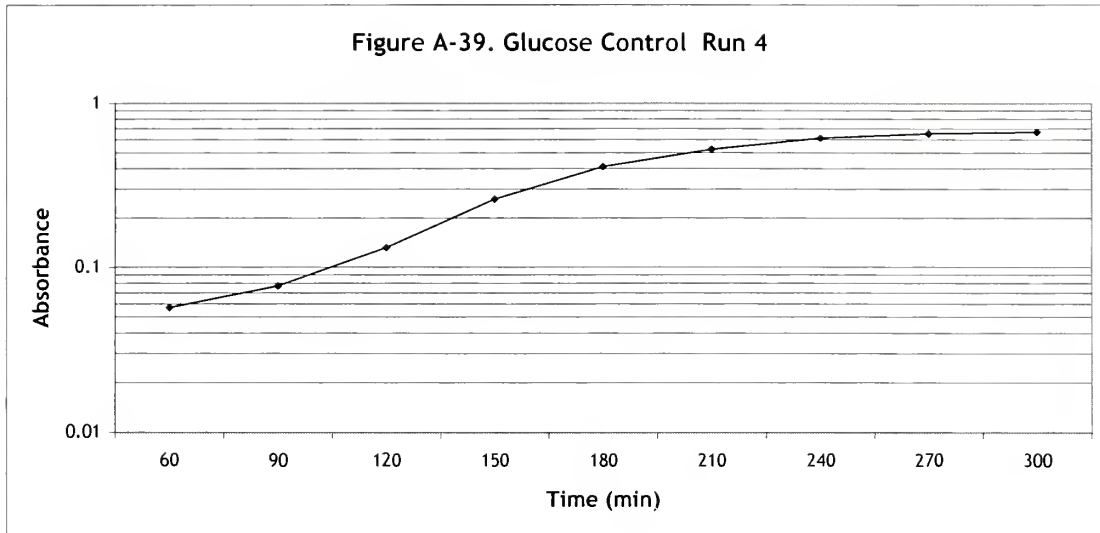
Absorbance increases from 0.147 to .0.292

Actual growth rate = (150 min -120 min) = 30 min



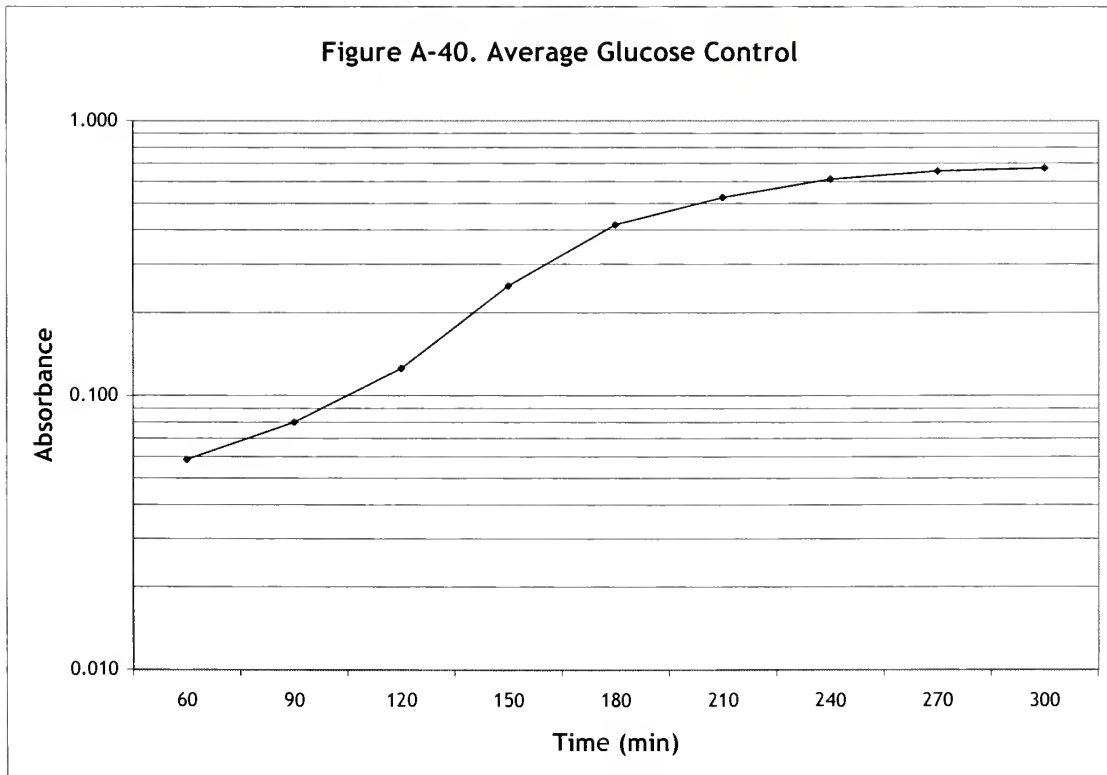
Absorbance increases from 0.116 to .0.234

Actual growth rate = (150 min -120 min) = 30 min



Absorbance increases from 0.132 to .0.261

Actual growth rate = (150 min - 120 min) = 30 min

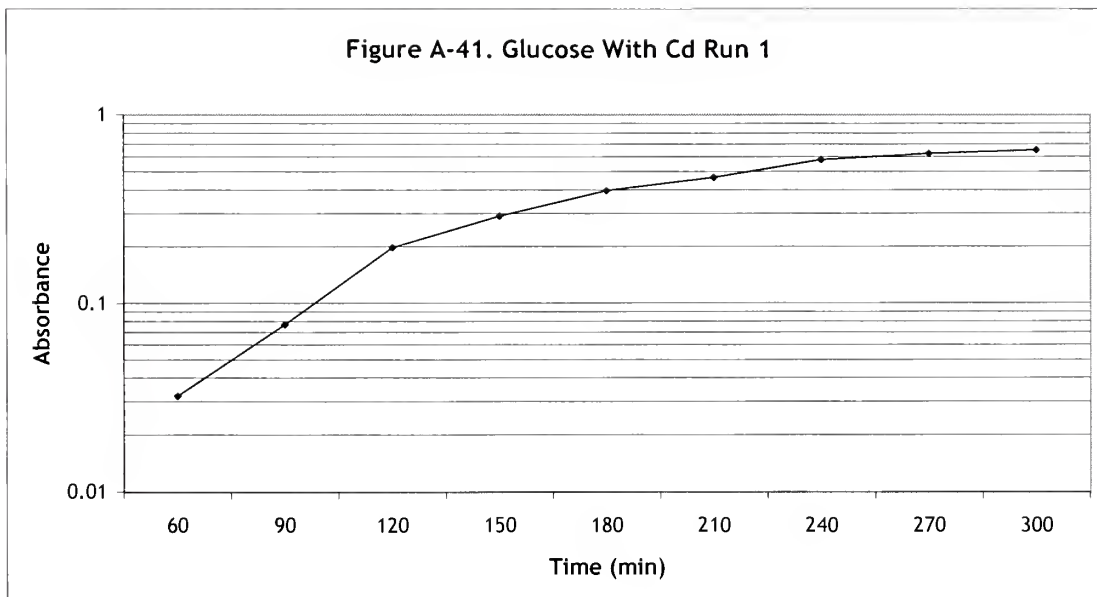


Absorbance increases from 0.126 to .0.251

Actual growth rate = (150 min - 120 min) = 30 min

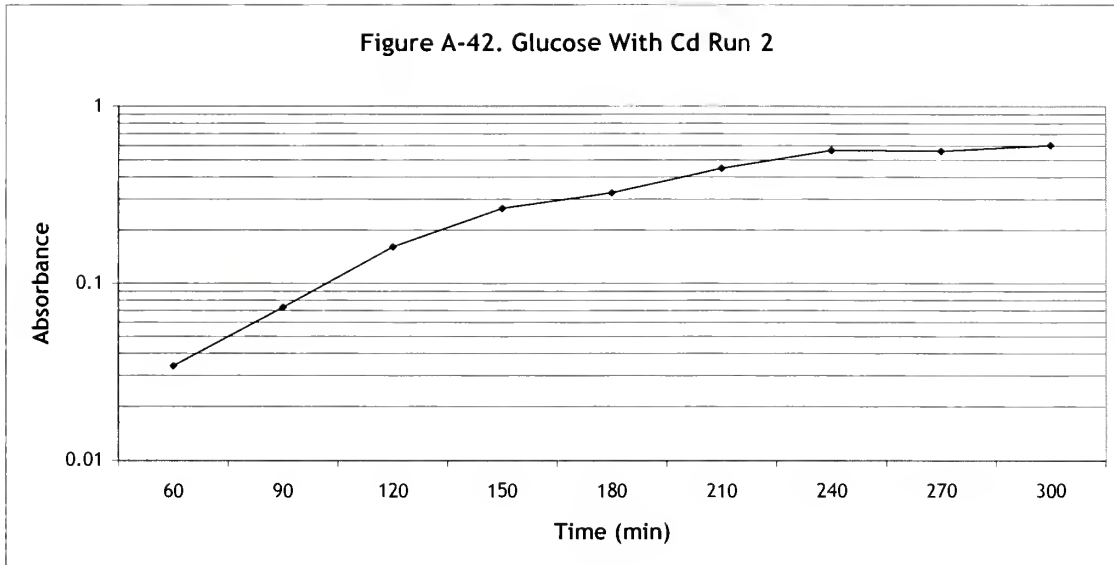
Data for *E.coli* from absorbance vs. time in LB Broth- Rich Media

Glucose With $1 \cdot 10^{-5}$ M Cd					
Time(min)	Run1	Run2	Run3	Run4	Average
0					
30					
60	0.032	0.034	0.028	0.031	0.031
90	0.077	0.073	0.069	0.072	0.073
120	0.198	0.161	0.172	0.176	0.177
150	0.291	0.266	0.254	0.258	0.267
180	0.397	0.326	0.344	0.351	0.355
210	0.467	0.448	0.432	0.441	0.447
240	0.578	0.567	0.598	0.561	0.576
270	0.624	0.562	0.642	0.641	0.617
300	0.652	0.605	0.682	0.678	0.654



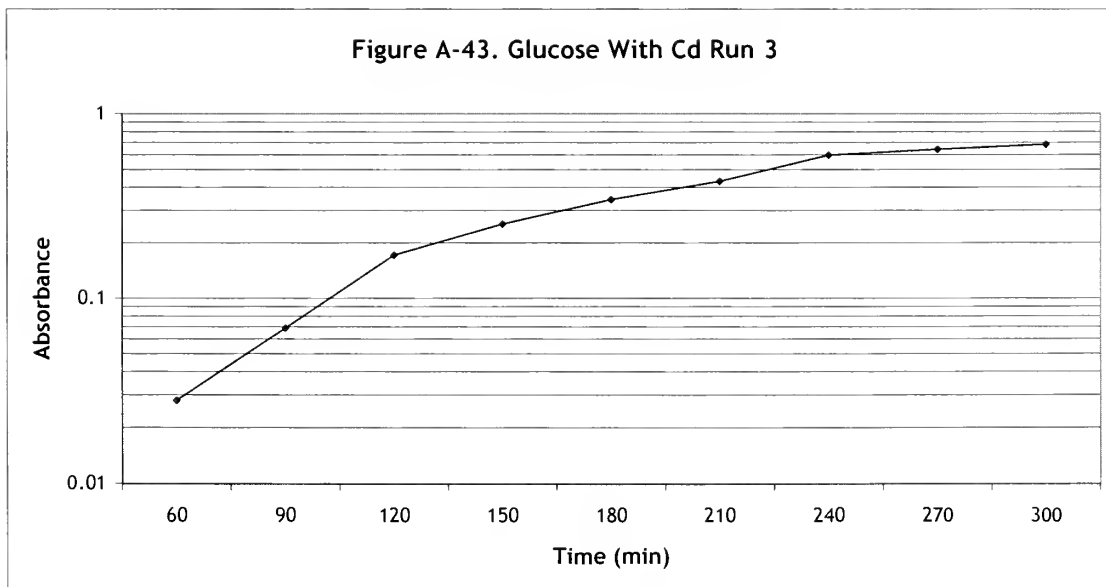
Absorbance increases from 0.198 to .0.397

Actual growth rate = (180 min - 120min) = 60 min



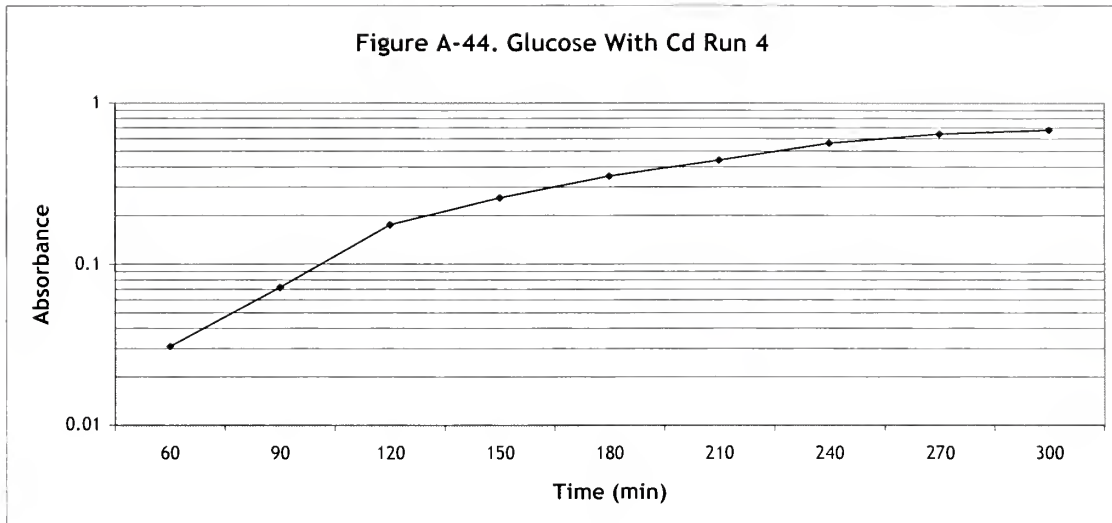
Absorbance increases from 0.161 to .0.326

Actual growth rate = (180 min - 120min) = 60 min



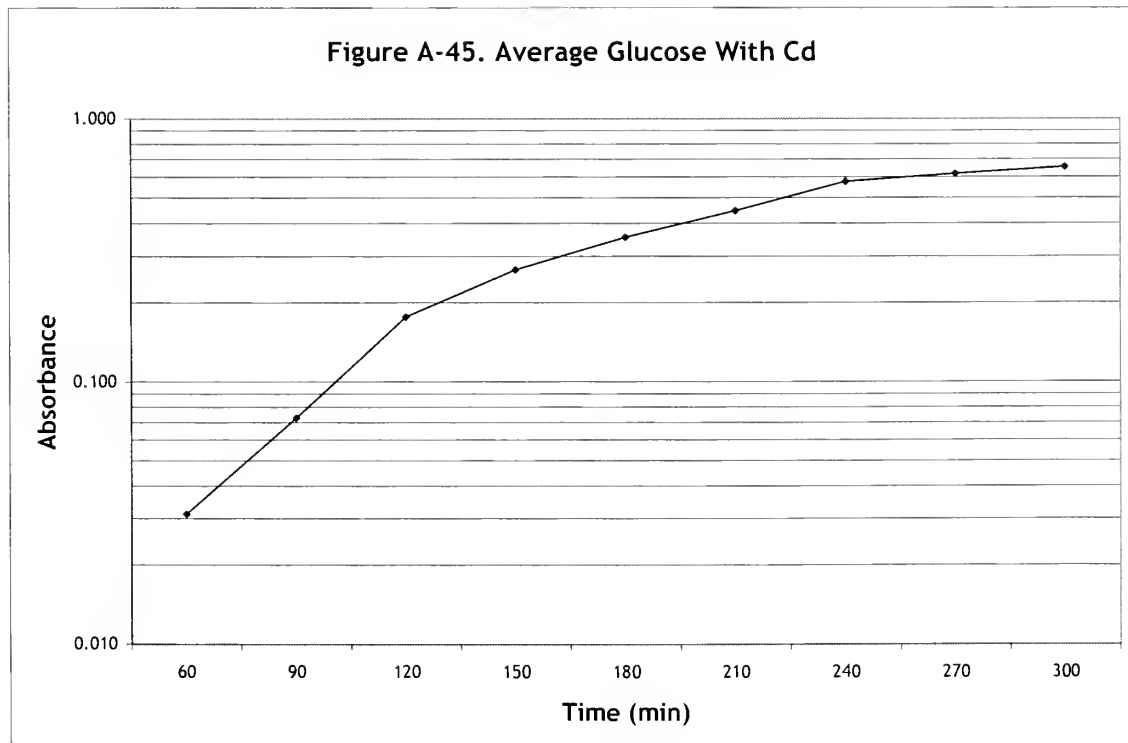
Absorbance increases from 0.172 to .0.344

Actual growth rate = (180 min - 120min) = 60 min



Absorbance increases from 0.176 to .0.351

Actual growth rate = (180 min - 120min) = 60 min

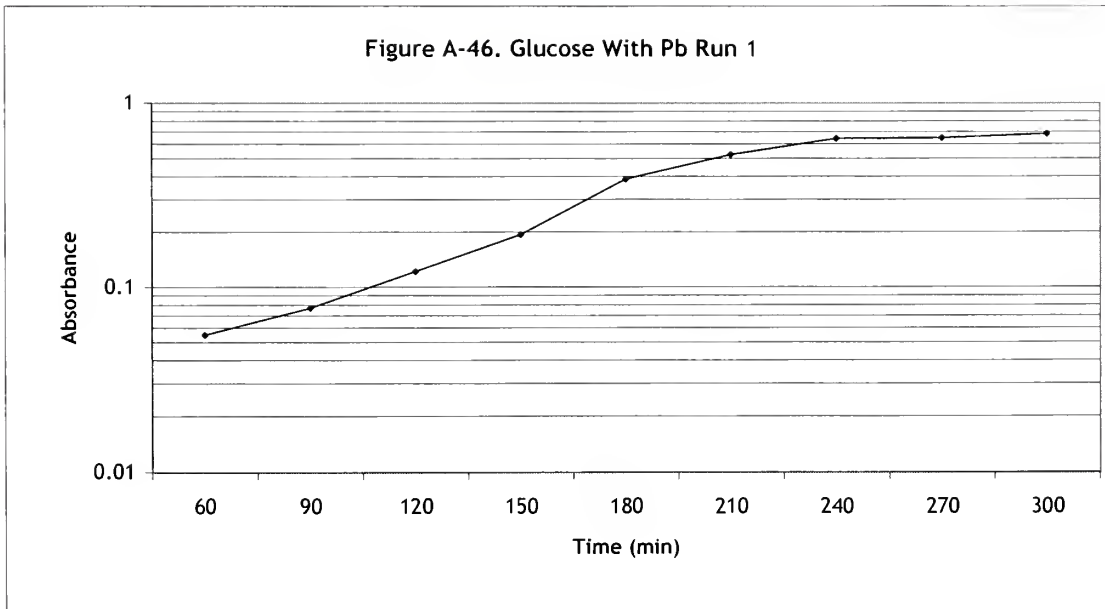


Absorbance increases from 0.177 to .0.355

Actual growth rate = (180 min - 120min) = 60 min

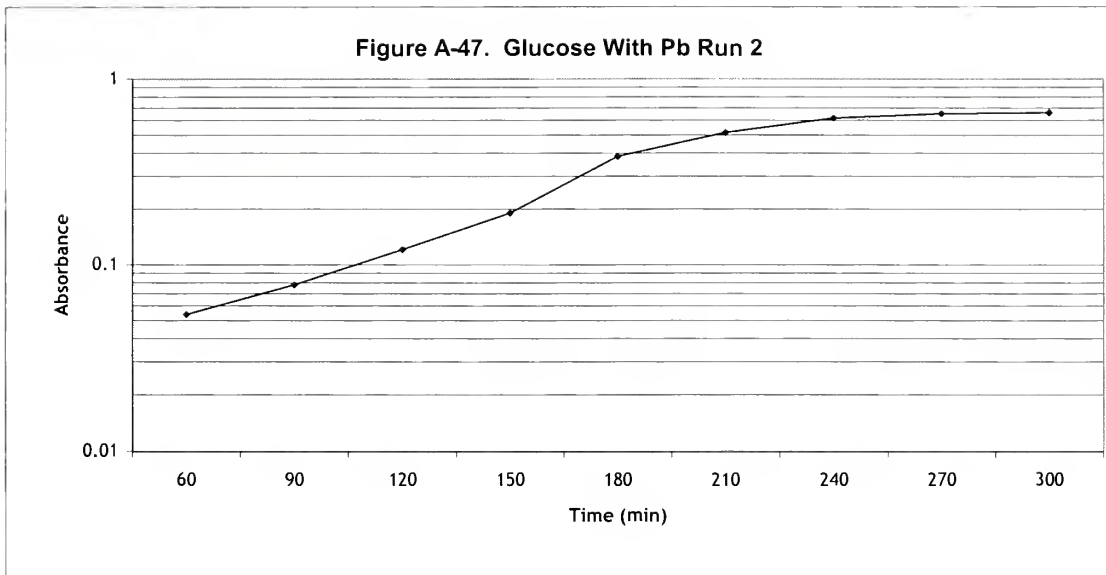
Data for *E. coli* from absorbance vs. time in LB Broth- Rich Media

Glucose With 1×10^{-5} M Pb					
Time(min)	Run1	Run2	Run3	Run4	Average
0					
30					
60	0.055	0.054	0.028	0.048	0.046
90	0.077	0.078	0.052	0.071	0.070
120	0.122	0.121	0.101	0.108	0.113
150	0.193	0.191	0.185	0.181	0.188
180	0.388	0.384	0.369	0.366	0.377
210	0.526	0.516	0.412	0.504	0.490
240	0.642	0.618	0.484	0.611	0.589
270	0.648	0.652	0.578	0.648	0.632
300	0.682	0.661	0.663	0.674	0.670



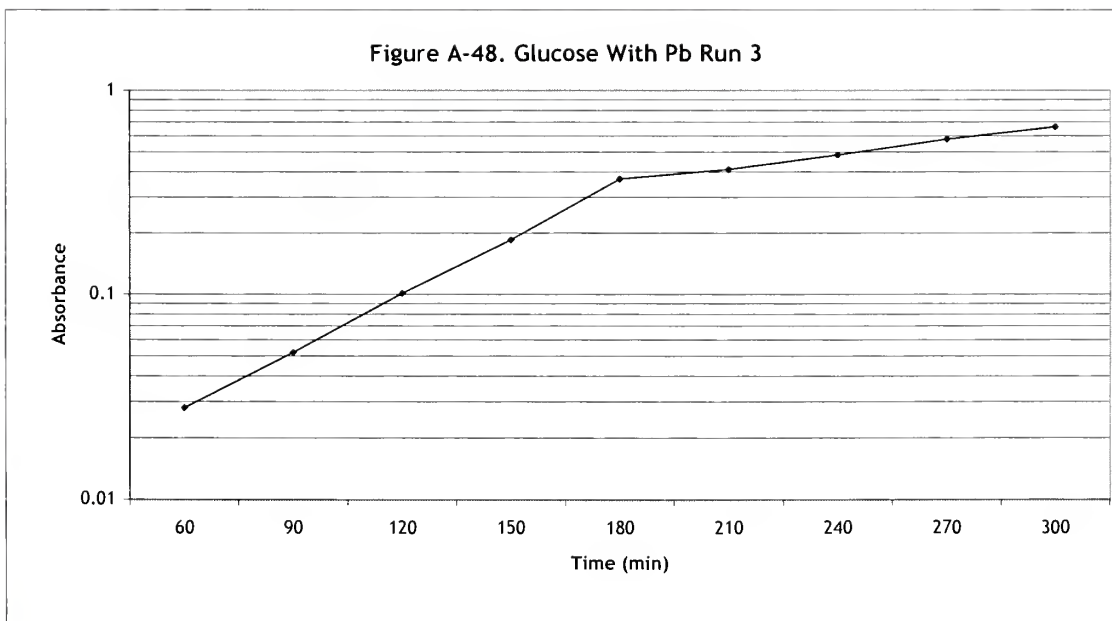
Absorbance increases from 0.193 to .0.388

Actual growth rate = (180 min -150 min) = 30 min



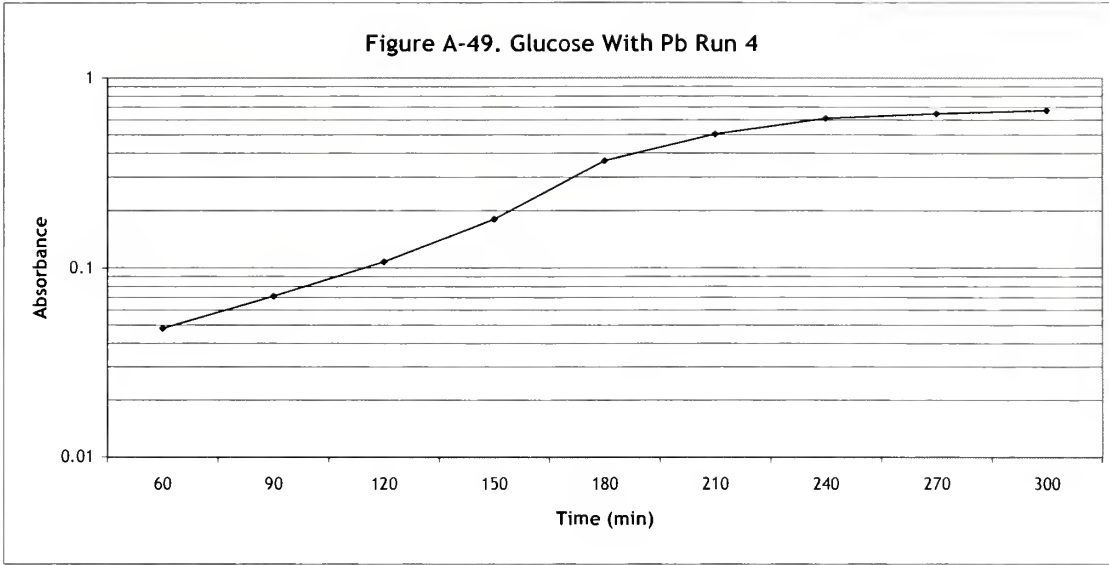
Absorbance increases from 0.191 to .0.384

Actual growth rate = (180 min - 150 min) = 30 min



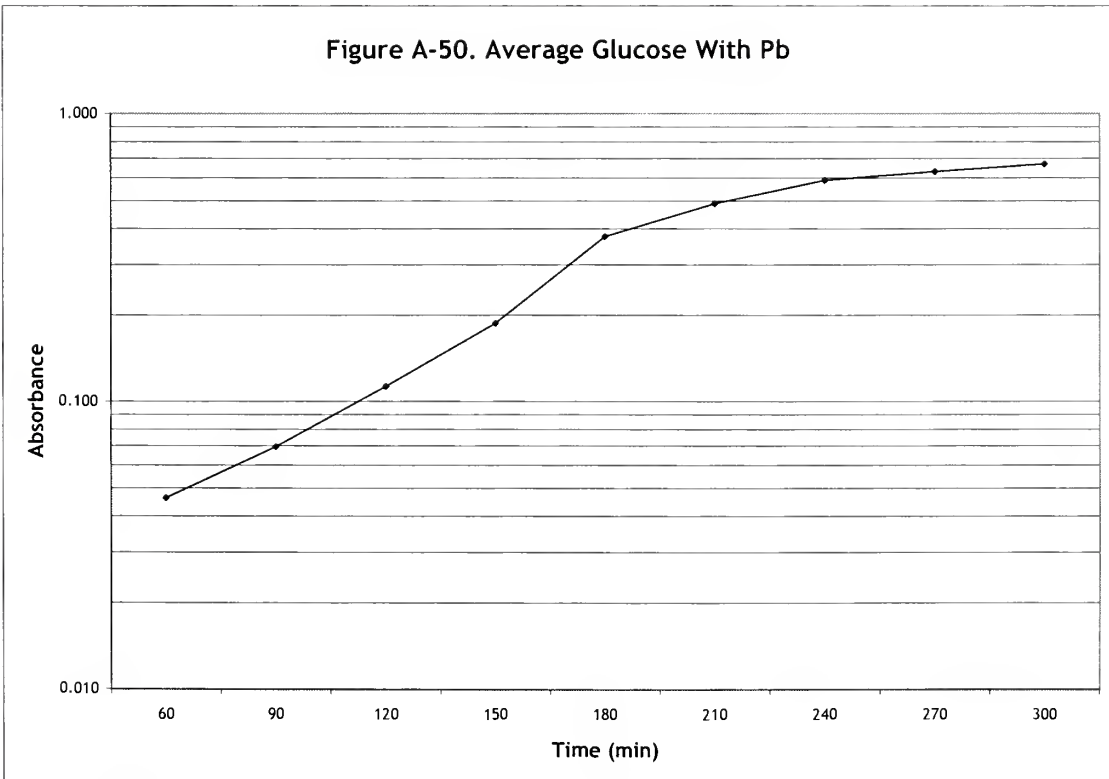
Absorbance increases from 0.185 to .0.369

Actual growth rate = (180 min - 150 min) = 30 min



Absorbance increases from 0.181 to .0.366

Actual growth rate = (150 min -120 min) = 30 min

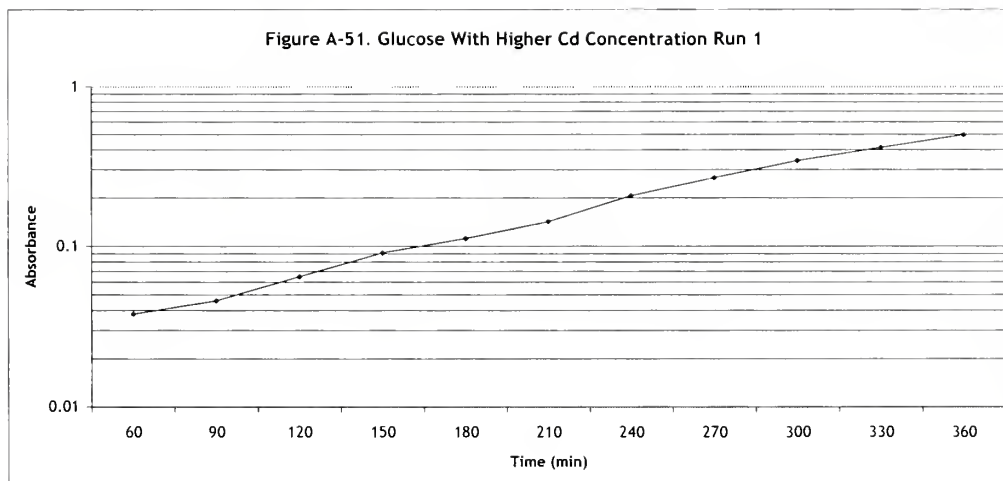


Absorbance increases from 0.188 to .0.377

Actual growth rate = (150 min -120 min) = 30 min

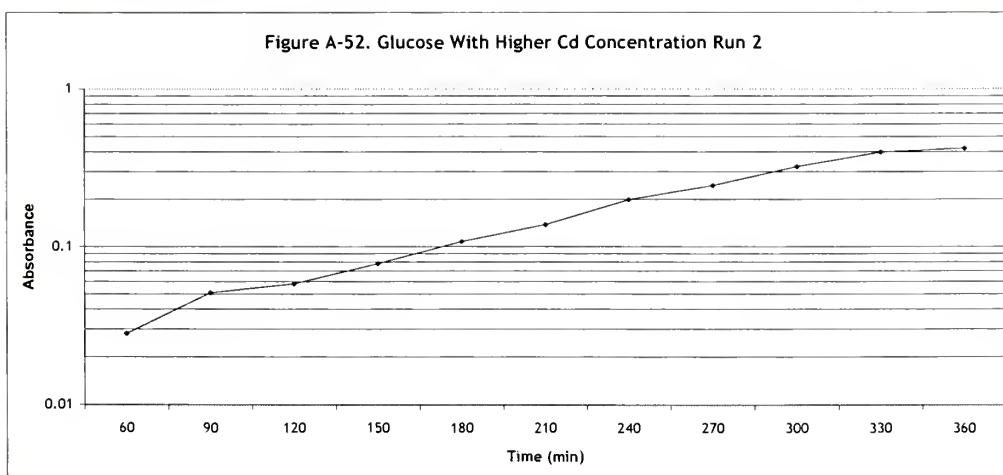
Data for *E.coli* from absorbance vs. time in LB Broth- Rich Media

Glucose With 1×10^{-3} M Cd				
Time(min)	Run1	Run2	Run3	Average
0				
30				
60	0.038	0.028	0.042	0.036
90	0.046	0.051	0.063	0.053
120	0.065	0.058	0.068	0.064
150	0.091	0.078	0.075	0.081
180	0.112	0.108	0.098	0.106
210	0.142	0.138	0.114	0.131
240	0.206	0.199	0.204	0.203
270	0.268	0.245	0.242	0.252
300	0.342	0.323	0.298	0.321
330	0.414	0.399	0.409	0.407
360	0.498	0.422	0.455	0.458



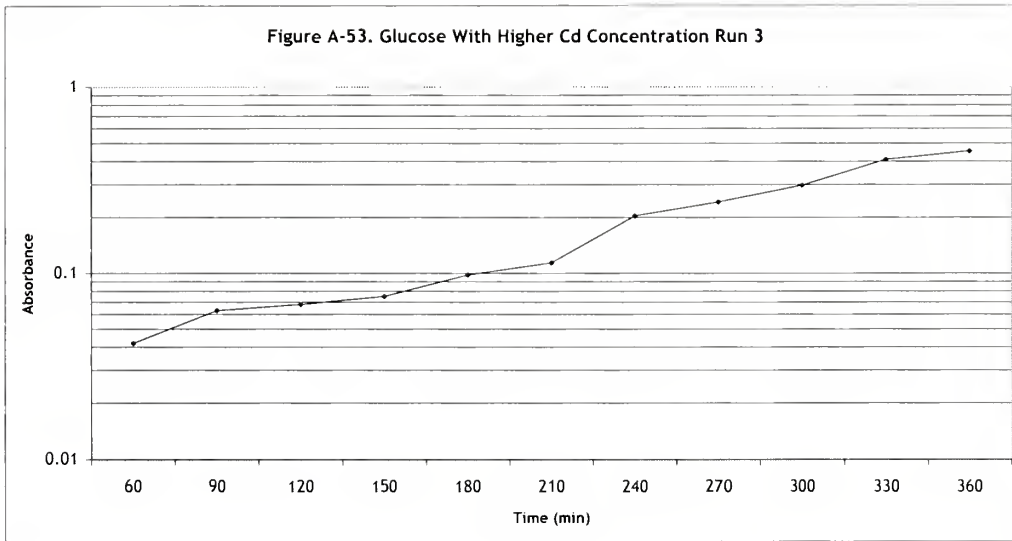
Absorbance increases from 0.206 to .0.414

Actual growth rate = (330 min - 240min) = 90 min



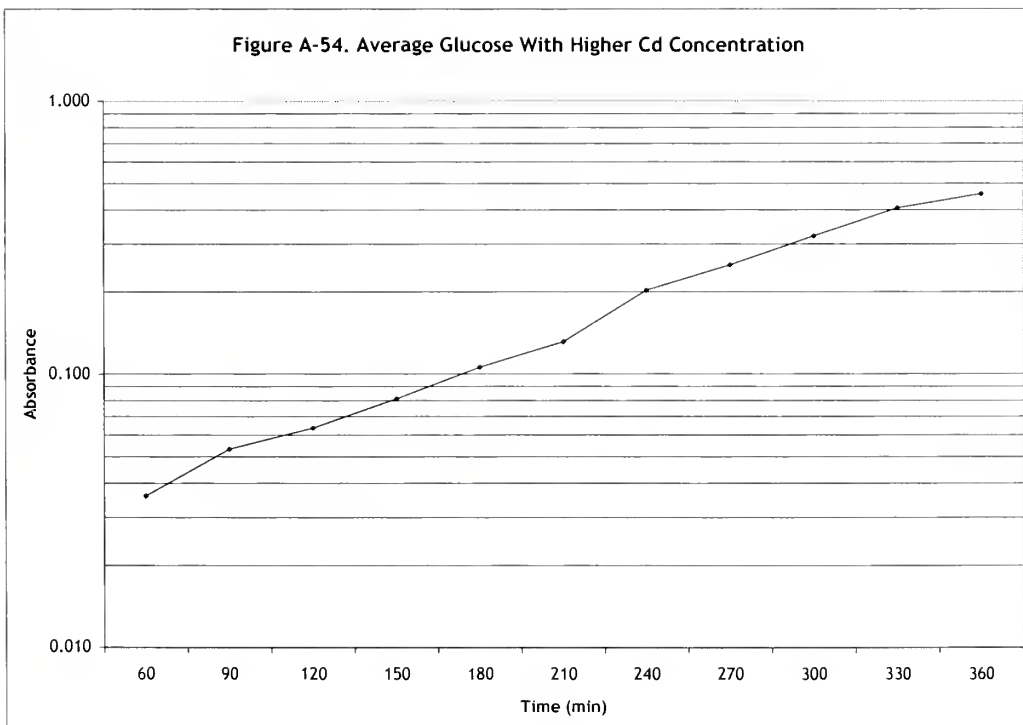
Absorbance increases from 0.199 to .0.399

Actual growth rate = (330 min - 240min) = 90 min



Absorbance increases from 0.204 to .0.409

Actual growth rate = (330 min - 240min) = 90 min



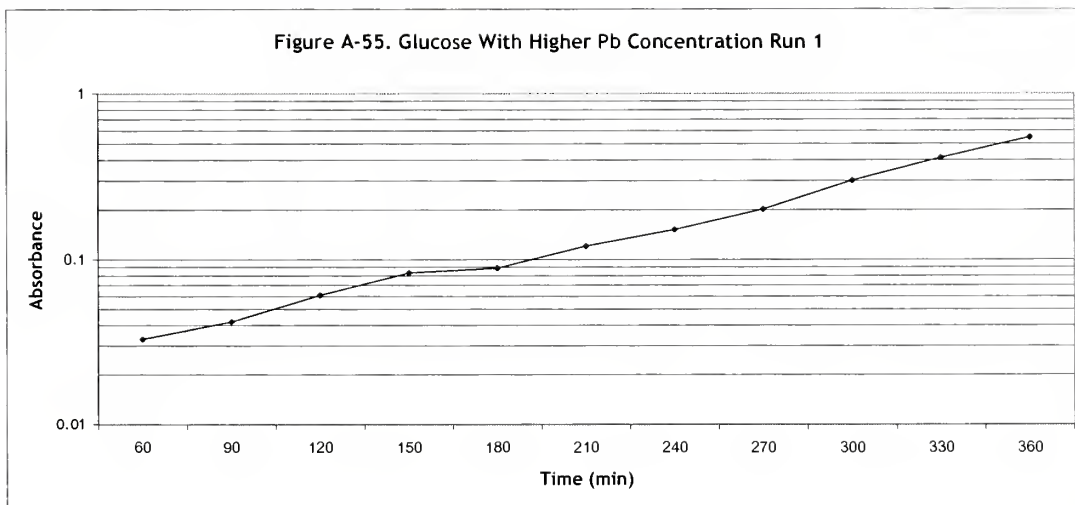
Absorbance increases from 0.203 to .0.407

Actual growth rate = (330 min - 240min) = 90 min

Data for *E. coli* from absorbance vs. time in LB Broth- Rich Media

Glucose With 1×10^{-3} M Pb				
Time(min)	Run1	Run2	Run3	Average
0				
30				
60	0.033	0.028	0.039	0.033
90	0.042	0.034	0.054	0.043
120	0.061	0.039	0.071	0.057
150	0.083	0.052	0.092	0.076
180	0.089	0.071	0.096	0.085
210	0.121	0.101	0.108	0.110
240	0.152	0.138	0.144	0.145
270	0.202	0.196	0.175	0.191
300	0.301	0.279	0.217	0.266
330	0.412	0.375	0.354	0.380
360	0.548	0.414	0.428	0.463

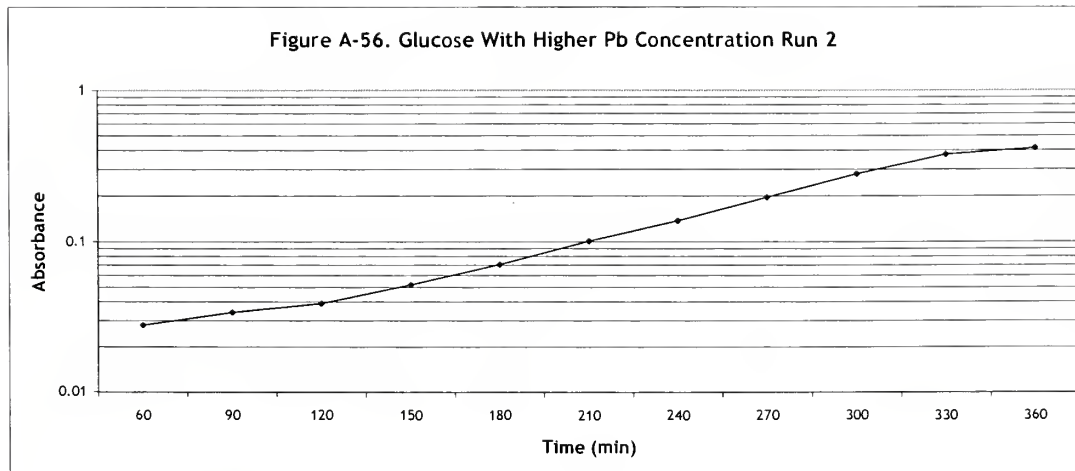
Figure A-55. Glucose With Higher Pb Concentration Run 1



Absorbance increases from 0.152 to .0.301

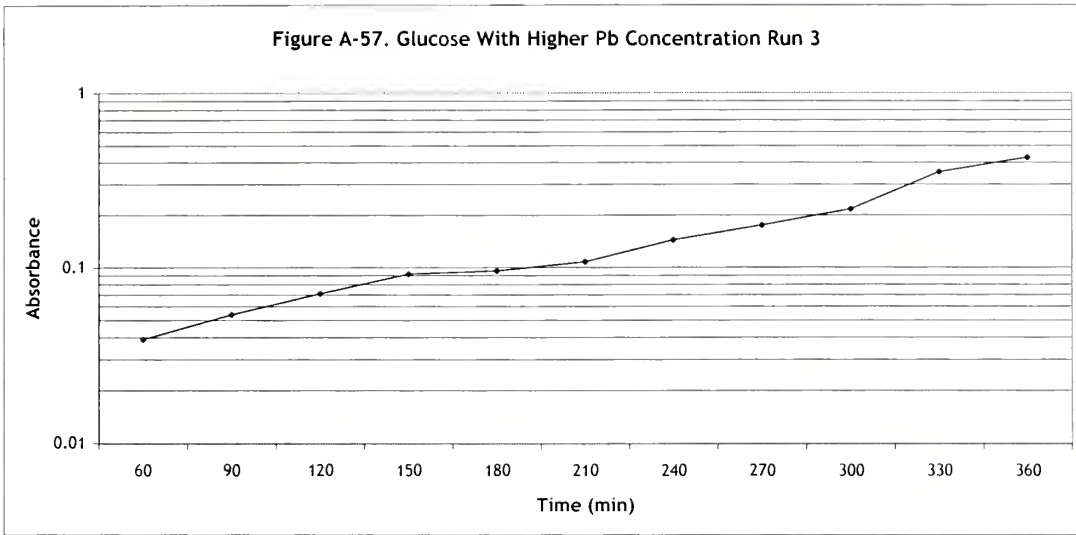
Actual growth rate = (300 min - 240min) = 60 min

Figure A-56. Glucose With Higher Pb Concentration Run 2



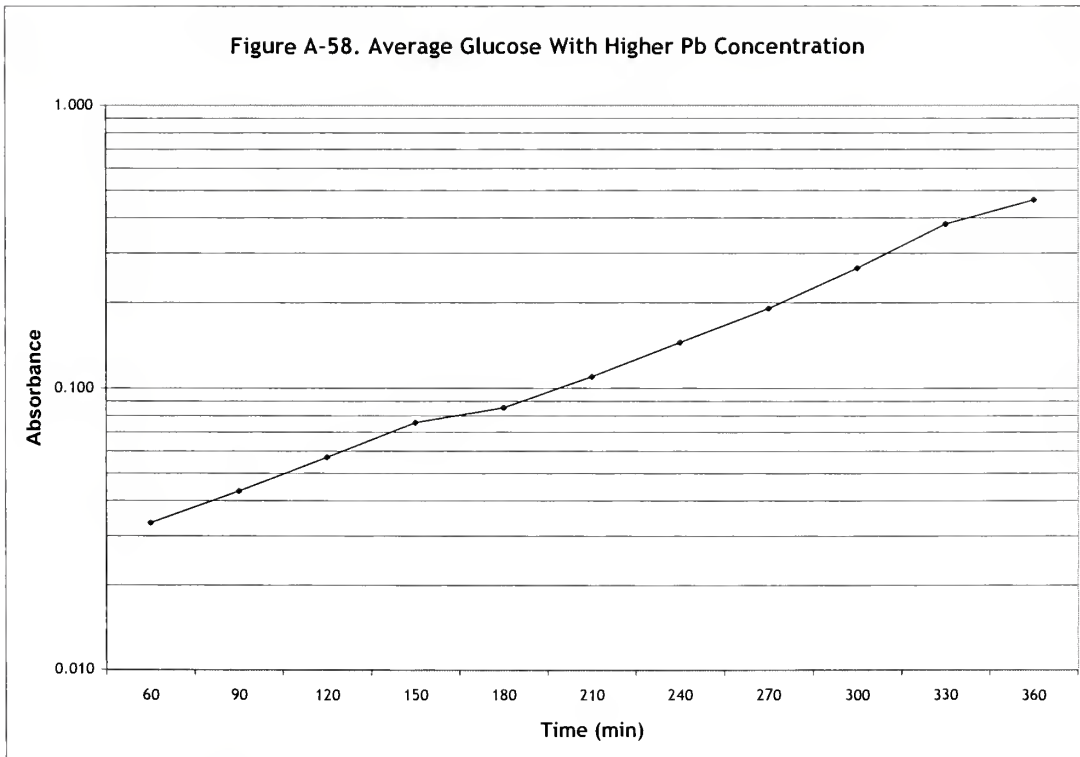
Absorbance increases from 0.138 to .0.279

Actual growth rate = (300 min - 240min) = 60 min



Absorbance increases from 0.108 to .0.217

Actual growth rate = (300 min - 210min) = 90 min



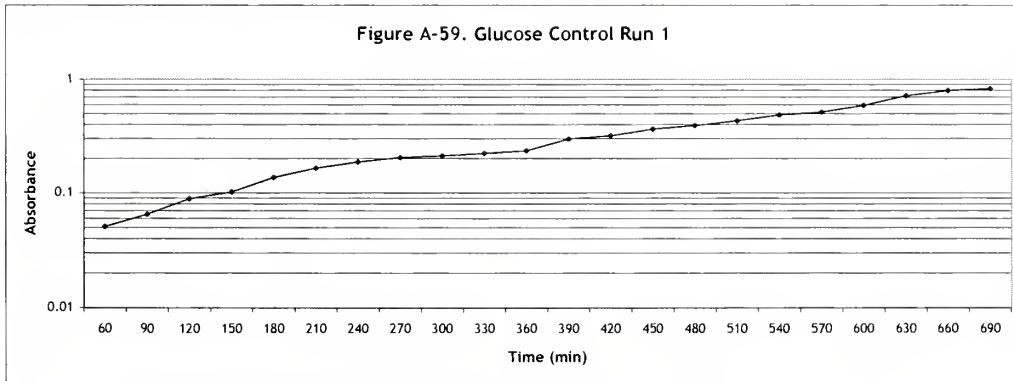
Absorbance increases from 0.191 to .0.380

Actual growth rate = (330 min - 270min) = 60 min

Data for *E. coli* from absorbance vs. time in M9 media with no casamino acids (poor media)

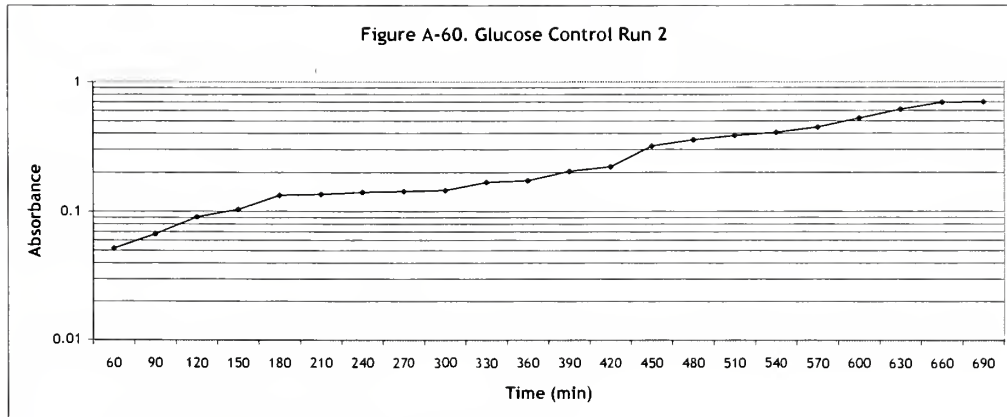
Glucose control				
Time(min)	Run1	Run2	Run3	Average
0				
30				
60	0.051	0.052	0.058	0.052
90	0.065	0.067	0.071	0.066
120	0.089	0.091	0.099	0.090
150	0.102	0.104	0.106	0.103
180	0.137	0.134	0.141	0.136
210	0.165	0.136	0.158	0.151
240	0.187	0.141	0.167	0.164
270	0.204	0.143	0.172	0.174
300	0.211	0.146	0.181	0.179
330	0.222	0.168	0.193	0.195
360	0.234	0.173	0.205	0.204
390	0.298	0.205	0.227	0.243
420	0.318	0.222	0.265	0.268
450	0.365	0.321	0.321	0.336
480	0.391	0.358	0.384	0.378
510	0.432	0.386	0.41	0.409
540	0.485	0.407	0.456	0.446
570	0.514	0.446	0.492	0.480
600	0.592	0.526	0.555	0.559
630	0.718	0.613	0.687	0.666
660	0.794	0.698	0.753	0.746
690	0.826	0.702	0.812	0.764

Figure A-59. Glucose Control Run 1

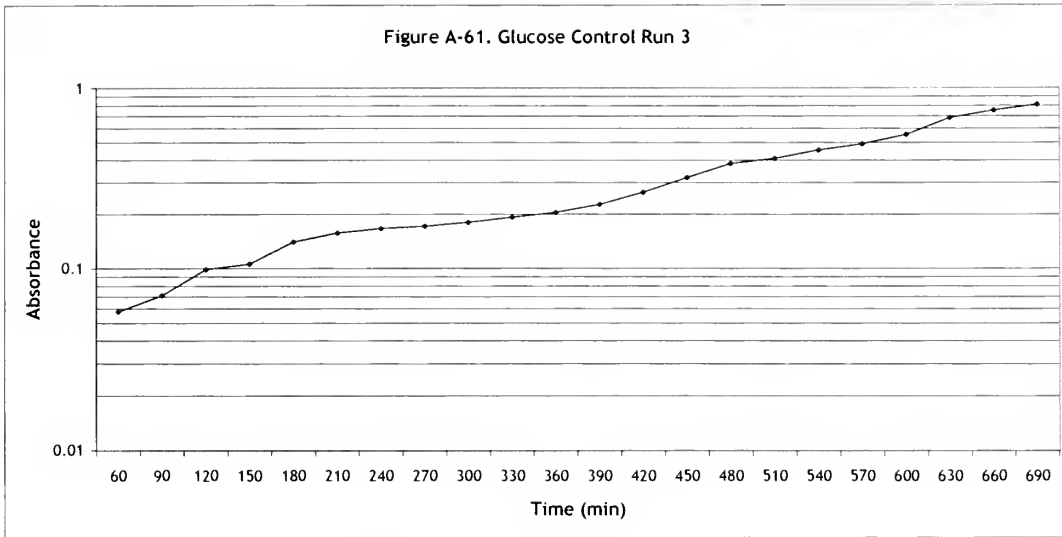


Absorbance increases from 0.102 to .0.204
 Actual growth rate = (270 min - 150 min) = 120 min

Figure A-60. Glucose Control Run 2

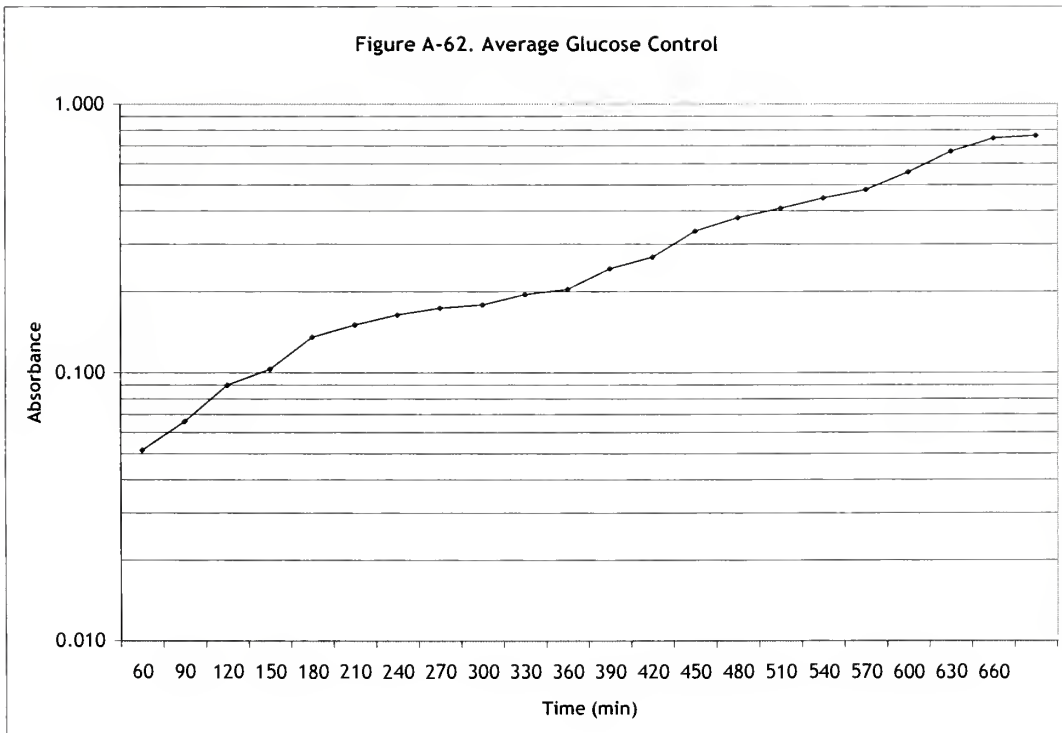


Absorbance increases from 0.205 to .0.407
 Actual growth rate = (540 min - 390 min) = 150 min



Absorbance increases from 0.205 to .0.41

Actual growth rate = (510 min - 360 min) = 150 min



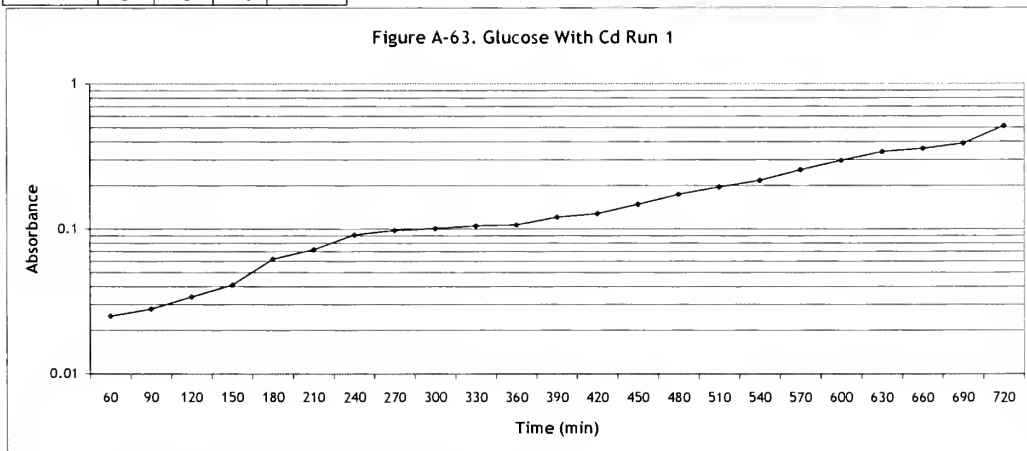
Absorbance increases from 0.103 to .0.204

Actual growth rate = (510 min - 360 min) = 150 min

Data for *E. coli* from absorbance vs. time in M9 media with no casamino acids (poor media)

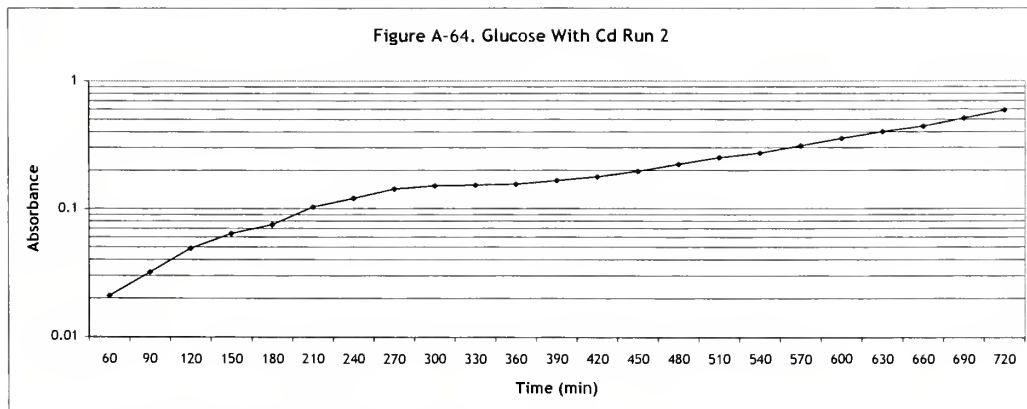
Glucose With 1×10^{-5} M Cd				
Time(min)	Run1	Run2	Run3	Average
0				
30				
60	0.025	0.021	0.032	0.023
90	0.028	0.032	0.038	0.030
120	0.034	0.049	0.042	0.042
150	0.041	0.064	0.059	0.053
180	0.062	0.075	0.079	0.069
210	0.072	0.103	0.095	0.088
240	0.091	0.121	0.108	0.106
270	0.098	0.143	0.126	0.121
300	0.101	0.151	0.142	0.126
330	0.105	0.153	0.148	0.129
360	0.107	0.156	0.153	0.132
390	0.121	0.166	0.165	0.144
420	0.128	0.178	0.174	0.153
450	0.148	0.196	0.187	0.172
480	0.174	0.223	0.201	0.199
510	0.195	0.251	0.224	0.223
540	0.217	0.273	0.251	0.247
570	0.256	0.312	0.292	0.287
600	0.298	0.356	0.328	0.327
630	0.342	0.402	0.364	0.369
660	0.359	0.443	0.4	0.401
690	0.391	0.511	0.498	0.467
720	0.514	0.59	0.569	0.558

Figure A-63. Glucose With Cd Run 1

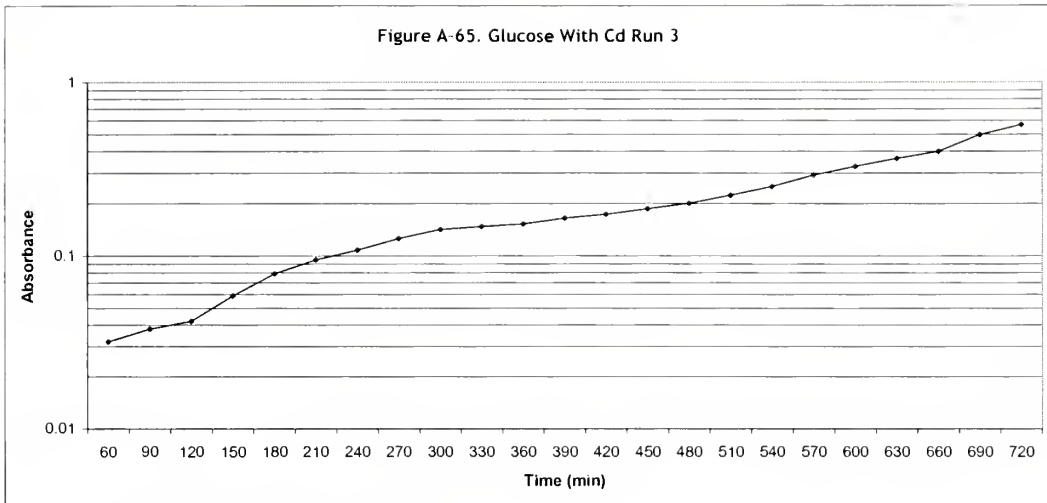


Absorbance increases from 0.195 to 0.387
 Actual growth rate = (690 min - 510min) = 180 min

Figure A-64. Glucose With Cd Run 2

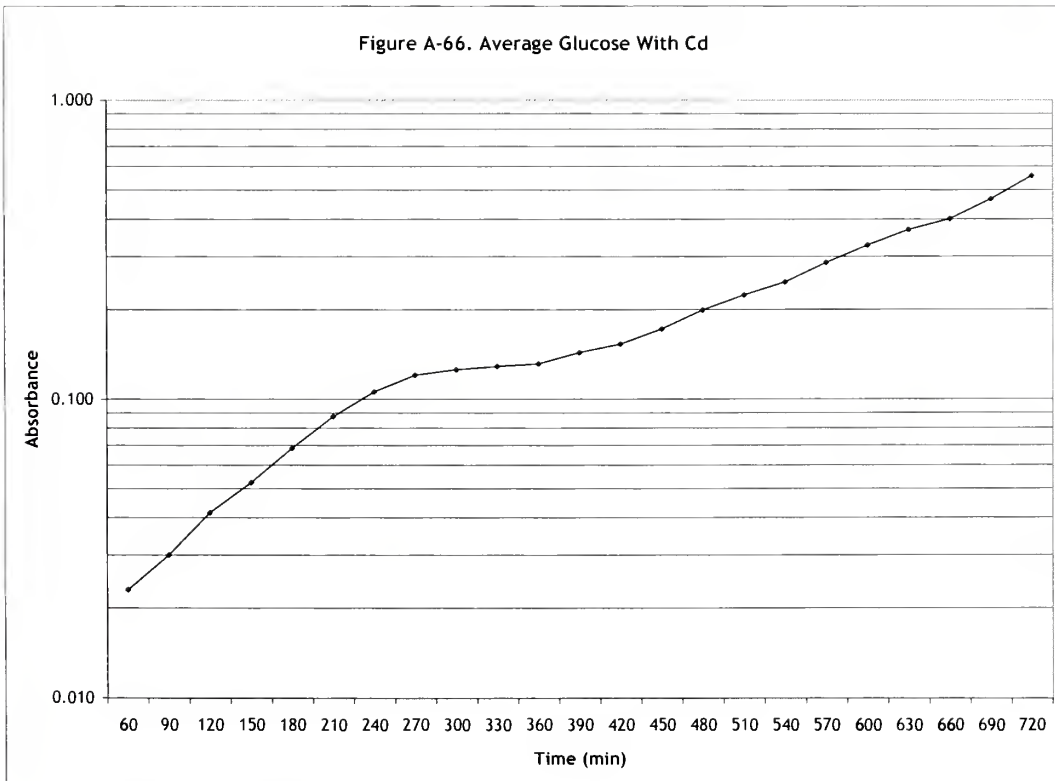


Absorbance increases from 0.223 to 0.443
 Actual growth rate = (660 min - 480min) = 180 min



Absorbance increases from 0.201 to .0.4

Actual growth rate = (660 min - 480min) = 180 min

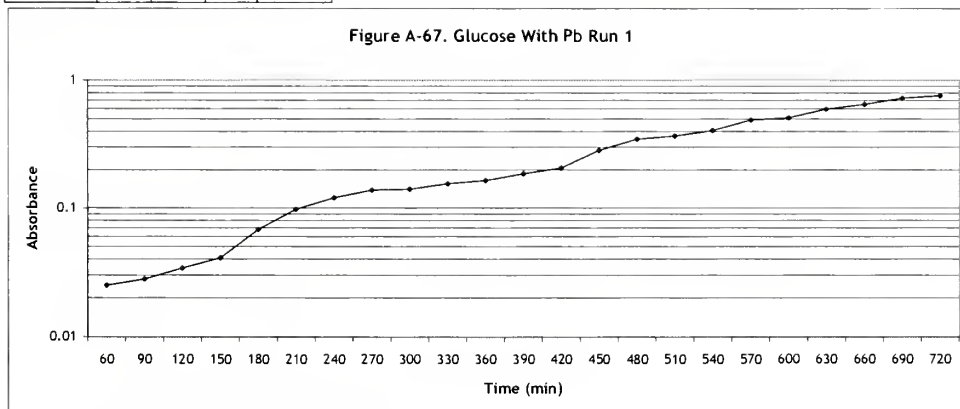


Absorbance increases from 0.199 to .0.401

Actual growth rate = (660 min - 480min) = 180 min

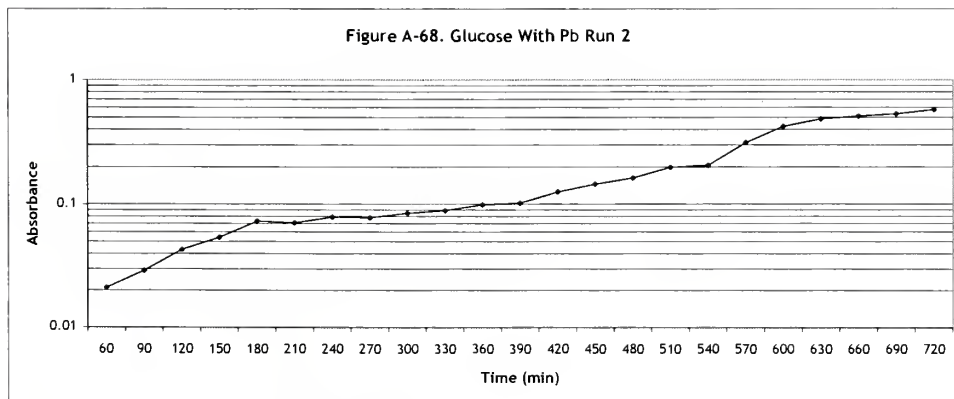
Data for *E.coli* from absorbance vs. time in M9 media with no casamino acids (poor media)

Glucose With 1×10^{-5} M Pb				
Time(min)	Run1	Run2	Run3	Average
60	0.025	0.021	0.032	0.026
90	0.028	0.029	0.033	0.030
120	0.034	0.043	0.036	0.038
150	0.041	0.054	0.048	0.048
180	0.068	0.073	0.071	0.071
210	0.098	0.071	0.099	0.089
240	0.121	0.079	0.102	0.101
270	0.139	0.078	0.114	0.110
300	0.141	0.085	0.121	0.116
330	0.156	0.089	0.128	0.124
360	0.165	0.099	0.131	0.132
390	0.186	0.102	0.148	0.145
420	0.207	0.126	0.163	0.165
450	0.283	0.145	0.174	0.201
480	0.347	0.163	0.196	0.235
510	0.369	0.198	0.227	0.265
540	0.406	0.206	0.294	0.302
570	0.491	0.312	0.362	0.388
600	0.512	0.423	0.454	0.463
630	0.597	0.484	0.512	0.531
660	0.653	0.511	0.582	0.582
690	0.726	0.533	0.631	0.630
720	0.762	0.576	0.694	0.677



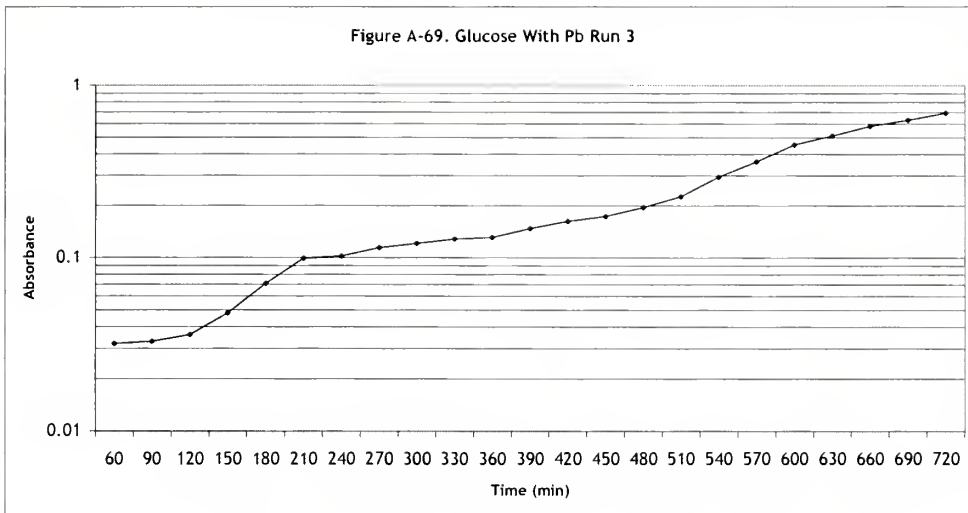
Absorbance increases from 0.141 to .0.283

Actual growth rate = (450 min - 300min) = 150 min



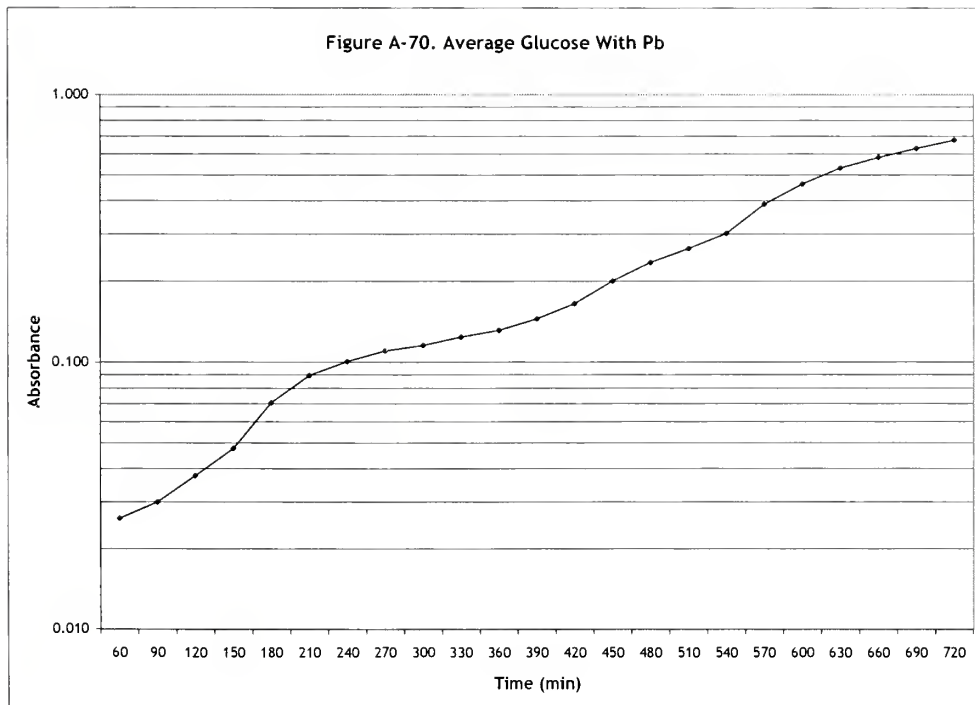
Absorbance increases from 0.102 to .0.206

Actual growth rate = (540 min - 390min) = 150 min



Absorbance increases from 0.148 to .0.294

Actual growth rate = (540 min - 390min) = 150 min



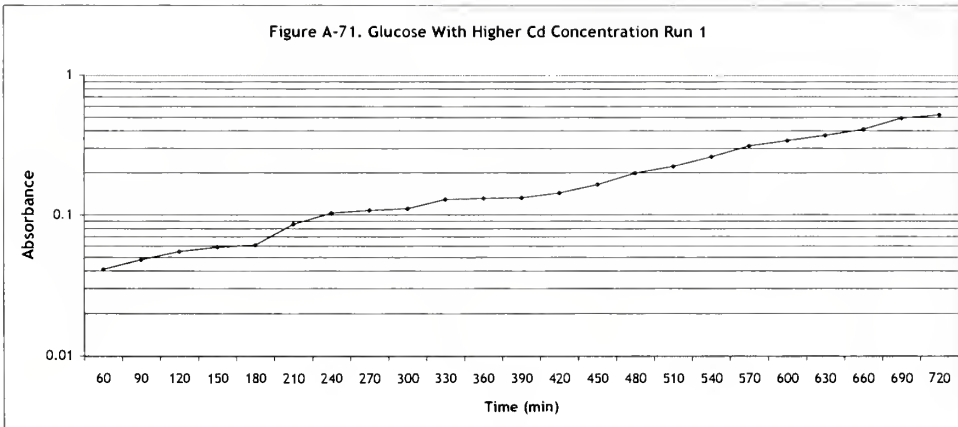
Absorbance increases from 0.132 to .0.265

Actual growth rate = (510 min - 360min) = 150 min

Data for *E. coli* from absorbance vs. time in M9 media with no casamino acids (poor media)

Glucose With $1 \cdot 10^{-3}$ M Cd				
Time(min)	Run1	Run2	Run3	Average
60	0.041	0.053	0.051	0.048
90	0.048	0.068	0.056	0.057
120	0.055	0.073	0.069	0.066
150	0.059	0.112	0.075	0.082
180	0.061	0.131	0.098	0.097
210	0.086	0.128	0.102	0.105
240	0.103	0.135	0.108	0.115
270	0.108	0.131	0.114	0.118
300	0.111	0.135	0.121	0.122
330	0.129	0.142	0.128	0.133
360	0.131	0.149	0.136	0.139
390	0.133	0.151	0.149	0.144
420	0.144	0.155	0.153	0.151
450	0.165	0.168	0.162	0.165
480	0.199	0.175	0.171	0.182
510	0.223	0.184	0.213	0.207
540	0.261	0.193	0.243	0.232
570	0.312	0.248	0.258	0.273
600	0.342	0.301	0.306	0.316
630	0.373	0.342	0.351	0.355
660	0.412	0.381	0.396	0.396
690	0.495	0.438	0.449	0.461
720	0.522	0.489	0.482	0.498

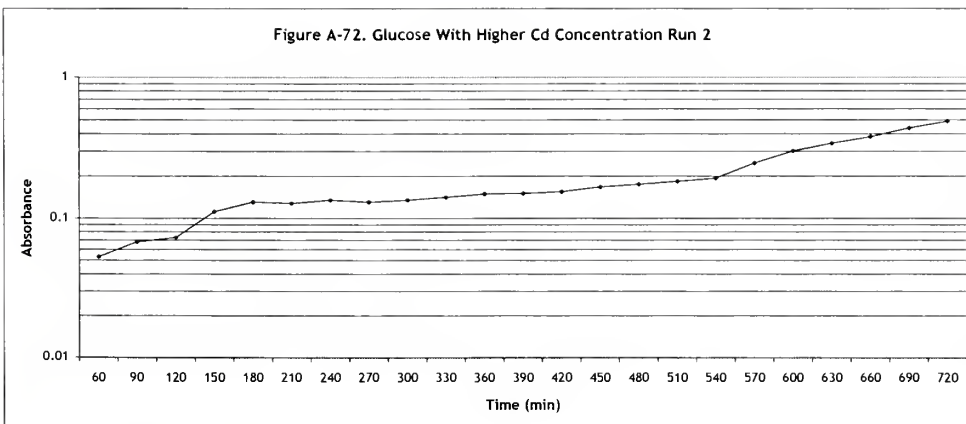
Figure A-71. Glucose With Higher Cd Concentration Run 1



Absorbance increases from 0.129 to .0.261

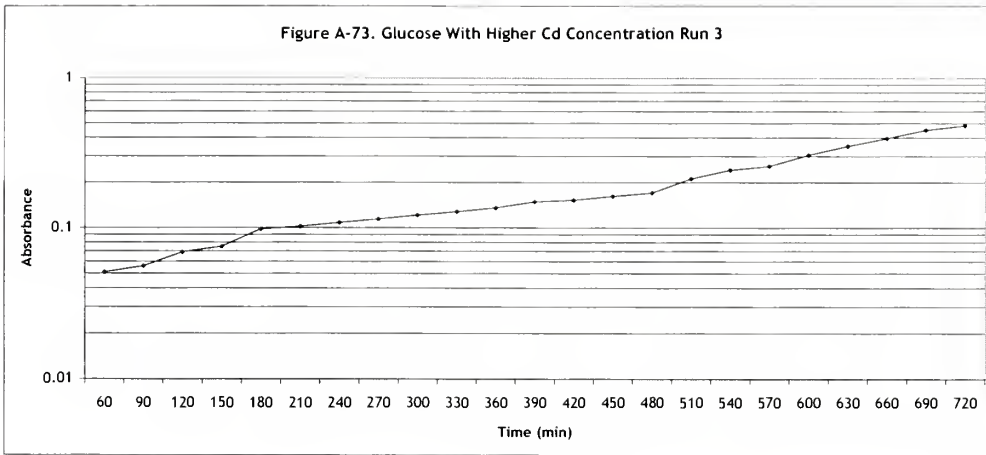
Actual growth rate = (540 min - 330min) = 210 min

Figure A-72. Glucose With Higher Cd Concentration Run 2



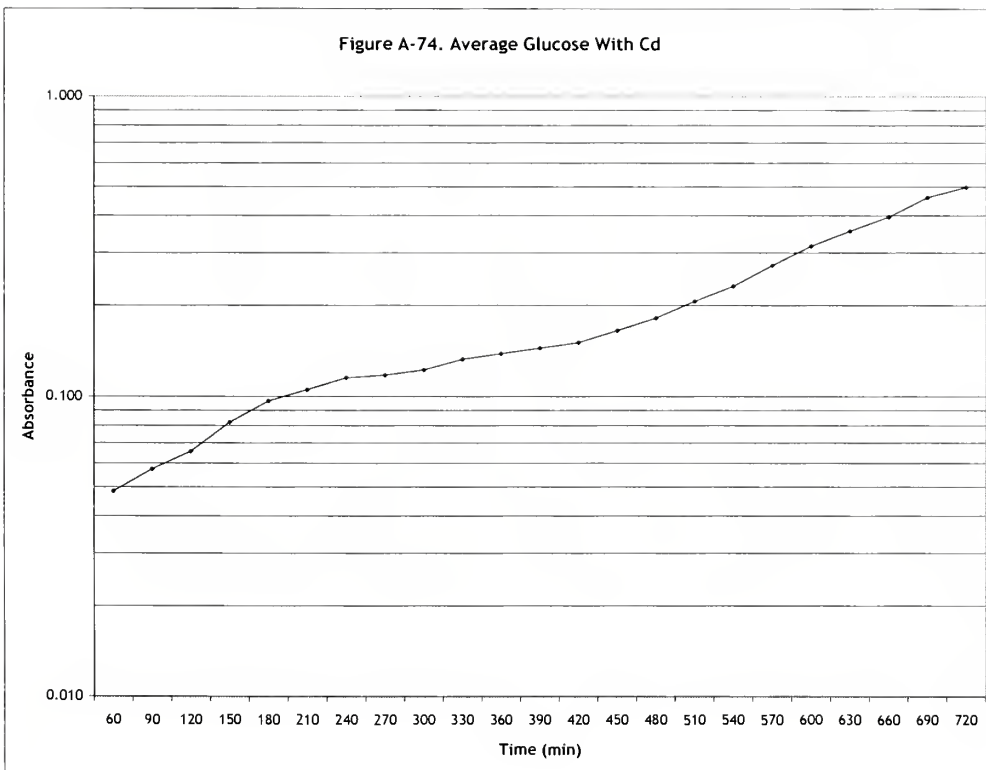
Absorbance increases from 0.151 to .0.301

Actual growth rate = (600 min - 390min) = 210 min



Absorbance increases from 0.128 to 0.258

Actual growth rate = (570 min - 330min) = 240 min

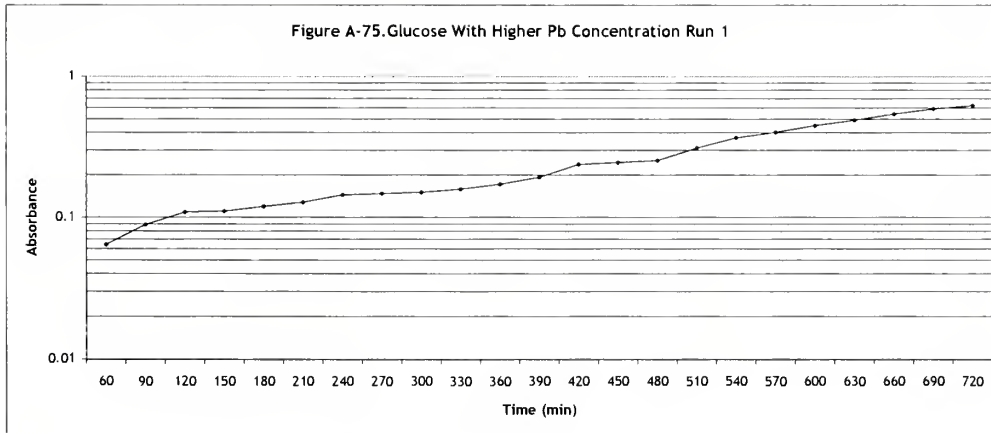


Absorbance increases from 0.139 to .0.280

Actual growth rate = (570 min - 360min) = 210 min

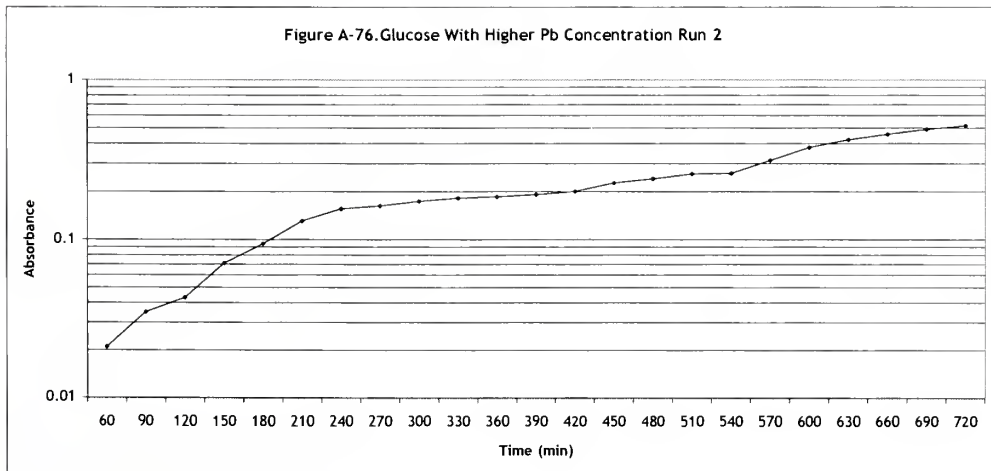
Data for *E. coli* from absorbance vs. time in M9 media with no casamino acids (poor media)

Glucose With 1×10^{-3} M Pb				
Time(min)	Run1	Run2	Run3	Average
60	0.064	0.021	0.041	0.042
90	0.089	0.035	0.048	0.057
120	0.109	0.043	0.055	0.069
150	0.111	0.071	0.086	0.089
180	0.120	0.094	0.103	0.106
210	0.128	0.131	0.111	0.123
240	0.145	0.156	0.121	0.141
270	0.148	0.163	0.142	0.151
300	0.151	0.174	0.158	0.161
330	0.159	0.182	0.163	0.168
360	0.172	0.186	0.178	0.179
390	0.193	0.192	0.188	0.191
420	0.238	0.201	0.211	0.217
450	0.246	0.227	0.233	0.235
480	0.254	0.241	0.248	0.248
510	0.312	0.258	0.273	0.281
540	0.368	0.261	0.341	0.323
570	0.403	0.314	0.359	0.359
600	0.449	0.378	0.377	0.401
630	0.493	0.422	0.412	0.442
660	0.544	0.458	0.495	0.499
690	0.592	0.491	0.522	0.535
720	0.625	0.516	0.543	0.561



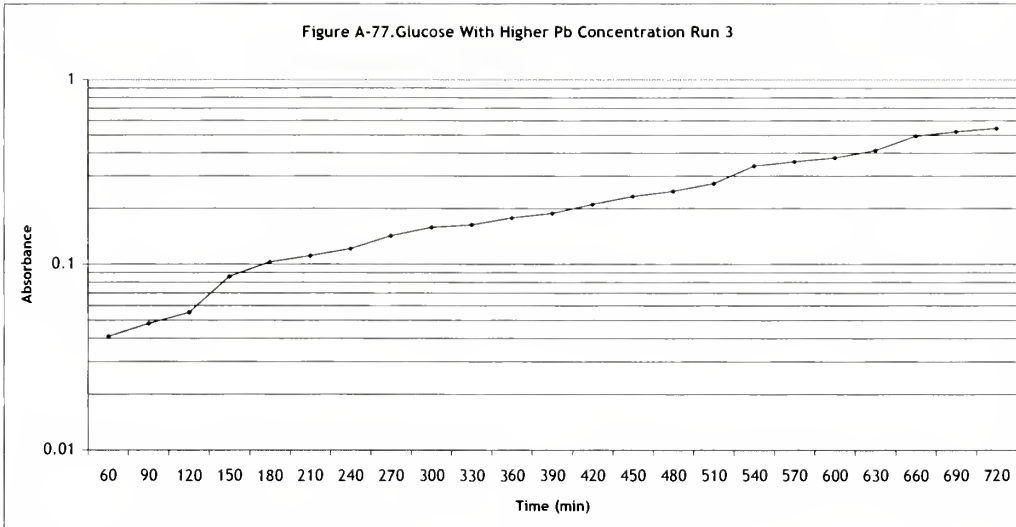
Absorbance increases from 0.179 to .0359

Actual growth rate = (570 min - 360min) = 210 min



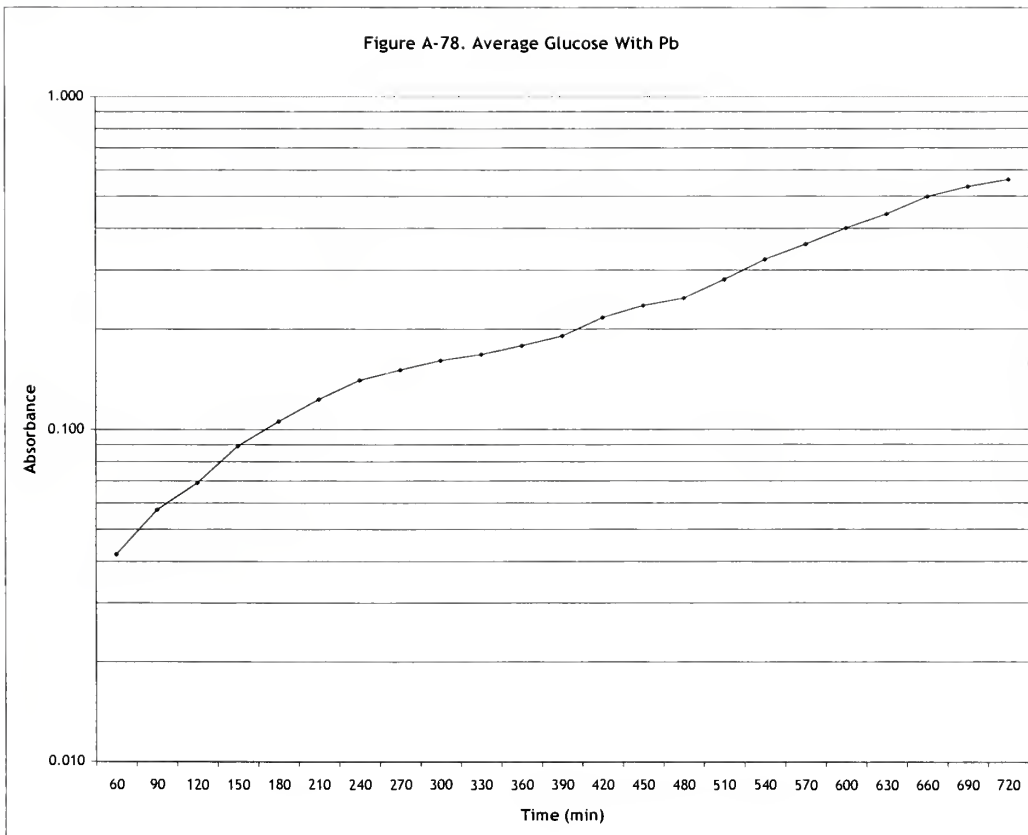
Absorbance increases from 0.179 to .0359

Actual growth rate = (570 min - 360min) = 210 min



Absorbance increases from 0.188 to .0.377

Actual growth rate = (600 min - 390min) = 210 min



Absorbance increases from 0.179 to .0.359

Actual growth rate = (570 min - 360min) = 210 min

E. coli growth data
Table A-1. E. coli cell growth over period of time

Dilution	1:25	1:50
	(twice for all media)	
	25	50
	6	183
	4	167
	5	175
	5	208
	4	133
	8	200
	Average	
	15	292
	7	225
	8	233
	12	267
	4	200
	11	258

growth time: 6 hours

Dilution	1:25	1:250
	(twice for all media)	
	25	250
	15	1
	7	225
	8	1
	12	1
	4	1
	11	1
	Average	
	25	250
	36	3
	12	1
	17	2
	31	2
	5	1
	13	1

growth time: 7 hours

Dilution	1:25	1:250
	(twice la., la.+Cd, la.+Pb, glu+Pb)	
	25	250
	36	3
	12	1
	17	2
	31	2
	5	1
	13	1
	Average	
	25	250
	36	3
	12	1
	17	2
	31	2
	5	1
	13	1

growth time: 8 hours

Dilution	1:25	1:250
	(twice la.+Cd, glu+Pb)	
	25	250
	49	5
	18	2
	19	2
	35	3
	6	1
	16	2
	Average	
	25	250
	56	5
	37	3
	21	2
	41	4
	7	1
	17	2

growth time: 9 hours

Dilution	1:25	1:250
	(twice la., la.+Cd, glu)	
	25	250
	56	5
	37	3
	21	2
	41	4
	7	1
	17	2
	Average	
	25	250
	56	5
	37	3
	21	2
	41	4
	7	1
	17	2

growth time: 11 hours

	Dilution		Average
	1:50 (twice la-Cd, glu-Cd, glu+Pb)	1:2,500 (twice la-Cd, glu-Cd, glu+Pb)	
lactose	50	2,500	5233
lactose and Cd	104	2	1033
lactose and Pb	21	0	2100
glucose	26	0	1283
glucose and Cd	27	0	533
glucose and Pb	11	0	733
	17	0	

growth time: 10 hours

	Dilution		Average
	1:50 (twice glu, glu-Cd, glu+Pb)	1:2,500 (twice la-Cd, la, +Pb)	
lactose	84	1	3400
lactose and Cd	17	0	950
lactose and Pb	22	0	1033
glucose	26	0	1200
glucose and Cd	9	0	467
glucose and Pb	13	0	600

growth time: 12.5 hours

	Dilution		Average
	1:50 twice (glu+Pb)	1:2,500 (twice la-Cd, glu, glu+Cd)	
lactose	321	8	18150
lactose and Cd	50	2	1583
lactose and Pb	25	1	3067
glucose	64	0	2050
glucose and Cd	33	0	950
glucose and Pb	17	0	1000
	20	0	

growth time: 12 hours

	Dilution		Average
	1:50 (twice glu, glu-Cd, glu+Pb)	1:2,500 (twice la-Cd, la, +Pb)	
lactose	171	4	9350
lactose and Cd	24	0	1133
lactose and Pb	44	1	2567
glucose	31	0	1567
glucose and Cd	17	0	733
glucose and Pb	18	0	767

growth time: 14 hours

	Dilution		Average
	1:50 (twice glu+Pb)	1:2,500 (twice glu+Cd)	
lactose	50	44	106333
lactose and Cd	100-	218	2467
lactose and Pb	48	5	12033
glucose	242	23	3767
glucose and Cd	66	6	1933
glucose and Pb	36	4	2667
	50	6	

growth time: 13 hours

	Dilution		Average
	1:50 (twice glu+Pb)	1:2,500 (twice la-Cd, glu, glu+Cd)	
lactose	341	9	19683
lactose and Cd	25	3	1750
lactose and Pb	69	8	4150
glucose	41	0	2517
glucose and Cd	21	0	1350
glucose and Pb	21	0	1033

growth time: 16 hours

	Dilution				Average
	1:50	1:2,500	1:25,000	1:125,000	
	50	500	25,000	125,000	
		twice(glu, glu+Cd)			
lactose	100+	100+	224	22	5
lactose and Cd	165	16	4	0	8750
lactose and Pb	100+	294	64	6	152333
glucose	100+	46	9	0	22667
glucose and Cd	96	10	0	0	4933
glucose and Pb	113	14	2	0	5883

growth time: 15 hours

	Dilution				Average
	1:50	1:2,500	1:25,000	1:125,000	
	50	500	25,000	125,000	
		twice(glu+Cd)			
lactose	100+	174	13	3	316667
lactose and Cd	82	2	0	0	4367
lactose and Pb	100+	48	1	0	23000
glucose	183	4	0	0	9383
glucose and Cd	68	2	0	0	4133
glucose and Pb	78	2	0	0	4467

growth time: 18 hours

	Dilution				Average
	1:50	1:2,500	1:25,000	1:125,000	
	50	500	25,000	125,000	
		twice(glu+Cd)			
lactose	100+	256	27	0	663333
lactose and Cd	264	26	6	0	13733
lactose and Pb	100+	175	16	0	425000
glucose	100+	258	53	5	128833
glucose and Cd	114	10	2	0	5400
glucose and Pb	149	17	3	0	7817

growth time: 17 hours

	Dilution				Average
	1:50	1:2,500	1:25,000	1:125,000	
	50	500	25,000	125,000	
		twice(glu+Cd)			
lactose	100+	232	25	5	610000
lactose and Cd	218	6	0	0	12300
lactose and Pb	100+	165	16	3	395833
glucose	100+	132	27	3	69500
glucose and Cd	115	10	0	0	5250
glucose and Pb	143	16	3	0	7550

growth time: 19.5 hours

	Dilution				Average
	1:2,500	1:25,000	1:125,000	1:125,000	
		twice(la+Cd)			
3 times(glu+Cd, glu+Pb)	2,500	25,000	125,000	125,000	
lactose	100+	29	6	0	733333
lactose and Cd	8	1	0	0	23333
lactose and Pb	181	18	4	0	467500
glucose	52	5	1	0	126667
glucose and Cd	2, 3-twice	0	0	0	6667
glucose and Pb	4, 3-twice	0	0	0	8333

growth time: 19 hours

	Dilution				Average
	1:2,500	1:25,000	1:125,000	1:125,000	
		twice(la+Cd, glu+Pb)			
3 times(la+Cd, glu+Pb)	2,500	25,000	125,000	125,000	
lactose	282	28	6	0	718333
lactose and Cd	7	0	0	0	17500
lactose and Pb	172	16	3	0	401667
glucose	51	4	1	0	117500
glucose and Cd	2-twice, 3	0	0	0	5833
glucose and Pb	3	0	0	0	7500

growth time:		21 hours	
Dilution		1:2,500	1:125,000
3 times(glu +Cd, glu +Pb)	twice(la+Cd)	25,000	125,000
Average			
lactose		330	7
lactose and Cd		11	0
lactose and Pb		199	4
glucose		105	2
glucose and Cd		3	0
glucose and Pb		4	0

growth time:		20 hours	
Dilution		1:2,500	1:125,000
3 times(glu +Cd, glu +Pb)	twice(la, la+Cd)	25,000	125,000
Average			
lactose		100+	6
lactose and Cd		9	0
lactose and Pb		192	4
glucose		98	2
glucose and Cd		3	0
glucose and Pb		4	0

growth time:		23 hours	
Dilution		1:2,500	1:125,000
3 times(glu +Pb)	twice(la, la+Cd)	25,000	125,000
Average			
lactose		100+	8
lactose and Cd		14	0
lactose and Pb		201	4
glucose		123	3
glucose and Cd	4, 5-twice	0	0
glucose and Pb		6	0

growth time:		22 hours	
Dilution		1:2,500	1:125,000
3 times (glu +Pb)	twice(la, la+Cd)	25,000	125,000
Average			
lactose		100+	8
lactose and Cd		15	0
lactose and Pb		198	4
glucose		119	2
glucose and Cd	3-twice,	0	0
glucose and Pb		5	0

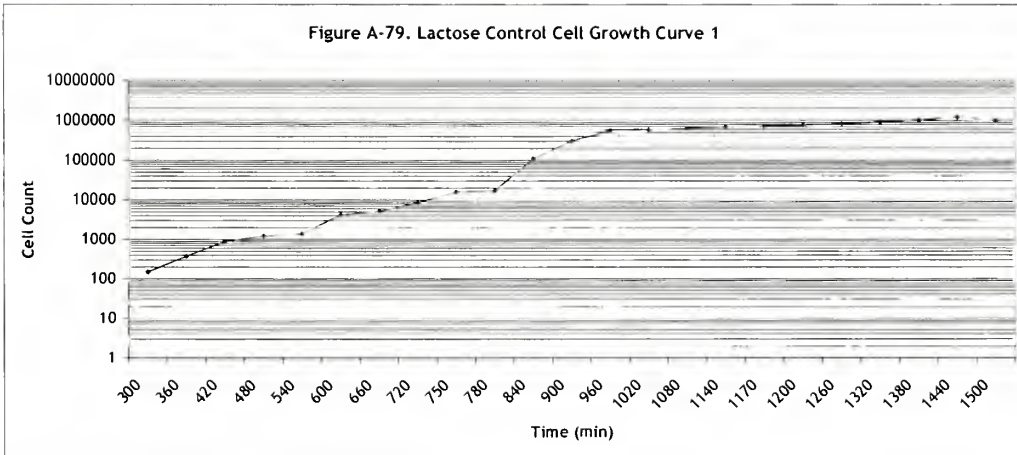
growth time:		25 hours	
Dilution		1:2,500	1:125,000
twice(la+Cd), 3 times(glu +Cd, glu +Pb)	twice(la, la+Cd)	25,000	125,000
Average			
lactose		100+	8
lactose and Cd		15	0
lactose and Pb		192	4
glucose		118	2
glucose and Cd	5	0	0
glucose and Pb	5	0	0

growth time:		24 hours	
Dilution		1:2,500	1:125,000
3 times(glu +Pb)	twice(la, la+Cd)	25,000	125,000
Average			
lactose		100+	9
lactose and Cd		18	0
lactose and Pb		214	5
glucose		141	3
glucose and Cd	4, 6-twice	0	0
glucose and Pb	7	0	0

LACTOSE ONLY CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Average
300	150	200	200	183
360	375	250	250	292
420	900	900	750	850
480	1225	1250	1250	1242
540	1400	1400	1250	1350
600	4200	3500	2500	3400
660	5200	5500	5000	5233
720	8550	9500	10000	9350
780	16050	19000	20000	18350
840	17050	19500	22500	19683
840	109000	110000	100000	106333
900	310000	325000	375000	336667
960	560000	550000	625000	578333
1020	580000	625000	625000	610000
1080	640000	675000	675000	663333
1140	705000	700000	750000	718333
1170	725000	725000	750000	733333
1200	775000	775000	750000	766667
1260	825000	775000	875000	825000
1320	925000	1000000	1000000	975000
1380	1025000	1025000	1000000	1016667
1440	1200000	1200000	1125000	1175000
1500	975000	1000000	1000000	991667

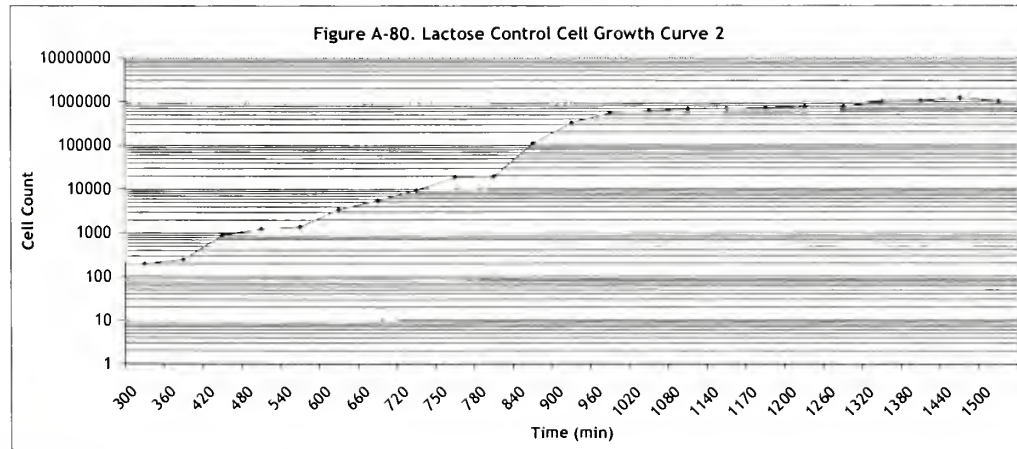
Figure A-79. Lactose Control Cell Growth Curve 1



Cell Count increases from 8550 to 17050

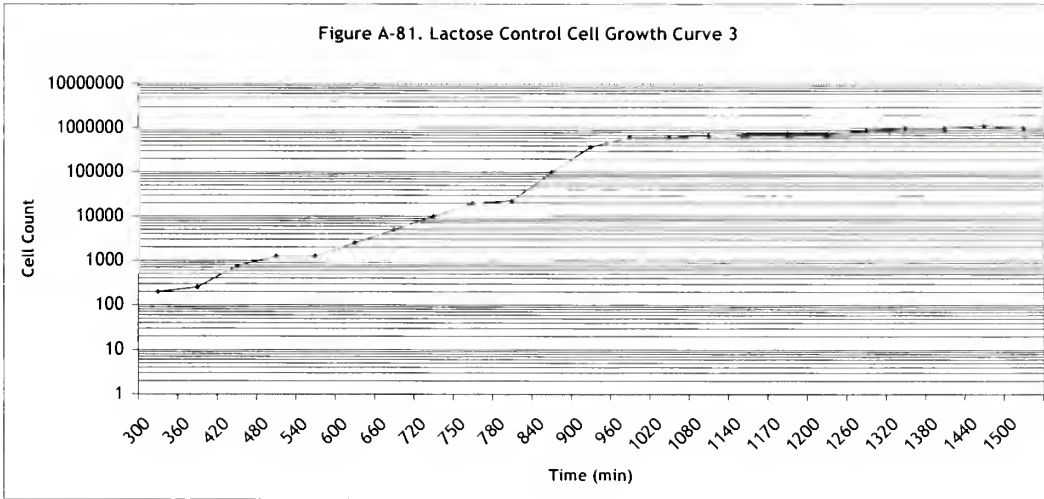
Actual growth rate = (780 min - 720min) = 60 min

Figure A-80. Lactose Control Cell Growth Curve 2



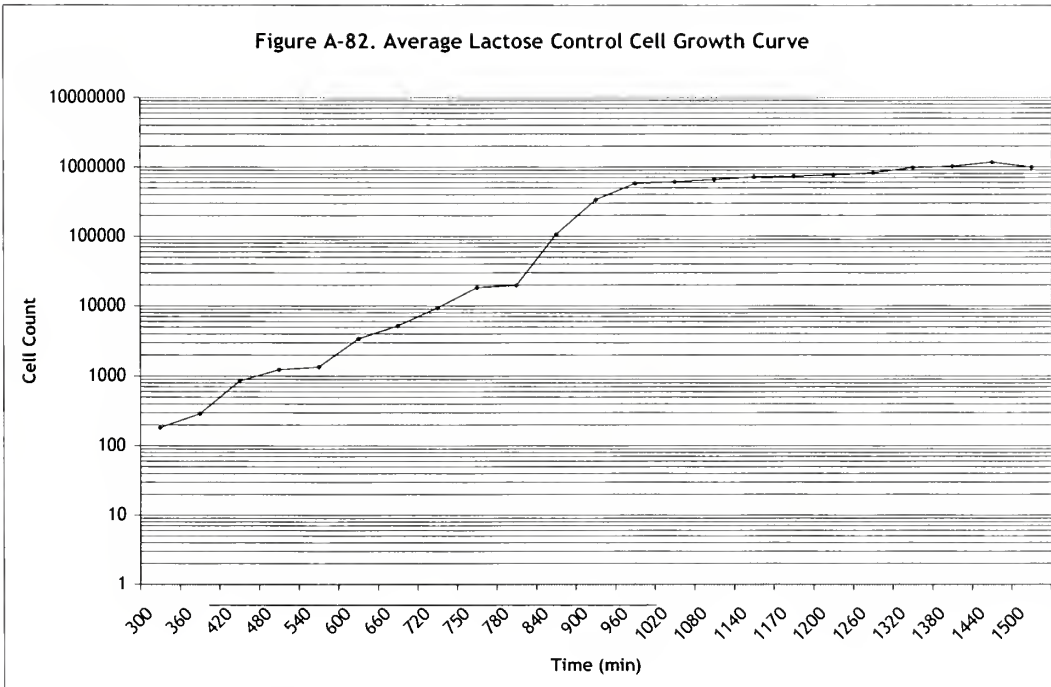
Cell Count increases from 9500 to 19000

Actual growth rate = (750 min - 720min) = 30 min



Cell Count increases from 10000 to 20000

Actual growth rate = (750 min - 720min) = 30 min



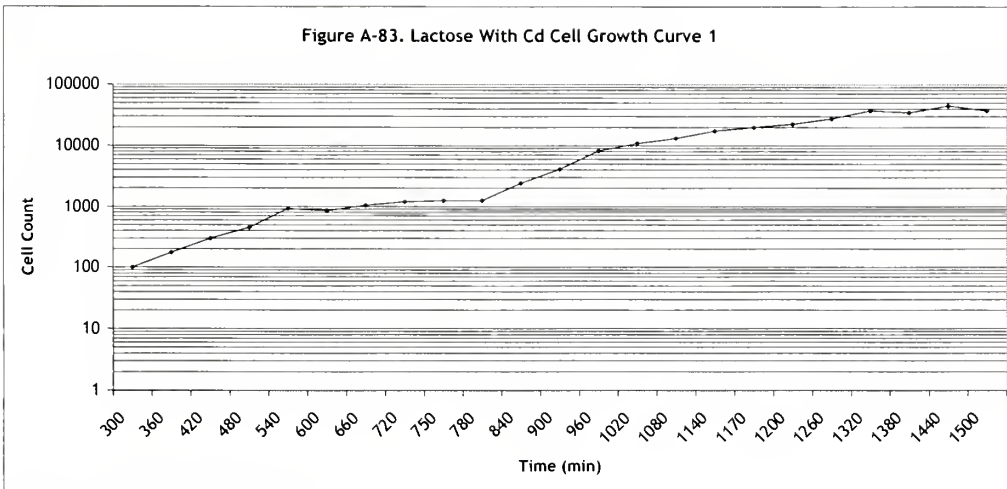
Cell Count increases from 9350 to 18350

Actual growth rate = (750 min - 720min) = 30 min

LACTOSE WITH Cd CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Average
300	100	200	200	167
360	175	250	250	225
420	300	300	250	283
480	450	450	500	467
540	925	925	750	867
600	850	1000	1000	950
660	1050	1050	1000	1033
720	1200	1200	1000	1133
750	1250	1000	2500	1583
780	1250	1500	2500	1750
840	2400	2500	2500	2467
900	4100	5000	4000	4367
960	8250	8000	10000	8750
1020	10900	11000	15000	12300
1080	13200	13000	15000	13733
1140	17500	17500	17500	17500
1170	20000	25000	25000	23333
1200	22500	25000	25000	24167
1260	27500	25000	25000	25833
1320	37500	25000	25000	29167
1380	35000	50000	50000	45000
1440	45000	50000	50000	48333
1500	37500	37500	25000	33333

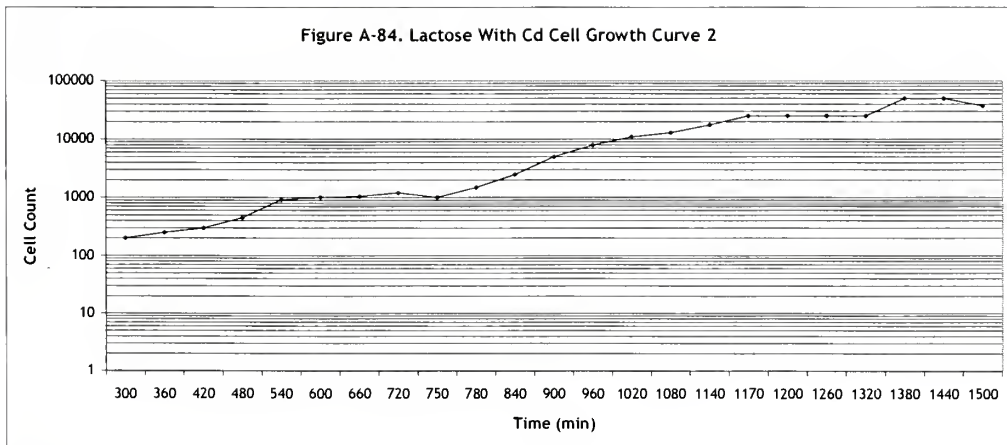
Figure A-83. Lactose With Cd Cell Growth Curve 1



Cell Count increases from 10900 to 22500

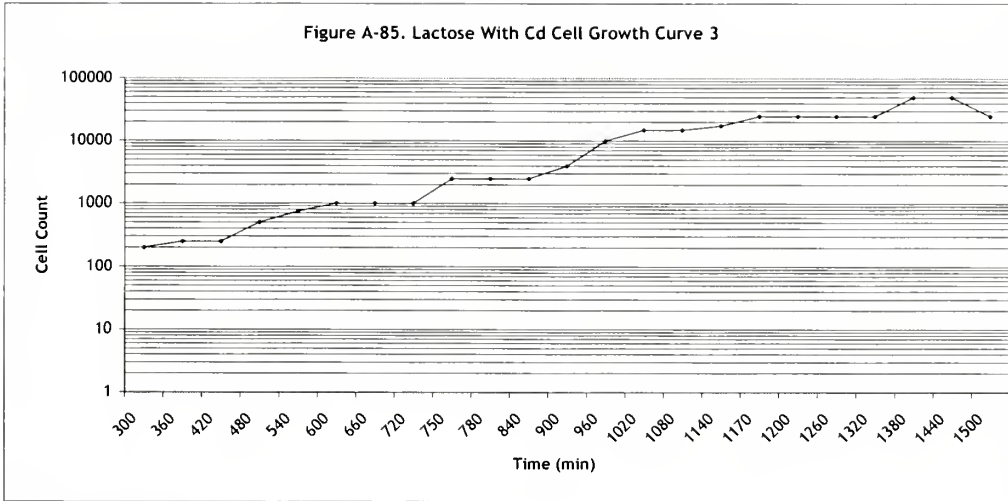
Actual growth rate = (1200 min - 1020min) = 180 min

Figure A-84. Lactose With Cd Cell Growth Curve 2



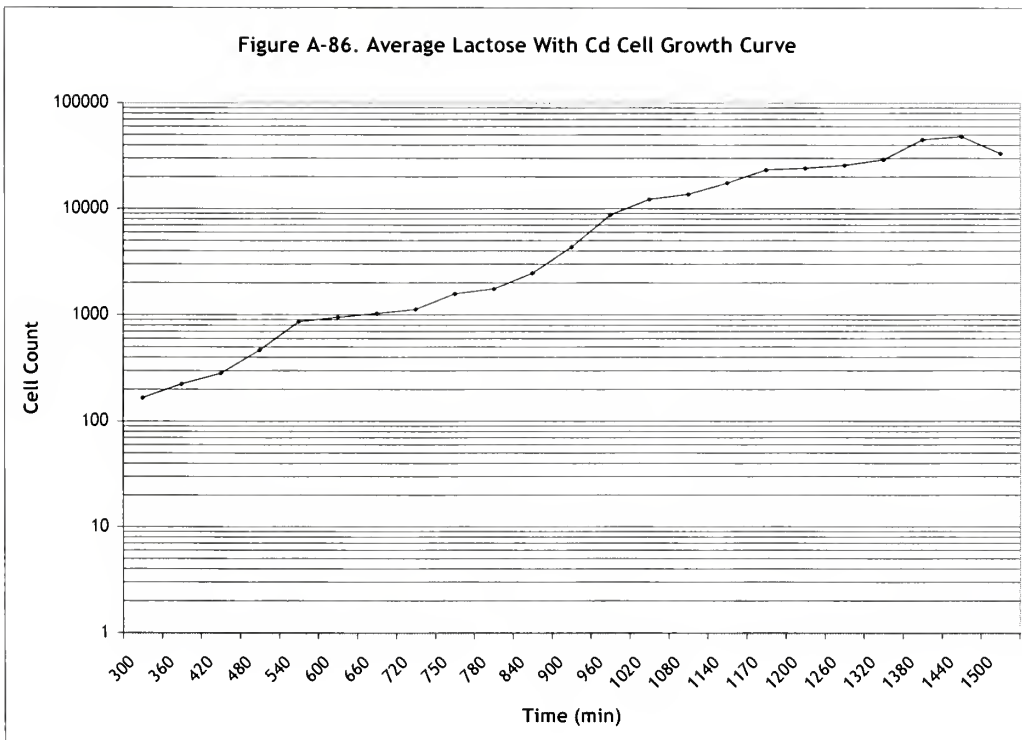
Cell Count increases from 25000 to 50000

Actual growth rate = (1380 min - 1170 min) = 210 min



Cell Count increases from 25000 to 50000

Actual growth rate = (1380 min - 1170 min) = 210 min



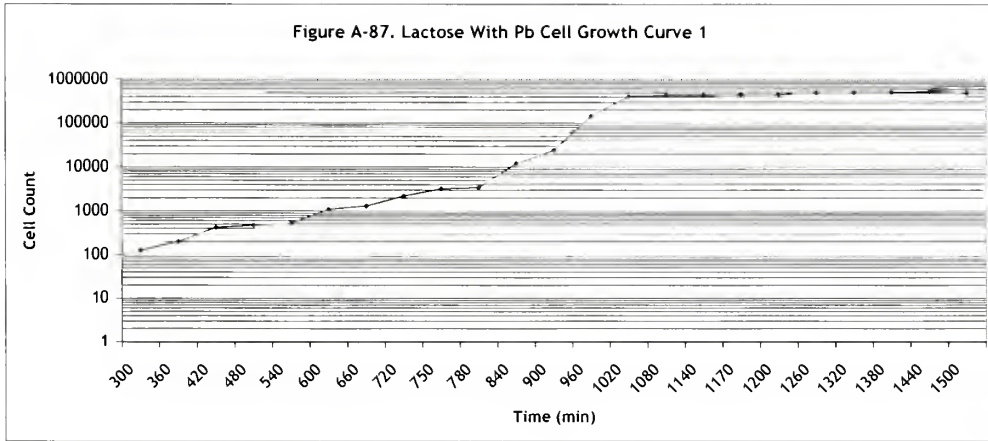
Cell Count increases from 8750 to 17500

Actual growth rate = (1140 min - 960min) = 180 min

LACTOSE WITH Pb CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Average
300	125	200	200	175
360	200	250	250	233
420	425	425	500	450
480	475	500	500	492
540	525	500	500	508
600	1100	1000	1000	1033
660	1300	2500	2500	2100
720	2200	3000	2500	2567
750	3200	3500	2500	3067
780	3450	4000	5000	4150
840	12100	11500	12500	12033
900	24000	20000	25000	23000
960	147000	160000	150000	152333
1020	412500	400000	375000	395833
1080	437500	437500	400000	425000
1140	430000	400000	375000	401667
1170	452500	450000	500000	467500
1200	452500	450000	500000	467500
1260	497500	475000	500000	490833
1320	495000	500000	500000	498333
1380	502500	575000	500000	525833
1440	535000	575000	625000	578333
1500	480000	450000	500000	476667

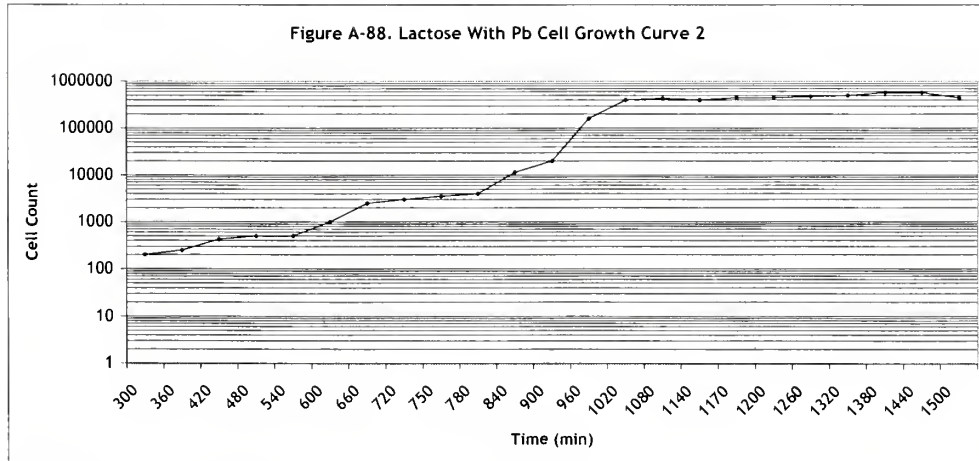
Figure A-87. Lactose With Pb Cell Growth Curve 1



Cell Count increases from 1100 to 2200

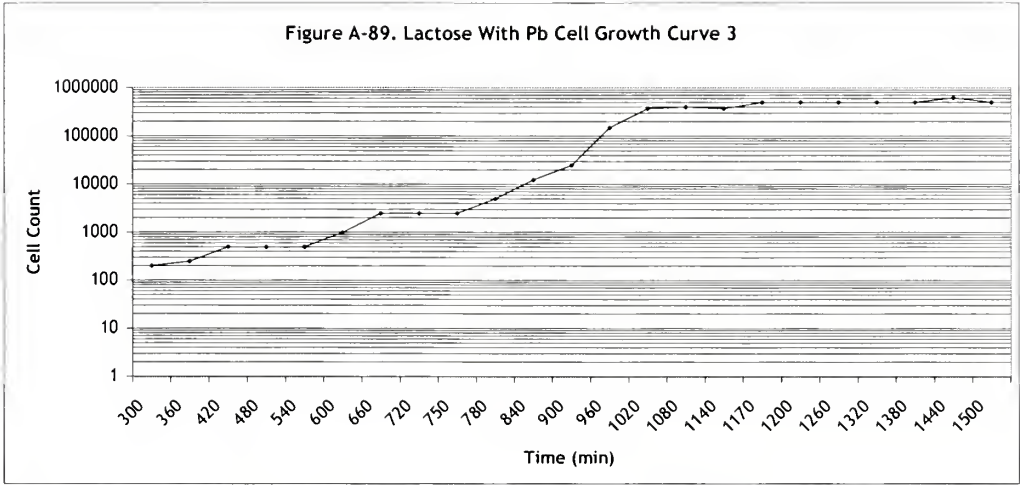
Actual growth rate = (720 min - 600min) = 120 min

Figure A-88. Lactose With Pb Cell Growth Curve 2



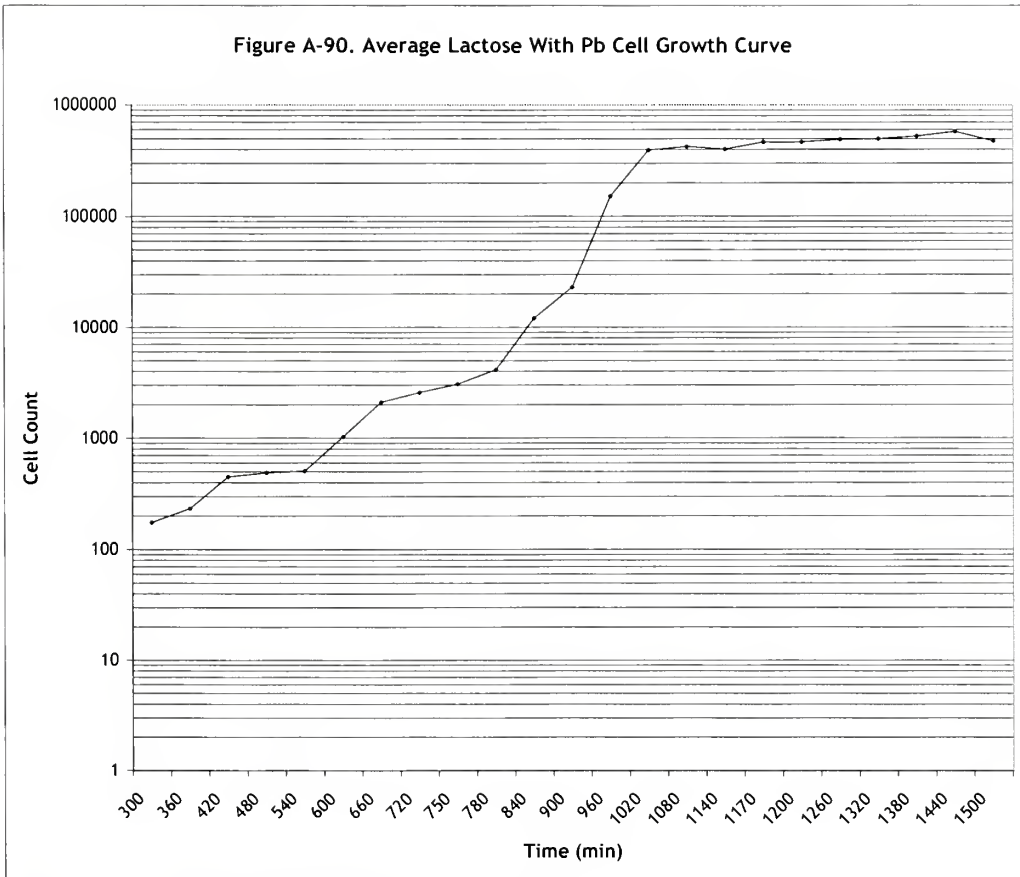
Cell Count increases from 500 to 1000

Actual growth rate = (600 min - 480min) = 120 min



Cell Count increases from 2500 to 5000

Actual growth rate = (780 min - 660 min) = 120 min

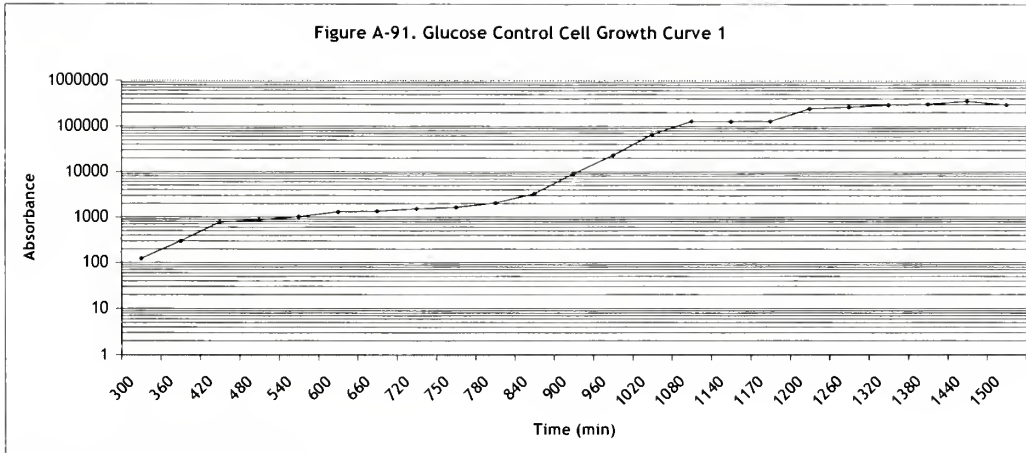


Cell Count increases from 2100 to 4150

Actual growth rate = (780 min - 660min) = 120 min

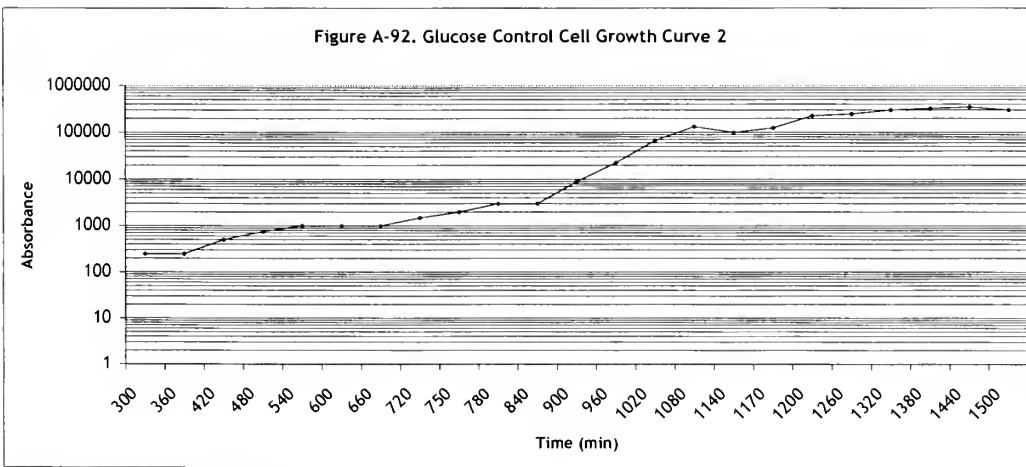
GLUCOSE ONLY CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Average
300	125	250	250	208
360	300	250	250	267
420	775	500	500	592
480	875	750	875	833
540	1025	1000	1025	1017
600	1300	1000	1300	1200
660	1350	1000	1500	1283
720	1550	1500	1650	1567
750	1650	2000	2500	2050
780	2050	3000	2500	2517
840	3300	3000	5000	3767
900	9150	9000	10000	9383
960	23000	22500	22500	22667
1020	66000	67500	75000	69500
1080	129000	132500	125000	128833
1140	127500	100000	125000	117500
1170	130000	125000	125000	126667
1200	245000	225000	250000	240000
1260	262500	250000	250000	254167
1320	297500	300000	250000	282500
1380	307500	325000	375000	335833
1440	352500	350000	375000	359167
1500	295000	300000	250000	281667



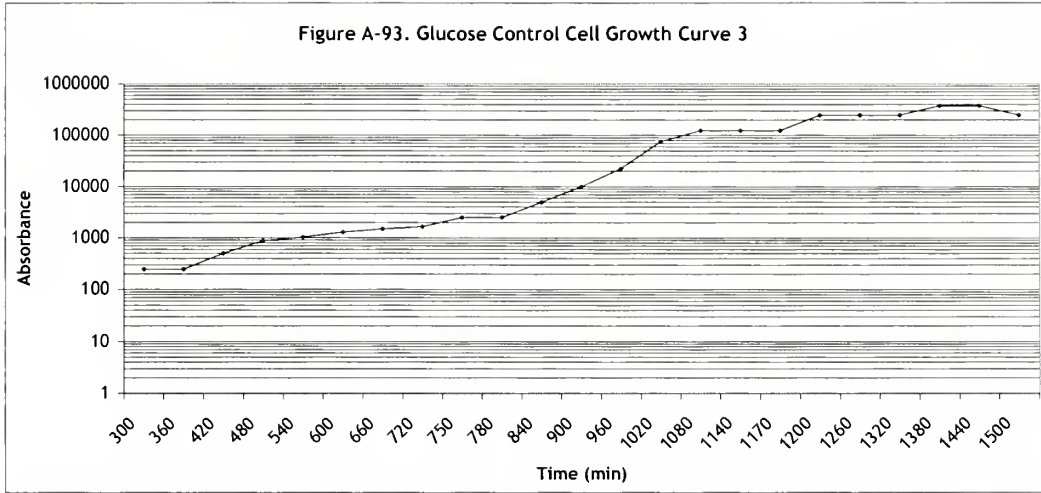
Cell Count increases from 1650 to 3300

Actual growth rate = (840min - 750min) = 90 min



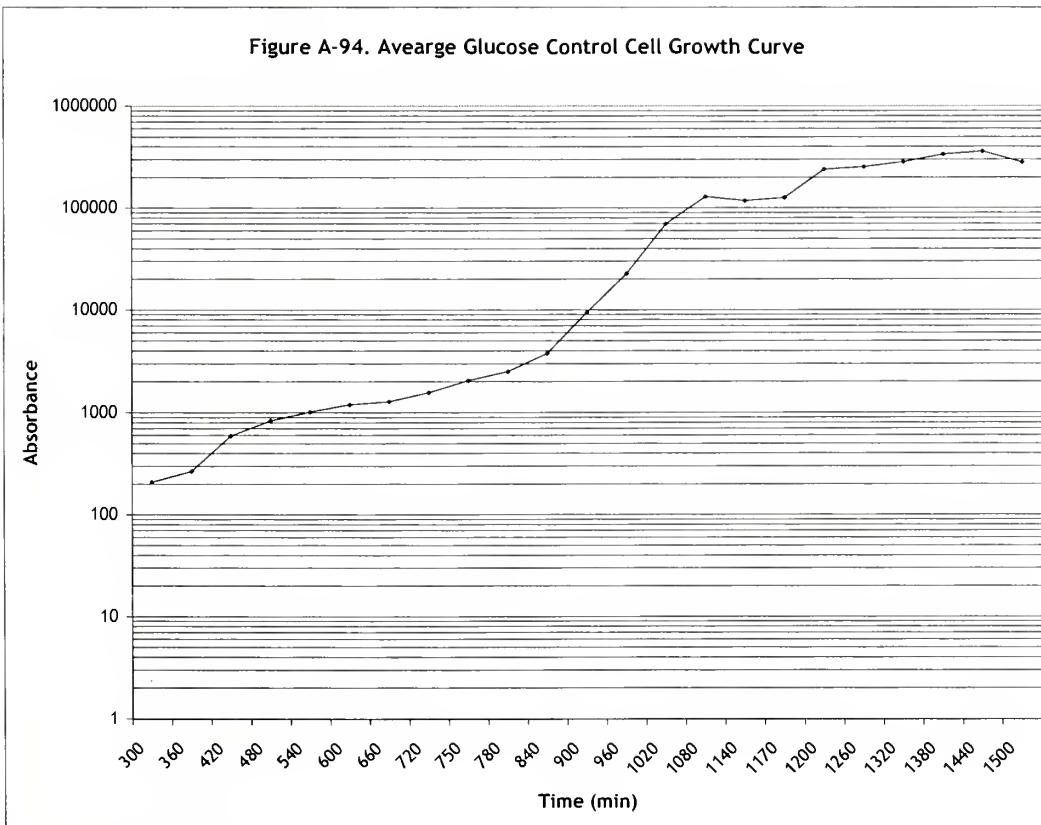
Cell Count increases from 1500 to 3000

Actual growth rate = (780min - 720min) = 60 min



Cell Count increases from 5000 to 10000

Actual growth rate = (900min - 840min) = 60 min

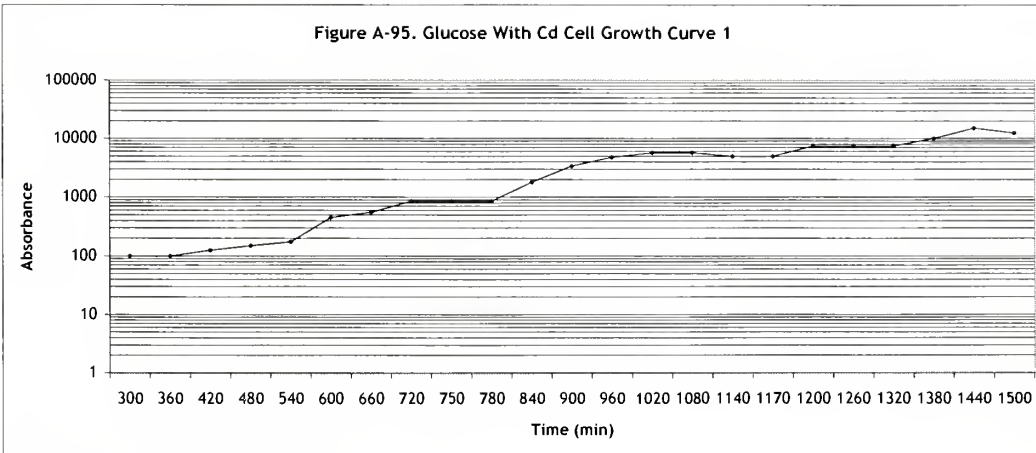


Cell Count increases from 126667 to 254167

Actual growth rate = (1260min - 1170min) = 90 min

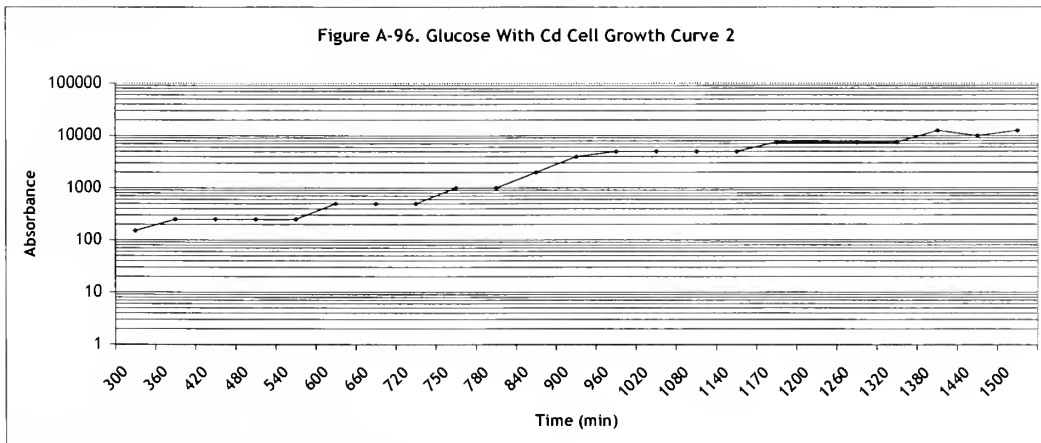
GLUCOSE WITH Cd CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Average
300	100	150	150	133
360	100	250	250	200
420	125	250	250	208
480	150	250	250	217
540	175	250	250	225
600	450	500	450	467
660	550	500	550	533
720	850	500	850	733
750	850	1000	1000	950
780	850	1000	1500	1117
840	1800	2000	2000	1933
900	3400	4000	5000	4133
960	4800	5000	5000	4933
1020	5750	5000	5000	5250
1080	5700	5000	5500	5400
1140	5000	5000	7500	5833
1170	5000	7500	7500	6667
1200	7500	7500	7500	7500
1260	7500	7500	7500	7500
1320	7500	7500	10000	8333
1380	10000	12500	12500	11667
1440	15000	10000	15000	13333
1500	12500	12500	12500	12500



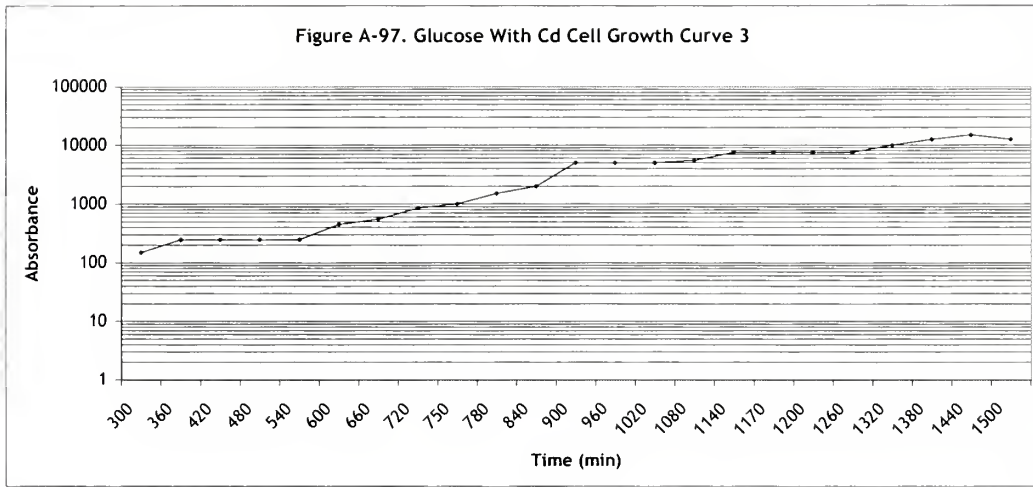
Cell Count increases from 5000 to 10000

Actual growth rate = (1380min - 1170min) = 210 min



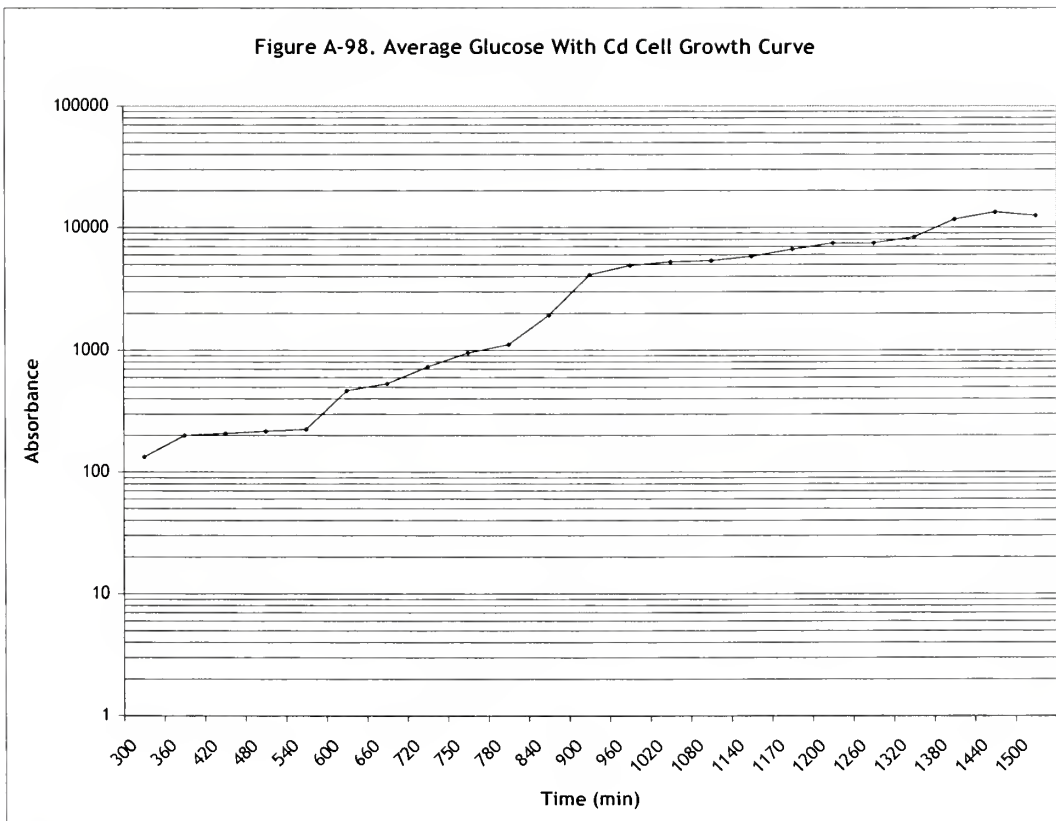
Cell Count increases from 5000 to 10000

Actual growth rate = (1440min - 1140min) = 300 min



Cell Count increases from 5000 to 10000

Actual growth rate = (1320min - 1020min) = 300 min

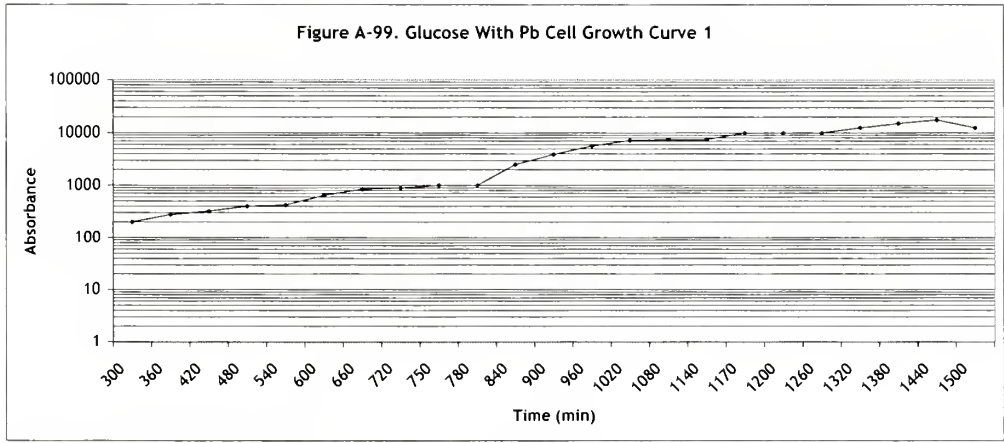


Cell Count increases from 6667 to 13333

Actual growth rate = (1440min - 1170min) = 270 min

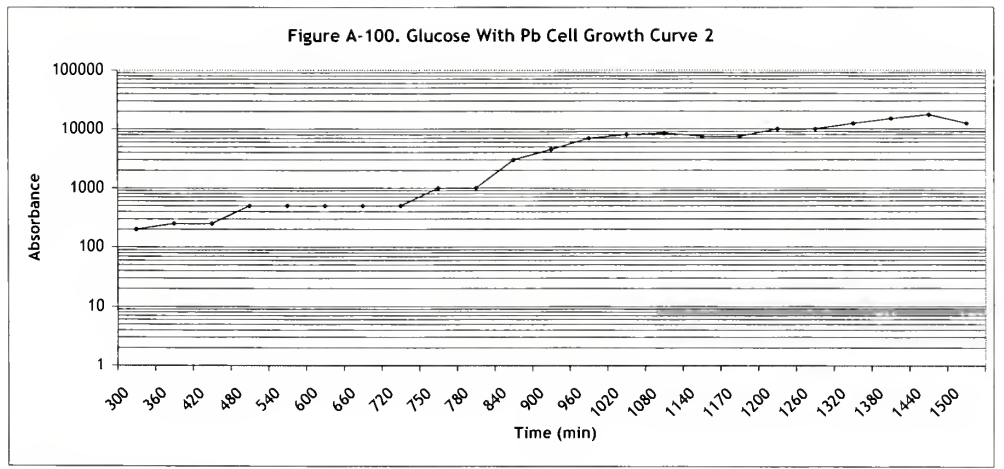
GLUCOSE WITH Pb CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Average
300	200	200	200	200
360	275	250	250	258
420	325	250	325	300
480	400	500	400	433
540	425	500	500	475
600	650	500	650	600
660	850	500	850	733
720	900	500	900	767
750	1000	1000	1000	1000
780	1000	1000	1050	1017
840	2500	3000	2500	2667
900	3900	4500	5000	4467
960	5650	7000	5000	5883
1020	7150	8000	7500	7550
1080	7450	8500	7500	7817
1140	7500	7500	7500	7500
1170	10000	7500	7500	8333
1200	10000	10000	10000	10000
1260	10000	10000	10000	10000
1320	12500	12500	12500	12500
1380	15000	15000	15000	15000
1440	17500	17500	17500	17500
1500	12500	12500	12500	12500



Cell Count increases from 425 to 850

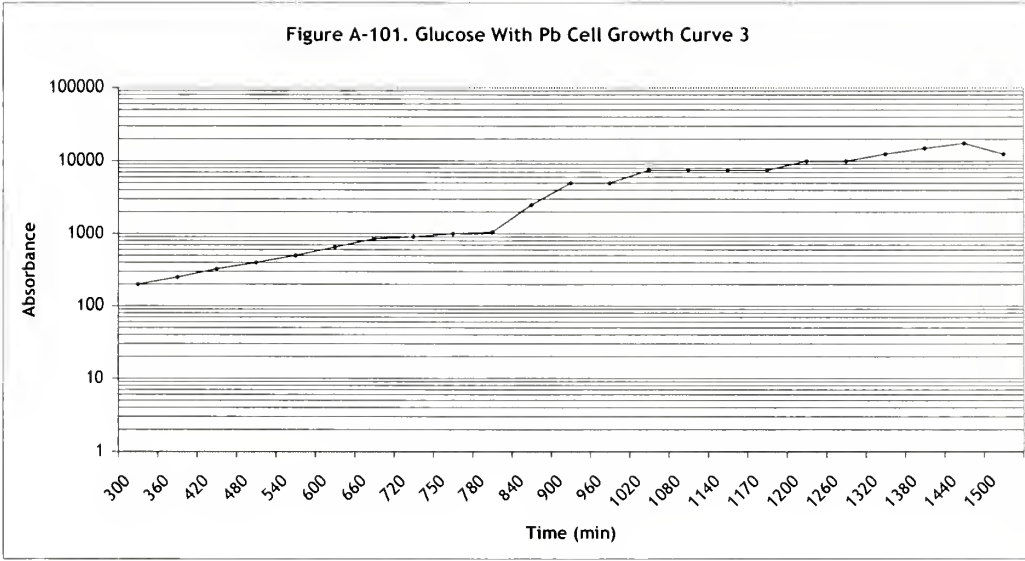
Actual growth rate = (660min - 540min) = 120 min



Cell Count increases from 7500 to 15000

Actual growth rate = (1380min - 1170min) = 210 min

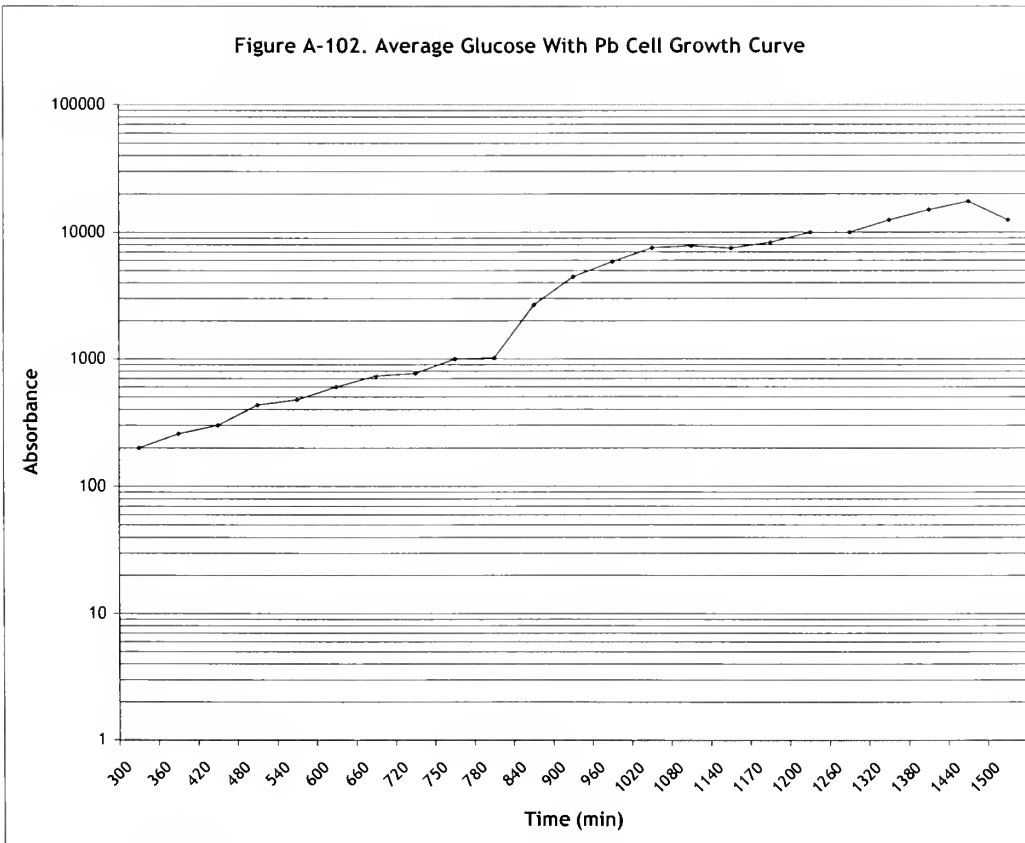
Figure A-101. Glucose With Pb Cell Growth Curve 3



Cell Count increases from 7500 to 15000

Actual growth rate = (1380min - 1170min) = 210 min

Figure A-102. Average Glucose With Pb Cell Growth Curve



Cell Count increases from 300 to 600

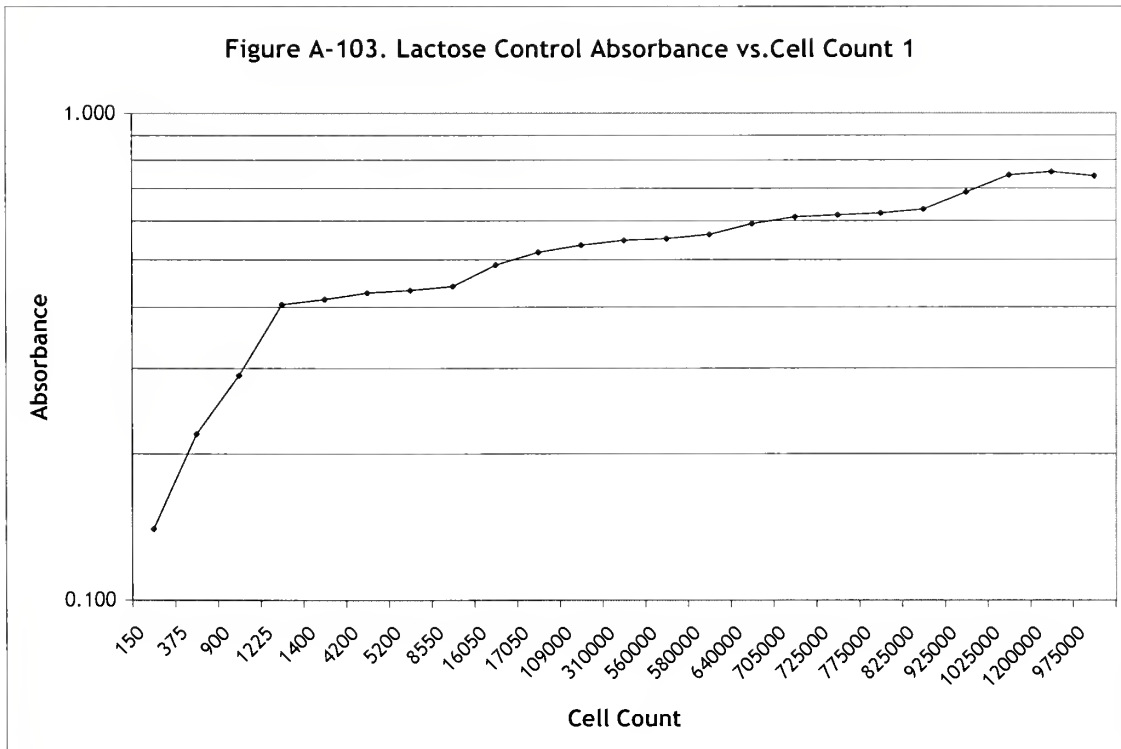
Actual growth rate = (600min - 420min) = 180 min

E. coli Cell growth vs. Absorbance

LACTOSE ONLY CELL COUNT

Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	150	0.140	1.38	0.724	150	0.140
360	375	0.220	1.66	0.603	375	0.220
420	900	0.290	1.95	0.513	900	0.290
480	1225	0.405	2.54	0.394	1225	0.405
540	1400	0.415	2.60	0.385	1400	0.415
600	4200	0.428	2.68	0.373	4200	0.428
660	5200	0.433	2.71	0.369	5200	0.433
720	8550	0.441	2.76	0.362	8550	0.441
750	16050	0.488	3.08	0.325	16050	0.488
780	17050	0.518	3.30	0.303	17050	0.518
840	109000	0.535	3.43	0.292	109000	0.535
900	310000	0.548	3.53	0.283	310000	0.548
960	560000	0.552	3.56	0.281	560000	0.552
1020	580000	0.563	3.66	0.274	580000	0.563
1080	640000	0.592	3.91	0.256	640000	0.592
1140	705000	0.611	4.08	0.245	705000	0.611
1170	725000	0.617	4.14	0.242	725000	0.617
1200	775000	0.623	4.20	0.238	775000	0.623
1260	825000	0.634	4.31	0.232	825000	0.634
1320	925000	0.687	4.86	0.206	925000	0.687
1380	1025000	0.745	5.56	0.180	1025000	0.745
1440	1200000	0.757	5.71	0.175	1200000	0.757
1500	975000	0.742	5.52	0.181	975000	0.742

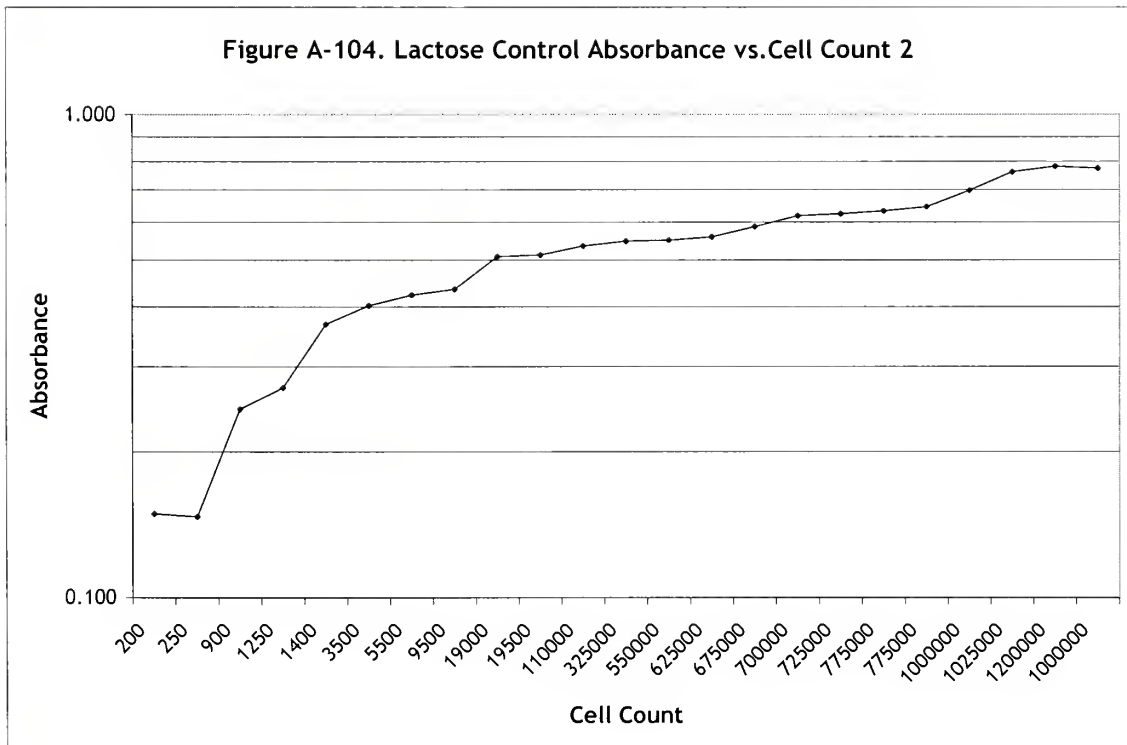
Figure A-103. Lactose Control Absorbance vs. Cell Count 1



Cell Count increases from 8550 to 17050

Difference in absorbance = (0.518 - 0.441) = 0.077

Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.149	1.41	0.710	200	0.149
360	250	0.147	1.40	0.713	250	0.147
420	900	0.245	1.76	0.569	900	0.245
480	1250	0.271	1.87	0.536	1250	0.271
540	1400	0.368	2.33	0.429	1400	0.368
600	3500	0.402	2.52	0.396	3500	0.402
660	5500	0.423	2.65	0.378	5500	0.423
720	9500	0.435	2.72	0.367	9500	0.435
750	19000	0.509	3.23	0.310	19000	0.509
780	19500	0.513	3.26	0.307	19500	0.513
840	110000	0.535	3.43	0.292	110000	0.535
900	325000	0.548	3.53	0.283	325000	0.548
960	550000	0.550	3.55	0.282	550000	0.550
1020	625000	0.559	3.62	0.276	625000	0.559
1080	675000	0.587	3.86	0.259	675000	0.587
1140	700000	0.618	4.15	0.241	700000	0.618
1170	725000	0.624	4.21	0.238	725000	0.624
1200	775000	0.633	4.30	0.233	775000	0.633
1260	775000	0.645	4.42	0.226	775000	0.645
1320	1000000	0.698	4.99	0.200	1000000	0.698
1380	1025000	0.762	5.78	0.173	1025000	0.762
1440	1200000	0.782	6.05	0.165	1200000	0.782
1500	1000000	0.775	5.96	0.168	1000000	0.775

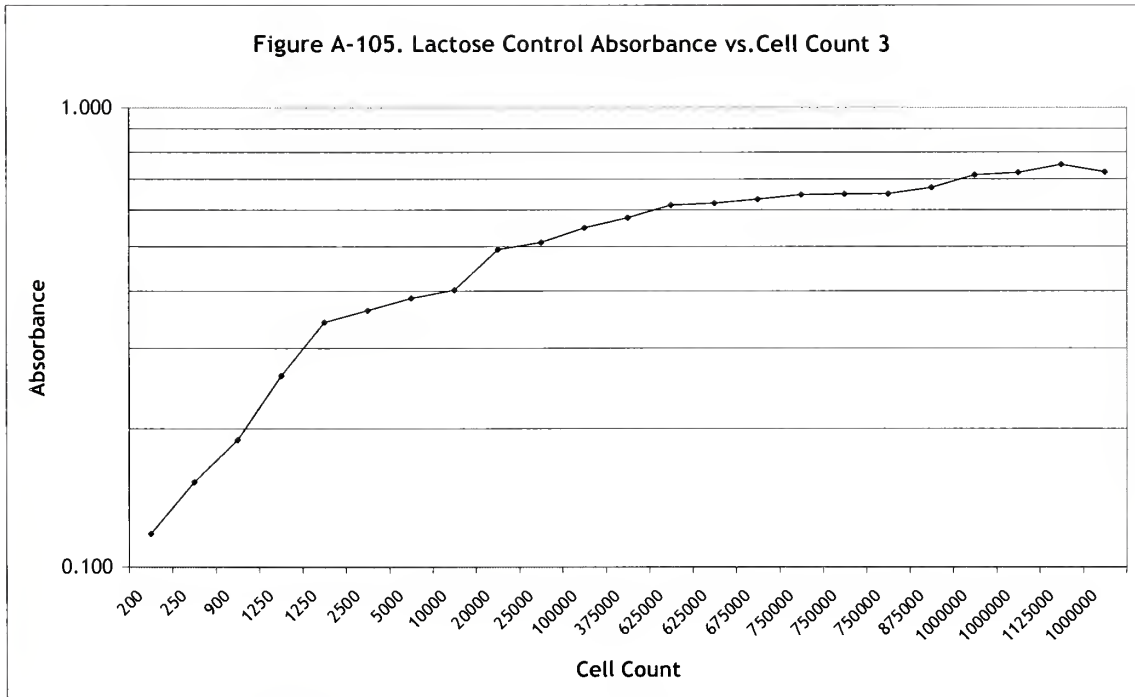


Cell Count increases from 9500 to 19000

Difference in absorbance = (0.509 - 0.435) = 0.074

Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.118	1.31	0.762	200	0.118
360	250	0.153	1.42	0.703	250	0.153
420	900	0.189	1.55	0.647	900	0.189
480	1250	0.261	1.82	0.548	1250	0.261
540	1250	0.342	2.20	0.455	1250	0.342
600	2500	0.363	2.31	0.434	2500	0.363
660	5000	0.386	2.43	0.411	5000	0.386
720	10000	0.402	2.52	0.396	10000	0.402
750	20000	0.493	3.11	0.321	20000	0.493
780	25000	0.511	3.24	0.308	25000	0.511
840	100000	0.549	3.54	0.282	100000	0.549
900	375000	0.577	3.78	0.265	375000	0.577
960	625000	0.615	4.12	0.243	625000	0.615
1020	625000	0.621	4.18	0.239	625000	0.621
1080	675000	0.633	4.30	0.233	675000	0.633
1140	750000	0.648	4.45	0.225	750000	0.648
1170	750000	0.650	4.47	0.224	750000	0.650
1200	750000	0.651	4.48	0.223	750000	0.651
1260	875000	0.671	4.69	0.213	875000	0.671
1320	1000000	0.715	5.19	0.193	1000000	0.715
1380	1000000	0.723	5.28	0.189	1000000	0.723
1440	1125000	0.752	5.65	0.177	1125000	0.752
1500	1000000	0.725	5.31	0.188	1000000	0.725

Figure A-105. Lactose Control Absorbance vs. Cell Count 3

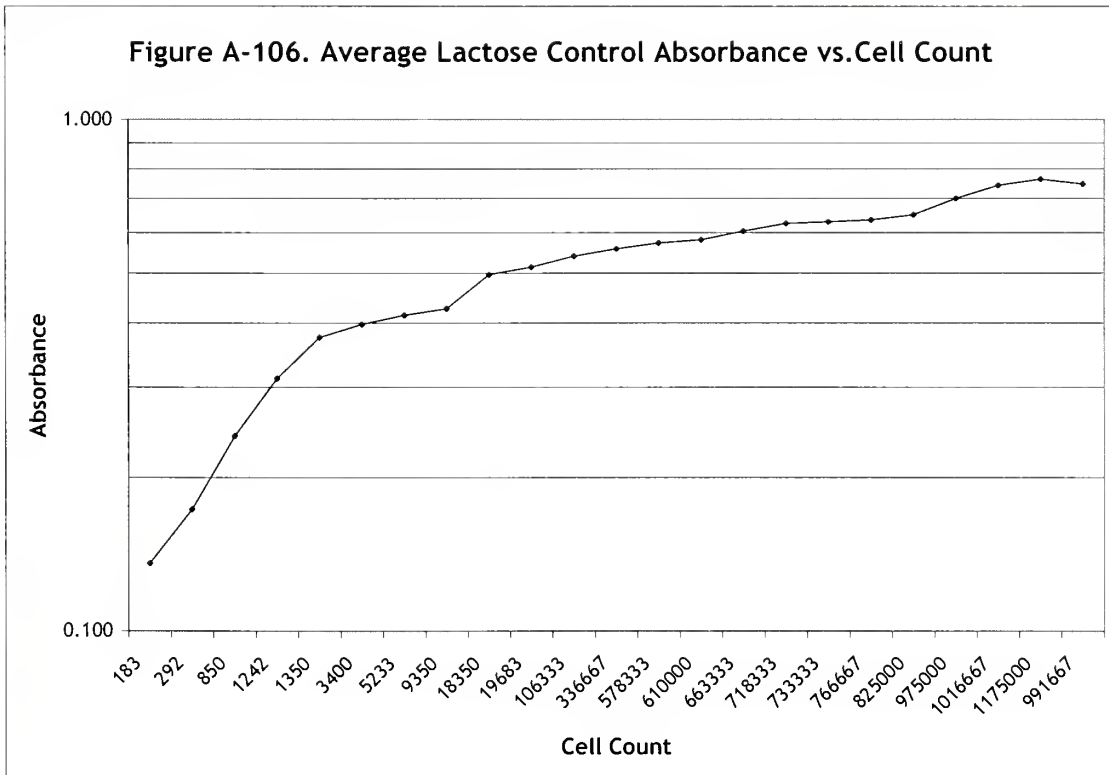


Cell Count increases from 10000 to 20000

Difference in absorbance = (0.493 - 0.402) = 0.091

Time(min)	Average				Cell Count	Absorbance
	Average Cell Count	Absorbance	Antilog	Transmittance		
300	183	0.136	1.37	0.732	183	0.136
360	292	0.173	1.49	0.671	292	0.173
420	850	0.241	1.74	0.574	850	0.241
480	1242	0.312	2.05	0.487	1242	0.312
540	1350	0.375	2.37	0.422	1350	0.375
600	3400	0.398	2.50	0.400	3400	0.398
660	5233	0.414	2.59	0.385	5233	0.414
720	9350	0.426	2.67	0.375	9350	0.426
750	18350	0.497	3.14	0.319	18350	0.497
780	19683	0.514	3.27	0.306	19683	0.514
840	106333	0.540	3.46	0.289	106333	0.540
900	336667	0.558	3.61	0.277	336667	0.558
960	578333	0.572	3.74	0.268	578333	0.572
1020	610000	0.581	3.81	0.262	610000	0.581
1080	663333	0.604	4.02	0.249	663333	0.604
1140	718333	0.626	4.22	0.237	718333	0.626
1170	733333	0.630	4.27	0.234	733333	0.630
1200	766667	0.636	4.32	0.231	766667	0.636
1260	825000	0.650	4.47	0.224	825000	0.650
1320	975000	0.700	5.01	0.200	975000	0.700
1380	1016667	0.743	5.54	0.181	1016667	0.743
1440	1175000	0.764	5.80	0.172	1175000	0.764
1500	991667	0.747	5.59	0.179	991667	0.747

Figure A-106. Average Lactose Control Absorbance vs. Cell Count

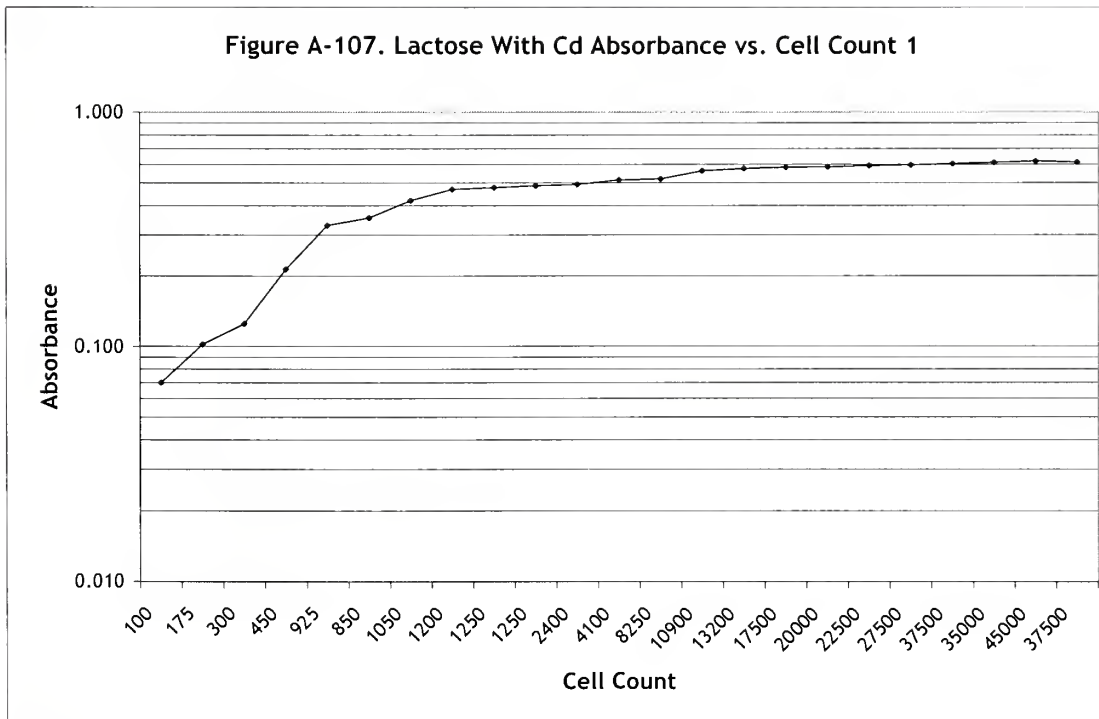


Cell Count increases from 9350 to 18350

Difference in absorbance = (0.497 - 0.426) = 0.071

LACTOSE WITH Cd CELL COUNT

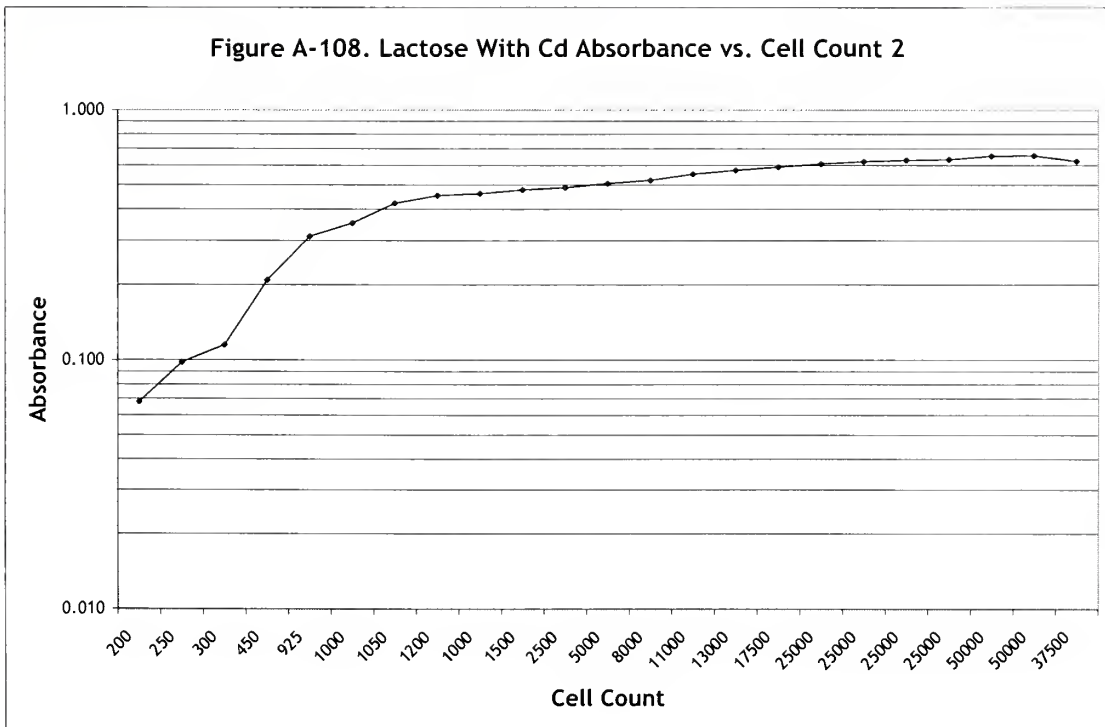
Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	100	0.070	1.17	0.851	100	0.070
360	175	0.102	1.26	0.791	175	0.102
420	300	0.125	1.33	0.750	300	0.125
480	450	0.214	1.64	0.611	450	0.214
540	925	0.330	2.14	0.468	925	0.330
600	850	0.355	2.26	0.442	850	0.355
660	1050	0.420	2.63	0.380	1050	0.420
720	1200	0.470	2.95	0.339	1200	0.470
750	1250	0.478	3.01	0.333	1250	0.478
780	1250	0.486	3.06	0.327	1250	0.486
840	2400	0.492	3.10	0.322	2400	0.492
900	4100	0.515	3.27	0.305	4100	0.515
960	8250	0.520	3.31	0.302	8250	0.520
1020	10900	0.562	3.65	0.274	10900	0.562
1080	13200	0.575	3.76	0.266	13200	0.575
1140	17500	0.582	3.82	0.262	17500	0.582
1170	20000	0.585	3.85	0.260	20000	0.585
1200	22500	0.591	3.90	0.256	22500	0.591
1260	27500	0.596	3.94	0.254	27500	0.596
1320	37500	0.603	4.01	0.249	37500	0.603
1380	35000	0.610	4.07	0.245	35000	0.610
1440	45000	0.618	4.15	0.241	45000	0.618
1500	37500	0.612	4.09	0.244	37500	0.612



Cell Count increases from 10900 to 22500

Difference in absorbance = (0.591 - 0.562) = 0.029

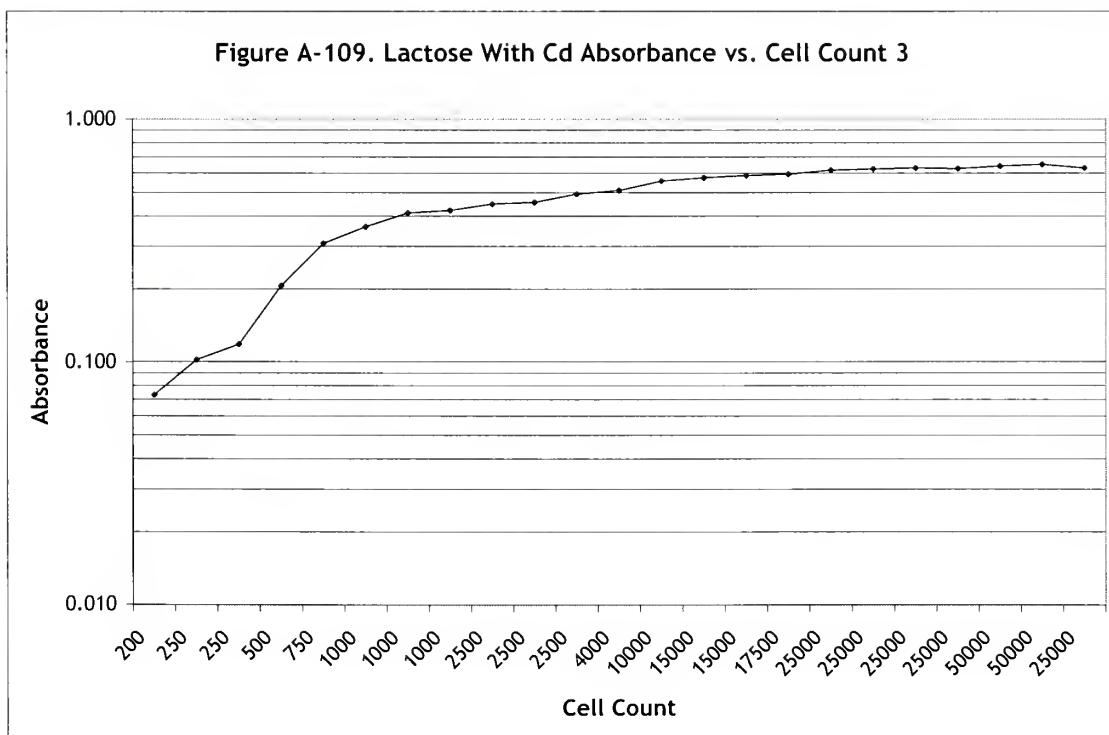
Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.068	1.17	0.855	200	0.068
360	250	0.098	1.25	0.798	250	0.098
420	300	0.115	1.30	0.767	300	0.115
480	450	0.209	1.62	0.618	450	0.209
540	925	0.312	2.05	0.488	925	0.312
600	1000	0.352	2.25	0.445	1000	0.352
660	1050	0.422	2.64	0.378	1050	0.422
720	1200	0.453	2.84	0.352	1200	0.453
750	1000	0.461	2.89	0.346	1000	0.461
780	1500	0.479	3.01	0.332	1500	0.479
840	2500	0.489	3.08	0.324	2500	0.489
900	5000	0.508	3.22	0.310	5000	0.508
960	8000	0.523	3.33	0.300	8000	0.523
1020	11000	0.554	3.58	0.279	11000	0.554
1080	13000	0.574	3.75	0.267	13000	0.574
1140	17500	0.591	3.90	0.256	17500	0.591
1170	25000	0.608	4.06	0.247	25000	0.608
1200	25000	0.622	4.19	0.239	25000	0.622
1260	25000	0.629	4.26	0.235	25000	0.629
1320	25000	0.632	4.29	0.233	25000	0.632
1380	50000	0.654	4.51	0.222	50000	0.654
1440	50000	0.655	4.52	0.221	50000	0.655
1500	37500	0.623	4.20	0.238	37500	0.623



Cell Count increases from 25000 to 50000

Difference in absorbance = (0.654 - 0.632) = 0.022

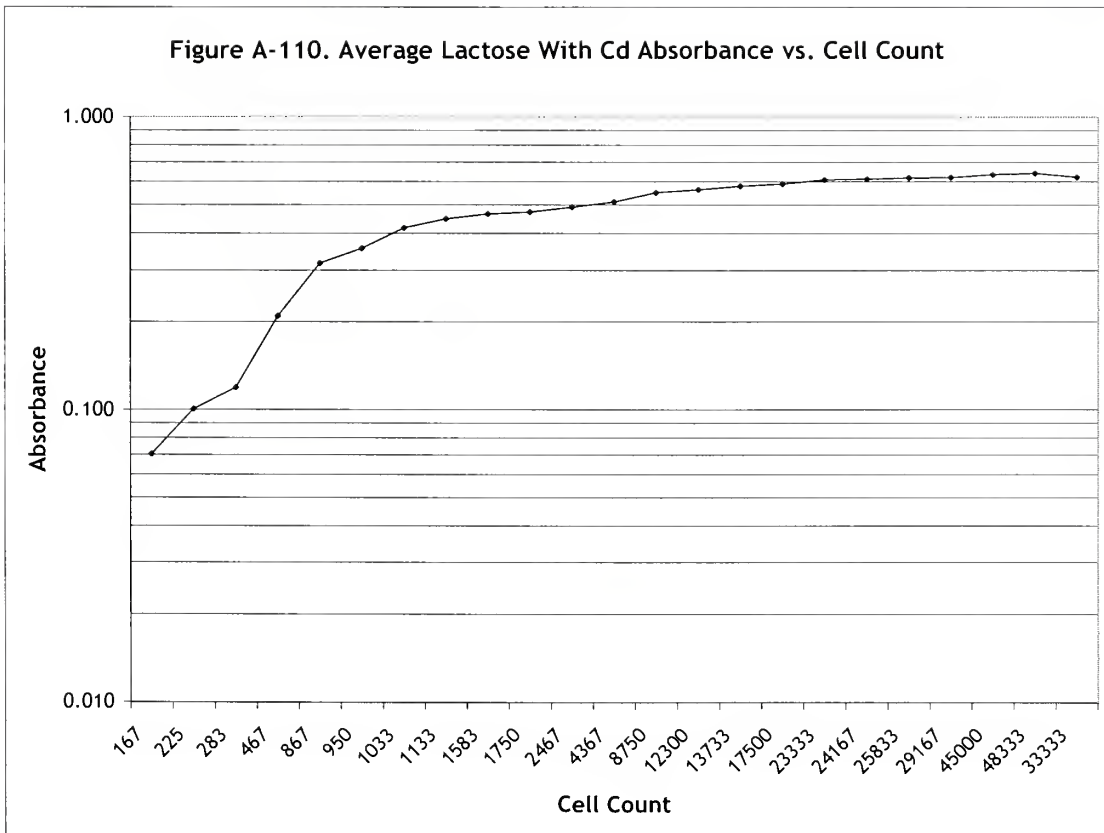
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.073	1.18	0.845	200	0.073
360	250	0.102	1.26	0.791	250	0.102
420	250	0.118	1.31	0.762	250	0.118
480	500	0.206	1.61	0.622	500	0.206
540	750	0.308	2.03	0.492	750	0.308
600	1000	0.362	2.30	0.435	1000	0.362
660	1000	0.412	2.58	0.387	1000	0.412
720	1000	0.422	2.64	0.378	1000	0.422
750	2500	0.449	2.81	0.356	2500	0.449
780	2500	0.456	2.86	0.350	2500	0.456
840	2500	0.493	3.11	0.321	2500	0.493
900	4000	0.511	3.24	0.308	4000	0.511
960	10000	0.558	3.61	0.277	10000	0.558
1020	15000	0.576	3.77	0.265	15000	0.576
1080	15000	0.588	3.87	0.258	15000	0.588
1140	17500	0.595	3.94	0.254	17500	0.595
1170	25000	0.617	4.14	0.242	25000	0.617
1200	25000	0.624	4.21	0.238	25000	0.624
1260	25000	0.631	4.28	0.234	25000	0.631
1320	25000	0.627	4.24	0.236	25000	0.627
1380	50000	0.643	4.40	0.228	50000	0.643
1440	50000	0.652	4.49	0.223	50000	0.652
1500	25000	0.631	4.28	0.234	25000	0.631



Cell Count increases from 25000 to 50000

Difference in absorbance = (0.643 - 0.617) = 0.026

Average						
Time(min)	Cell Count	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	167	0.070	1.18	0.850	167	0.070
360	225	0.101	1.26	0.793	225	0.101
420	283	0.119	1.32	0.760	283	0.119
480	467	0.210	1.62	0.617	467	0.210
540	867	0.317	2.07	0.482	867	0.317
600	950	0.356	2.27	0.440	950	0.356
660	1033	0.418	2.62	0.382	1033	0.418
720	1133	0.448	2.81	0.356	1133	0.448
750	1583	0.466	2.92	0.342	1583	0.466
780	1750	0.474	2.98	0.336	1750	0.474
840	2467	0.491	3.10	0.323	2467	0.491
900	4367	0.511	3.25	0.308	4367	0.511
960	8750	0.551	3.56	0.281	8750	0.551
1020	12300	0.564	3.66	0.273	12300	0.564
1080	13733	0.579	3.79	0.264	13733	0.579
1140	17500	0.589	3.88	0.257	17500	0.589
1170	23333	0.609	4.06	0.246	23333	0.609
1200	24167	0.612	4.10	0.244	24167	0.612
1260	25833	0.619	4.16	0.241	25833	0.619
1320	29167	0.621	4.18	0.240	29167	0.621
1380	45000	0.636	4.32	0.231	45000	0.636
1440	48333	0.642	4.38	0.228	48333	0.642
1500	33333	0.622	4.19	0.239	33333	0.622

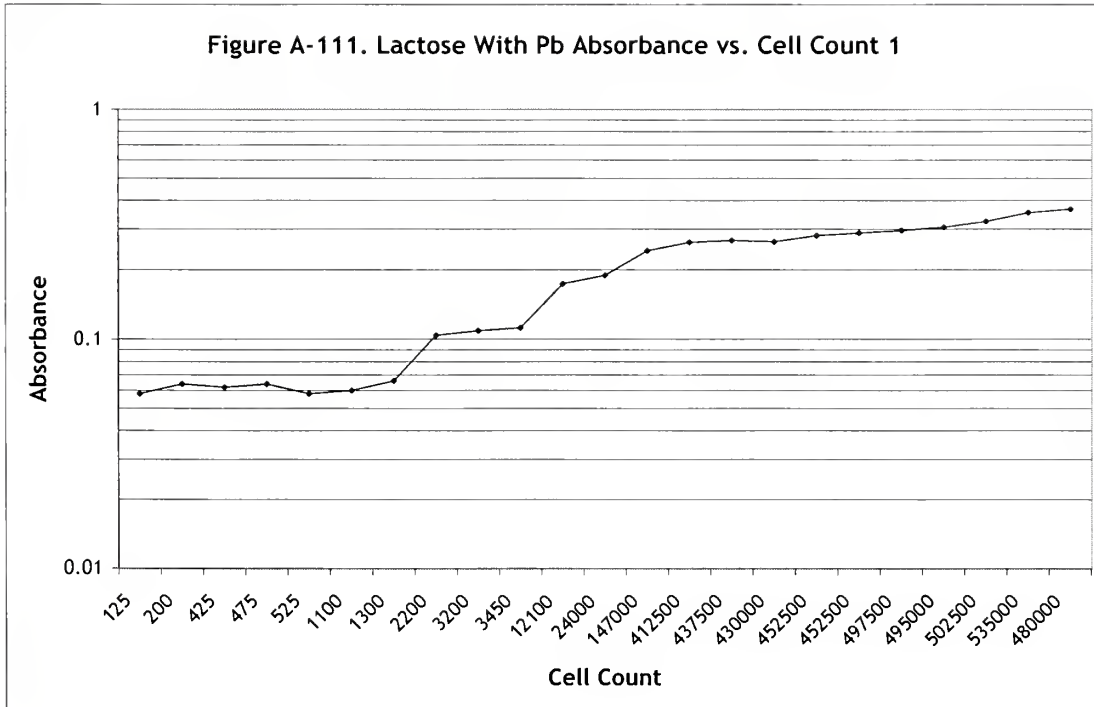


Cell Count increases from 8750 to 17500

Difference in absorbance = (0.589 - 0.551) = 0.038

LACTOSE WITH Pb CELL COUNT

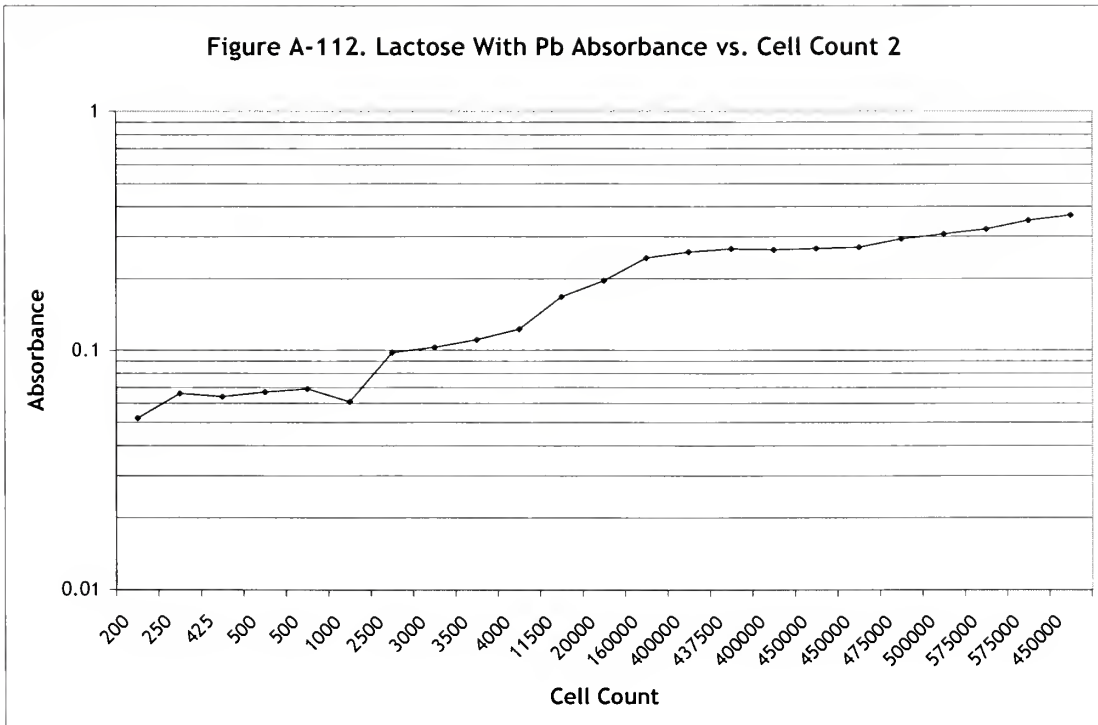
Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	125	0.058	1.14	0.875	125	0.058
360	200	0.064	1.16	0.863	200	0.064
420	425	0.062	1.15	0.867	425	0.062
480	475	0.064	1.16	0.863	475	0.064
540	525	0.058	1.14	0.875	525	0.058
600	1100	0.060	1.15	0.871	1100	0.060
660	1300	0.066	1.16	0.859	1300	0.066
720	2200	0.104	1.27	0.787	2200	0.104
750	3200	0.109	1.29	0.778	3200	0.109
780	3450	0.112	1.29	0.773	3450	0.112
840	12100	0.174	1.49	0.670	12100	0.174
900	24000	0.189	1.55	0.647	24000	0.189
960	147000	0.242	1.75	0.573	147000	0.242
1020	412500	0.263	1.83	0.546	412500	0.263
1080	437500	0.268	1.85	0.540	437500	0.268
1140	430000	0.265	1.84	0.543	430000	0.265
1170	452500	0.281	1.91	0.524	452500	0.281
1200	452500	0.289	1.95	0.514	452500	0.289
1260	497500	0.296	1.98	0.506	497500	0.296
1320	495000	0.306	2.02	0.494	495000	0.306
1380	502500	0.324	2.11	0.474	502500	0.324
1440	535000	0.354	2.26	0.443	535000	0.354
1500	480000	0.366	2.32	0.431	480000	0.366



Cell Count increases from 1100 to 2200

Difference in absorbance = (0.104 - 0.060) = 0.044

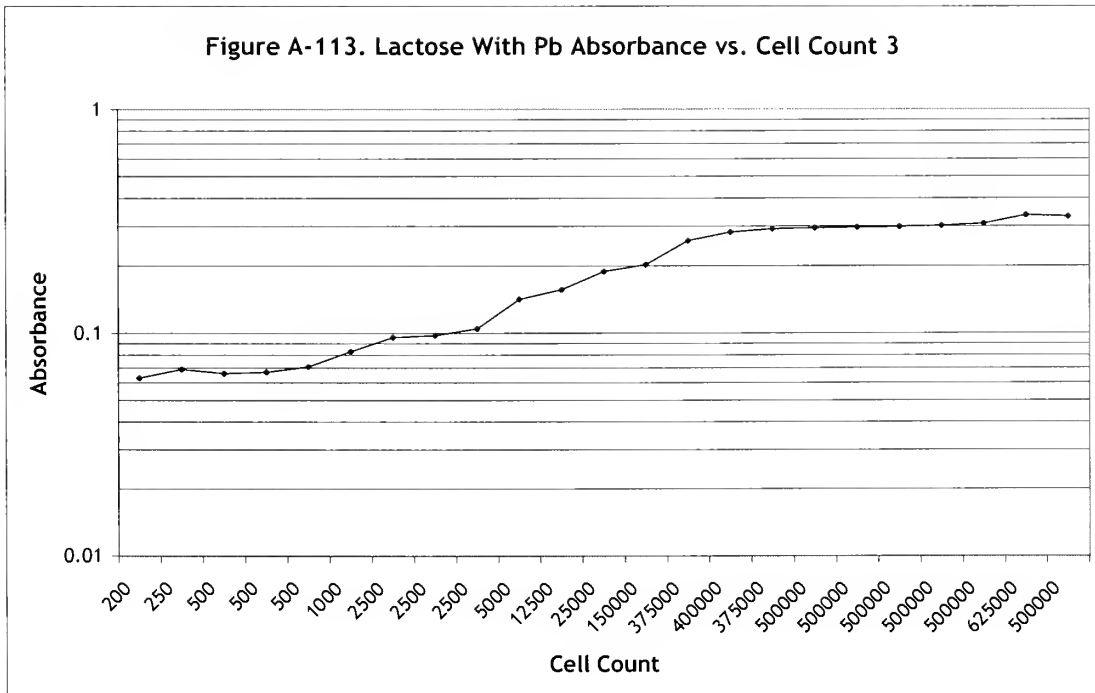
Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.052	1.13	0.887	200	0.052
360	250	0.066	1.16	0.859	250	0.066
420	425	0.064	1.16	0.863	425	0.064
480	500	0.067	1.17	0.857	500	0.067
540	500	0.069	1.17	0.853	500	0.069
600	1000	0.061	1.15	0.869	1000	0.061
660	2500	0.098	1.25	0.798	2500	0.098
720	3000	0.103	1.27	0.789	3000	0.103
750	3500	0.111	1.29	0.774	3500	0.111
780	4000	0.123	1.33	0.753	4000	0.123
840	11500	0.168	1.47	0.679	11500	0.168
900	20000	0.196	1.57	0.637	20000	0.196
960	160000	0.245	1.76	0.569	160000	0.245
1020	400000	0.259	1.82	0.551	400000	0.259
1080	437500	0.267	1.85	0.541	437500	0.267
1140	400000	0.264	1.84	0.545	400000	0.264
1170	450000	0.268	1.85	0.540	450000	0.268
1200	450000	0.271	1.87	0.536	450000	0.271
1260	475000	0.294	1.97	0.508	475000	0.294
1320	500000	0.308	2.03	0.492	500000	0.308
1380	575000	0.322	2.10	0.476	575000	0.322
1440	575000	0.351	2.24	0.446	575000	0.351
1500	450000	0.368	2.33	0.429	450000	0.368



Cell Count increases from 1000 to 2500

Difference in absorbance = (0.098 - 0.061) = 0.037

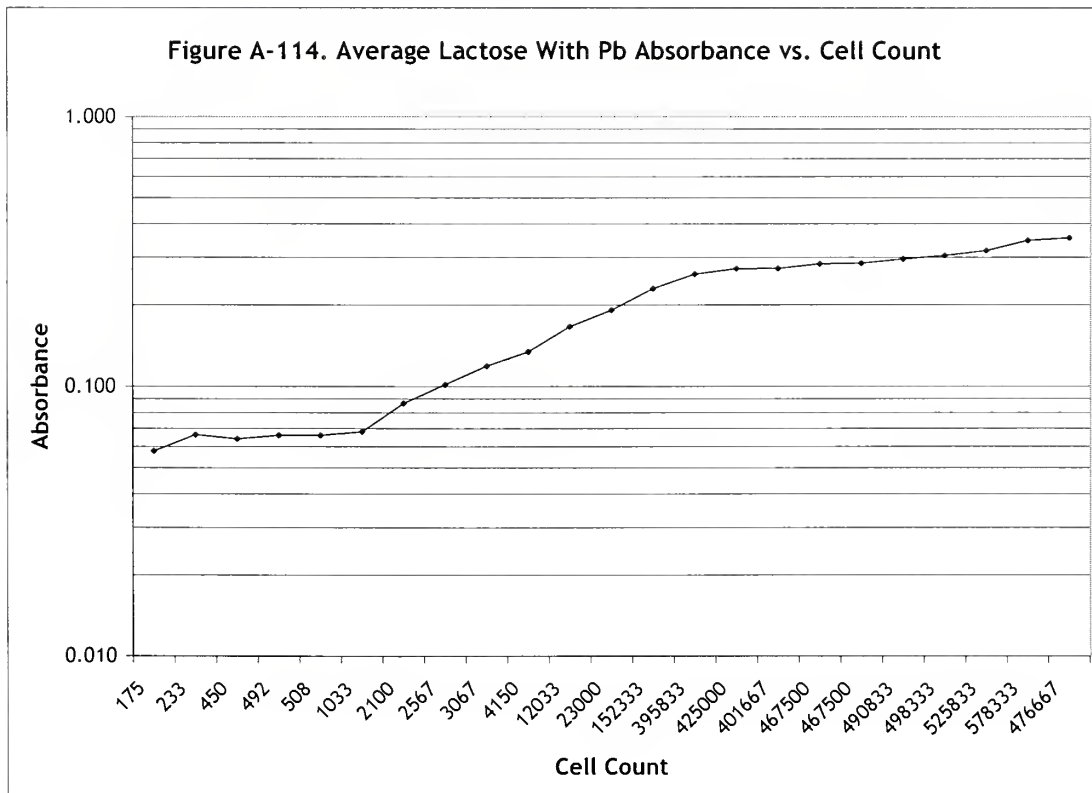
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.063	1.16	0.865	200	0.063
360	250	0.069	1.17	0.853	250	0.069
420	500	0.066	1.16	0.859	500	0.066
480	500	0.067	1.17	0.857	500	0.067
540	500	0.071	1.18	0.849	500	0.071
600	1000	0.083	1.21	0.826	1000	0.083
660	2500	0.096	1.25	0.802	2500	0.096
720	2500	0.098	1.25	0.798	2500	0.098
750	2500	0.105	1.27	0.785	2500	0.105
780	5000	0.142	1.39	0.721	5000	0.142
840	12500	0.157	1.44	0.697	12500	0.157
900	25000	0.189	1.55	0.647	25000	0.189
960	150000	0.203	1.60	0.627	150000	0.203
1020	375000	0.259	1.82	0.551	375000	0.259
1080	400000	0.283	1.92	0.521	400000	0.283
1140	375000	0.292	1.96	0.511	375000	0.292
1170	500000	0.295	1.97	0.507	500000	0.295
1200	500000	0.297	1.98	0.505	500000	0.297
1260	500000	0.299	1.99	0.502	500000	0.299
1320	500000	0.302	2.00	0.499	500000	0.302
1380	500000	0.308	2.03	0.492	500000	0.308
1440	625000	0.336	2.17	0.461	625000	0.336
1500	500000	0.332	2.15	0.466	500000	0.332



Cell Count increases from 2500 to 5000

Difference in absorbance = (0.142 - 0.096) = 0.046

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	175	0.058	1.14	0.876	175	0.058
360	233	0.066	1.17	0.858	233	0.066
420	450	0.064	1.16	0.863	450	0.064
480	492	0.066	1.16	0.859	492	0.066
540	508	0.066	1.16	0.859	508	0.066
600	1033	0.068	1.17	0.855	1033	0.068
660	2100	0.087	1.22	0.819	2100	0.087
720	2567	0.102	1.26	0.791	2567	0.102
750	3067	0.119	1.32	0.760	3067	0.119
780	4150	0.134	1.36	0.735	4150	0.134
840	12033	0.166	1.47	0.682	12033	0.166
900	23000	0.191	1.55	0.644	23000	0.191
960	152333	0.230	1.70	0.589	152333	0.230
1020	395833	0.260	1.82	0.549	395833	0.260
1080	425000	0.273	1.87	0.534	425000	0.273
1140	401667	0.274	1.88	0.533	401667	0.274
1170	467500	0.284	1.92	0.520	467500	0.284
1200	467500	0.286	1.93	0.518	467500	0.286
1260	490833	0.296	1.98	0.505	490833	0.296
1320	498333	0.305	2.02	0.495	498333	0.305
1380	525833	0.318	2.08	0.481	525833	0.318
1440	578333	0.347	2.22	0.450	578333	0.347
1500	476667	0.355	2.27	0.441	476667	0.355



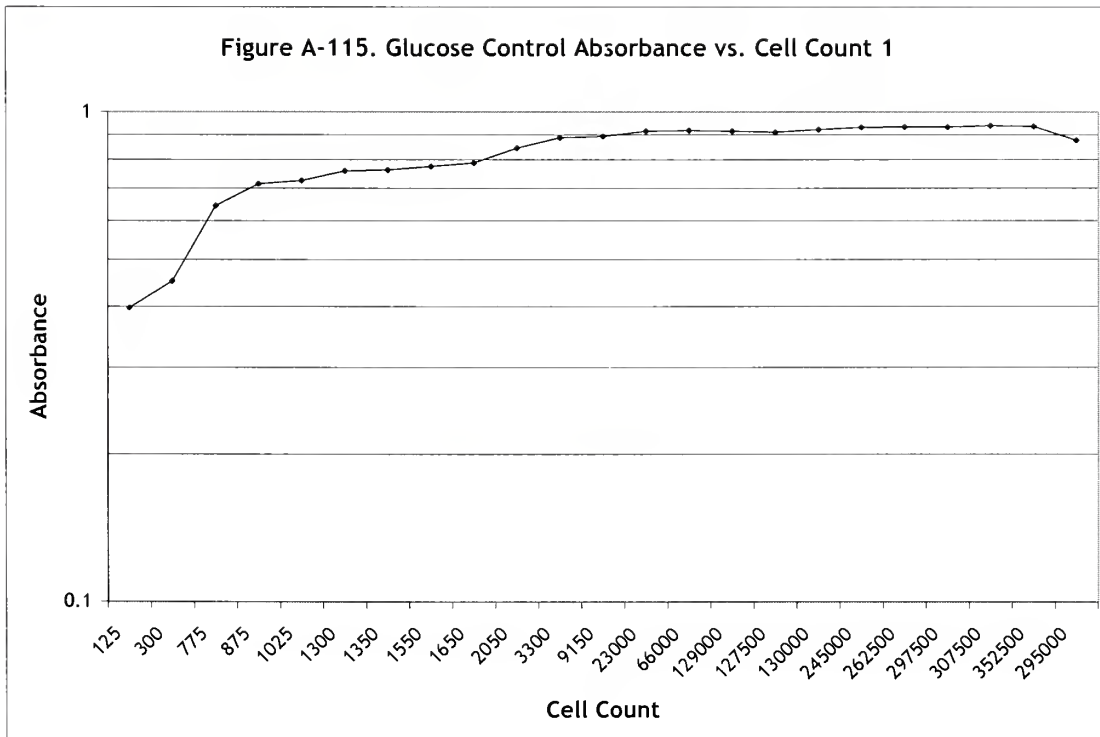
Cell Count increases from 2100 to 4150

Difference in absorbance = (0.134 - 0.087) = 0.047

GLUCOSE ONLY CELL COUNT

Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	125	0.398	2.50	0.400	125	0.398
360	300	0.452	2.83	0.353	300	0.452
420	775	0.645	4.42	0.226	775	0.645
480	875	0.715	5.19	0.193	875	0.715
540	1025	0.726	5.32	0.188	1025	0.726
600	1300	0.759	5.74	0.174	1300	0.759
660	1350	0.763	5.79	0.173	1350	0.763
720	1550	0.775	5.96	0.168	1550	0.775
750	1650	0.788	6.14	0.163	1650	0.788
780	2050	0.845	7.00	0.143	2050	0.845
840	3300	0.887	7.71	0.130	3300	0.887
900	9150	0.892	7.80	0.128	9150	0.892
960	23000	0.915	8.22	0.122	23000	0.915
1020	66000	0.918	8.28	0.121	66000	0.918
1080	129000	0.915	8.22	0.122	129000	0.915
1140	127500	0.911	8.15	0.123	127500	0.911
1170	130000	0.921	8.34	0.120	130000	0.921
1200	245000	0.932	8.55	0.117	245000	0.932
1260	262500	0.933	8.57	0.117	262500	0.933
1320	297500	0.933	8.57	0.117	297500	0.933
1380	307500	0.938	8.67	0.115	307500	0.938
1440	352500	0.935	8.61	0.116	352500	0.935
1500	295000	0.877	7.53	0.133	295000	0.877

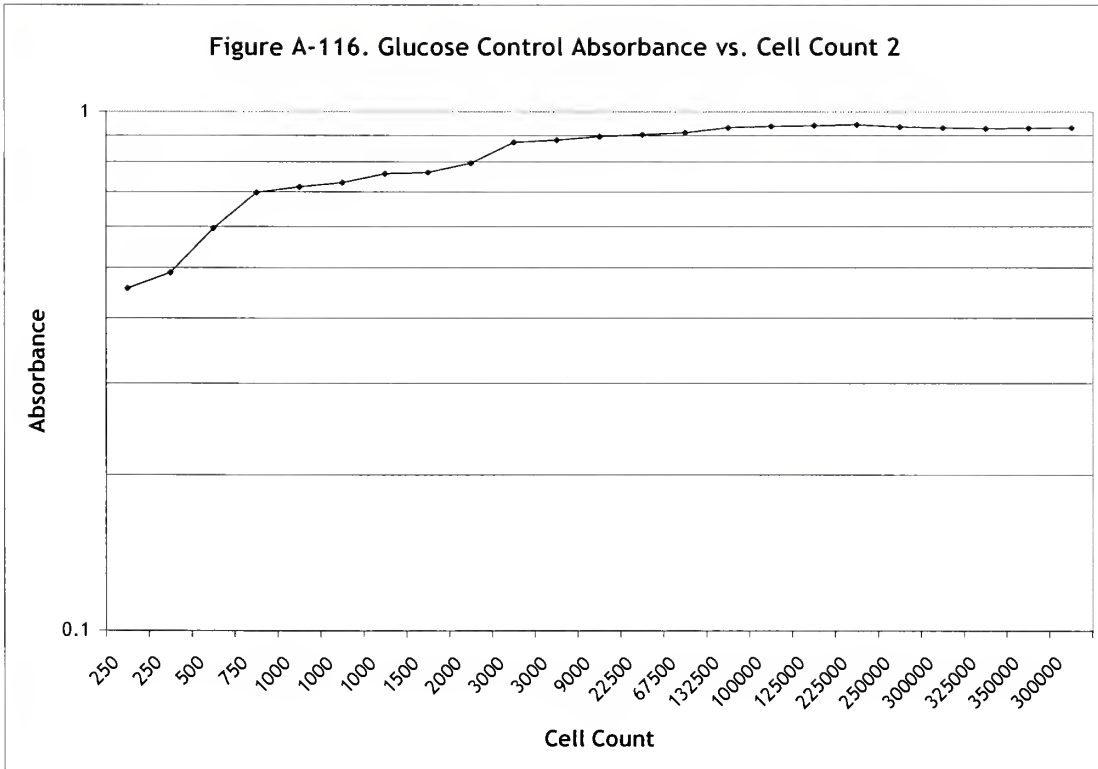
Figure A-115. Glucose Control Absorbance vs. Cell Count 1



Cell Count increases from 1650 to 3300

Difference in absorbance = (0.887 - 0.788) = 0.099

Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	250	0.456	2.86	0.350	250	0.456
360	250	0.489	3.08	0.324	250	0.489
420	500	0.596	3.94	0.254	500	0.596
480	750	0.698	4.99	0.200	750	0.698
540	1000	0.716	5.20	0.192	1000	0.716
600	1000	0.729	5.36	0.187	1000	0.729
660	1000	0.759	5.74	0.174	1000	0.759
720	1500	0.764	5.81	0.172	1500	0.764
750	2000	0.796	6.25	0.160	2000	0.796
780	3000	0.873	7.46	0.134	3000	0.873
840	3000	0.882	7.62	0.131	3000	0.882
900	9000	0.896	7.87	0.127	9000	0.896
960	22500	0.903	8.00	0.125	22500	0.903
1020	67500	0.912	8.17	0.122	67500	0.912
1080	132500	0.933	8.57	0.117	132500	0.933
1140	100000	0.938	8.67	0.115	100000	0.938
1170	125000	0.941	8.73	0.115	125000	0.941
1200	225000	0.945	8.81	0.114	225000	0.945
1260	250000	0.935	8.61	0.116	250000	0.935
1320	300000	0.931	8.53	0.117	300000	0.931
1380	325000	0.928	8.47	0.118	325000	0.928
1440	350000	0.930	8.51	0.117	350000	0.930
1500	300000	0.931	8.53	0.117	300000	0.931

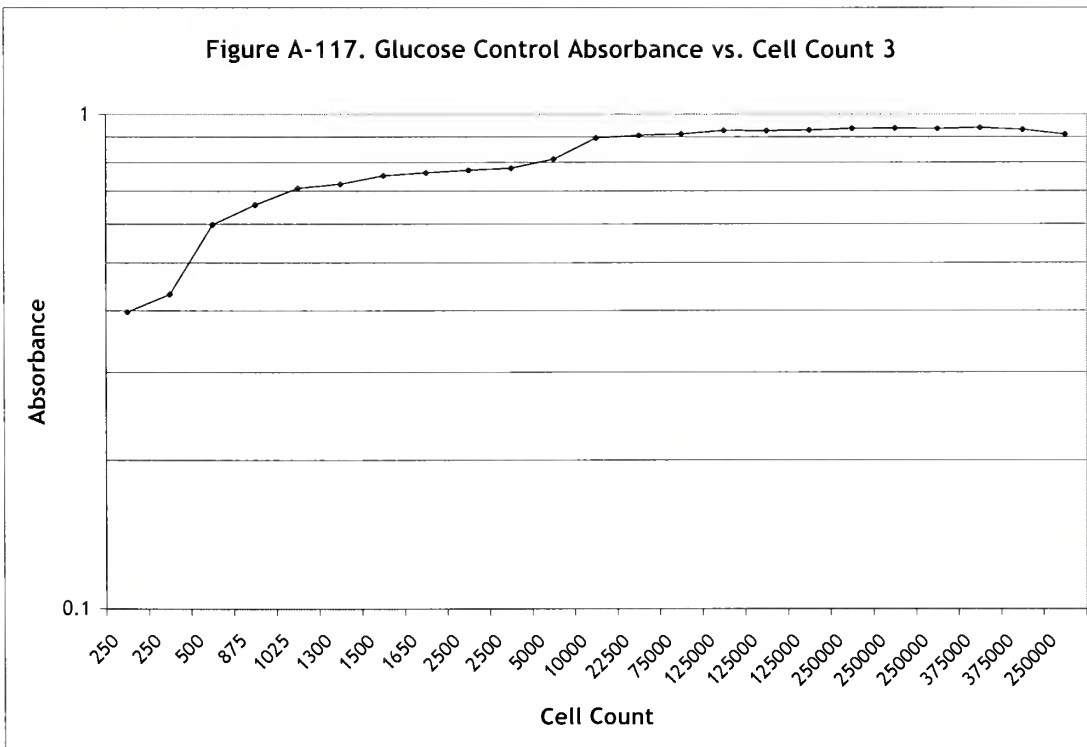


Cell Count increases from 1500 to 3000

Difference in absorbance = (0.873 - 0.764) = 0.109

Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	250	0.398	2.50	0.400	250	0.398
360	250	0.432	2.70	0.370	250	0.432
420	500	0.598	3.96	0.252	500	0.598
480	875	0.656	4.53	0.221	875	0.656
540	1025	0.709	5.12	0.195	1025	0.709
600	1300	0.723	5.28	0.189	1300	0.723
660	1500	0.752	5.65	0.177	1500	0.752
720	1650	0.762	5.78	0.173	1650	0.762
750	2500	0.771	5.90	0.169	2500	0.771
780	2500	0.779	6.01	0.166	2500	0.779
840	5000	0.812	6.49	0.154	5000	0.812
900	10000	0.896	7.87	0.127	10000	0.896
960	22500	0.907	8.07	0.124	22500	0.907
1020	75000	0.912	8.17	0.122	75000	0.912
1080	125000	0.928	8.47	0.118	125000	0.928
1140	125000	0.927	8.45	0.118	125000	0.927
1170	125000	0.929	8.49	0.118	125000	0.929
1200	250000	0.936	8.63	0.116	250000	0.936
1260	250000	0.938	8.67	0.115	250000	0.938
1320	250000	0.936	8.63	0.116	250000	0.936
1380	375000	0.941	8.73	0.115	375000	0.941
1440	375000	0.932	8.55	0.117	375000	0.932
1500	250000	0.911	8.15	0.123	250000	0.911

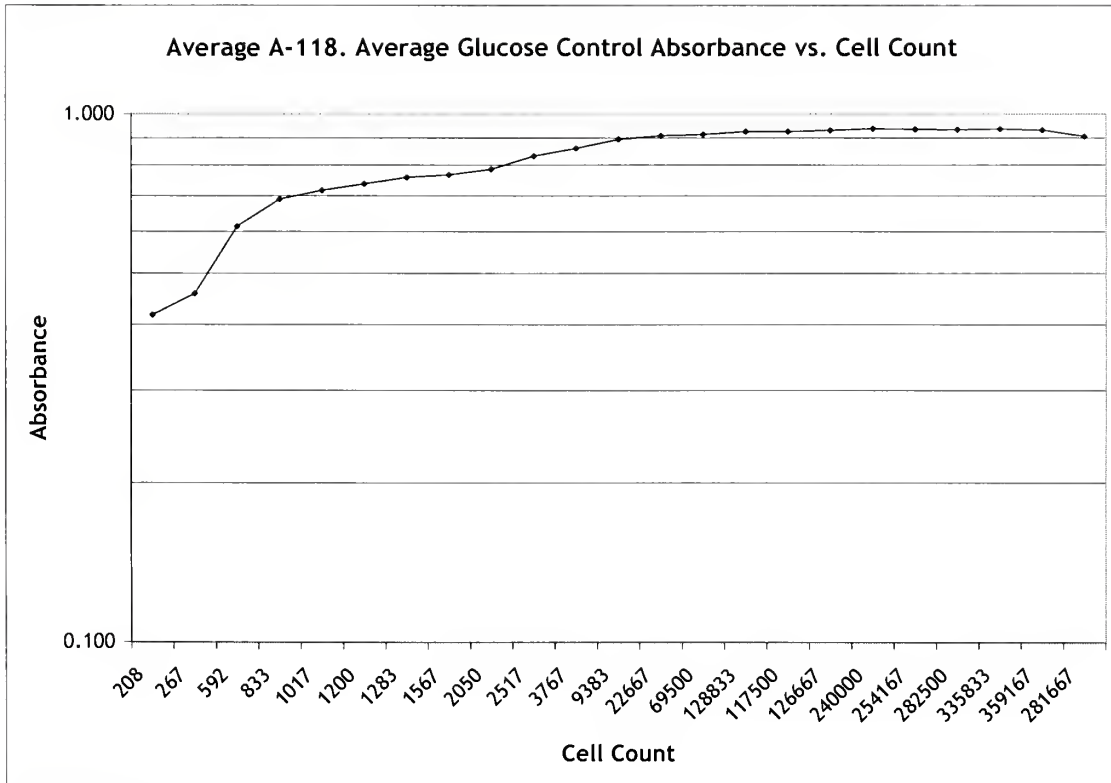
Figure A-117. Glucose Control Absorbance vs. Cell Count 3



Cell Count increases from 5000 to 10000

Difference in absorbance = (0.896 - 0.812) = 0.084

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	208	0.417	2.61	0.383	208	0.417
360	267	0.458	2.87	0.349	267	0.458
420	592	0.613	4.10	0.244	592	0.613
480	833	0.690	4.89	0.204	833	0.690
540	1017	0.717	5.21	0.192	1017	0.717
600	1200	0.737	5.46	0.183	1200	0.737
660	1283	0.758	5.73	0.175	1283	0.758
720	1567	0.767	5.85	0.171	1567	0.767
750	2050	0.785	6.10	0.164	2050	0.785
780	2517	0.832	6.80	0.147	2517	0.832
840	3767	0.860	7.25	0.138	3767	0.860
900	9383	0.895	7.85	0.127	9383	0.895
960	22667	0.908	8.10	0.123	22667	0.908
1020	69500	0.914	8.20	0.122	69500	0.914
1080	128833	0.925	8.42	0.119	128833	0.925
1140	117500	0.925	8.42	0.119	117500	0.925
1170	126667	0.930	8.52	0.117	126667	0.930
1200	240000	0.938	8.66	0.115	240000	0.938
1260	254167	0.935	8.62	0.116	254167	0.935
1320	282500	0.933	8.58	0.117	282500	0.933
1380	335833	0.936	8.62	0.116	335833	0.936
1440	359167	0.932	8.56	0.117	359167	0.932
1500	281667	0.906	8.06	0.124	281667	0.906

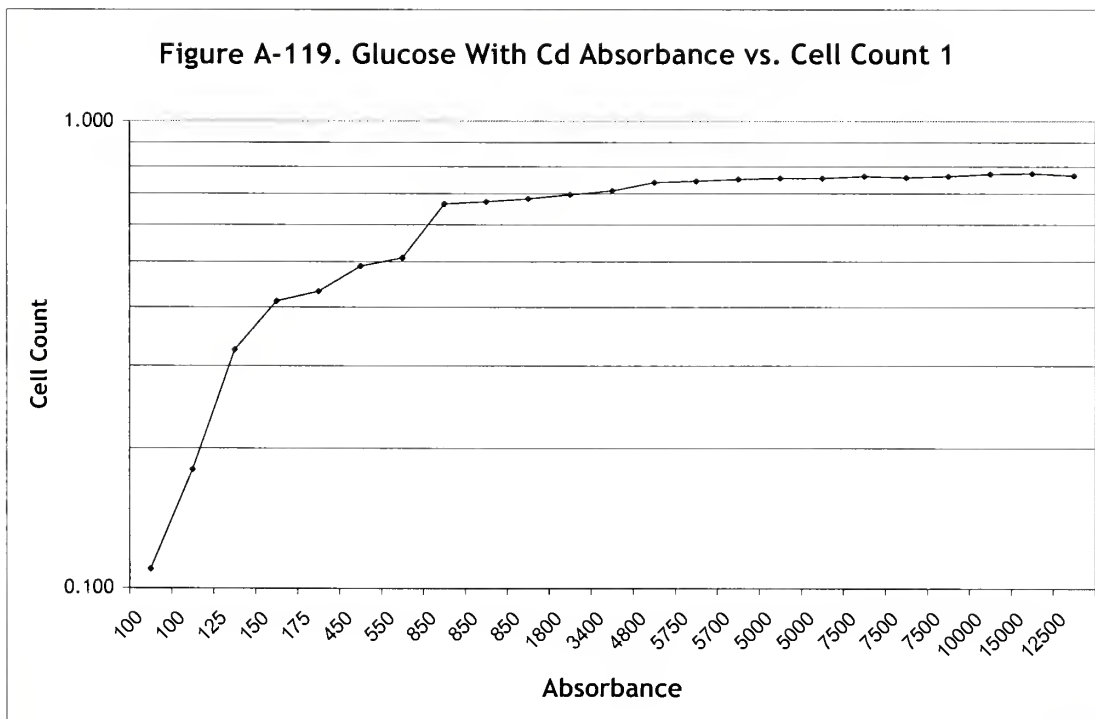


Cell Count increases from 592 to 1200

Difference in absorbance = (0.737 - 0.613) = 0.124

GLUCOSE WITH Cd CELL COUNT

Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	100	0.110	1.29	0.776	100	0.110
360	100	0.180	1.51	0.661	100	0.180
420	125	0.325	2.11	0.473	125	0.325
480	150	0.412	2.58	0.387	150	0.412
540	175	0.432	2.70	0.370	175	0.432
600	450	0.489	3.08	0.324	450	0.489
660	550	0.510	3.24	0.309	550	0.510
720	850	0.665	4.62	0.216	850	0.665
750	850	0.673	4.71	0.212	850	0.673
780	850	0.682	4.81	0.208	850	0.682
840	1800	0.697	4.98	0.201	1800	0.697
900	3400	0.710	5.13	0.195	3400	0.710
960	4800	0.740	5.50	0.182	4800	0.740
1020	5750	0.745	5.56	0.180	5750	0.745
1080	5700	0.752	5.65	0.177	5700	0.752
1140	5000	0.755	5.69	0.176	5000	0.755
1170	5000	0.755	5.69	0.176	5000	0.755
1200	7500	0.762	5.78	0.173	7500	0.762
1260	7500	0.758	5.73	0.175	7500	0.758
1320	7500	0.763	5.79	0.173	7500	0.763
1380	10000	0.771	5.90	0.169	10000	0.771
1440	15000	0.774	5.94	0.168	15000	0.774
1500	12500	0.765	5.82	0.172	12500	0.765

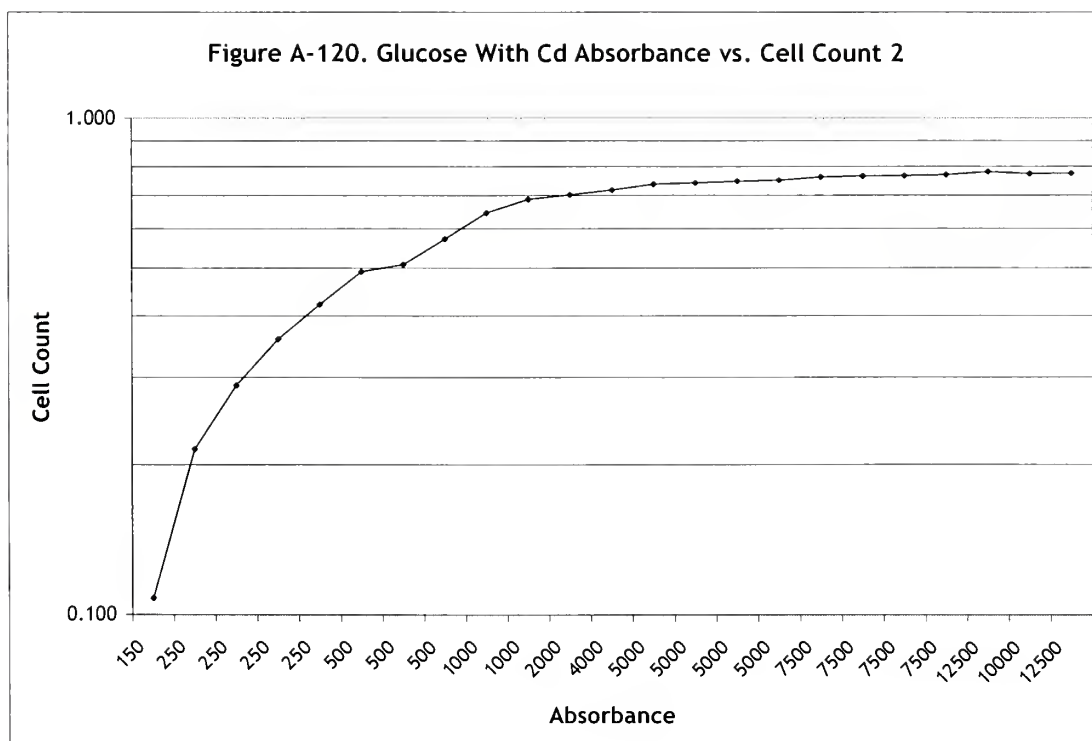


Cell Count increases from 5000 to 10000

Difference in absorbance = (0.771 - 0.755) = 0.016

Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	150	0.108	1.28	0.780	150	0.108
360	250	0.215	1.64	0.610	250	0.215
420	250	0.289	1.95	0.514	250	0.289
480	250	0.359	2.29	0.438	250	0.359
540	250	0.422	2.64	0.378	250	0.422
600	500	0.492	3.10	0.322	500	0.492
660	500	0.508	3.22	0.310	500	0.508
720	500	0.572	3.73	0.268	500	0.572
750	1000	0.646	4.43	0.226	1000	0.646
780	1000	0.688	4.88	0.205	1000	0.688
840	2000	0.702	5.04	0.199	2000	0.702
900	4000	0.718	5.22	0.191	4000	0.718
960	5000	0.738	5.47	0.183	5000	0.738
1020	5000	0.742	5.52	0.181	5000	0.742
1080	5000	0.748	5.60	0.179	5000	0.748
1140	5000	0.751	5.64	0.177	5000	0.751
1170	7500	0.763	5.79	0.173	7500	0.763
1200	7500	0.766	5.83	0.171	7500	0.766
1260	7500	0.767	5.85	0.171	7500	0.767
1320	7500	0.771	5.90	0.169	7500	0.771
1380	12500	0.781	6.04	0.166	12500	0.781
1440	10000	0.774	5.94	0.168	10000	0.774
1500	12500	0.775	5.96	0.168	12500	0.775

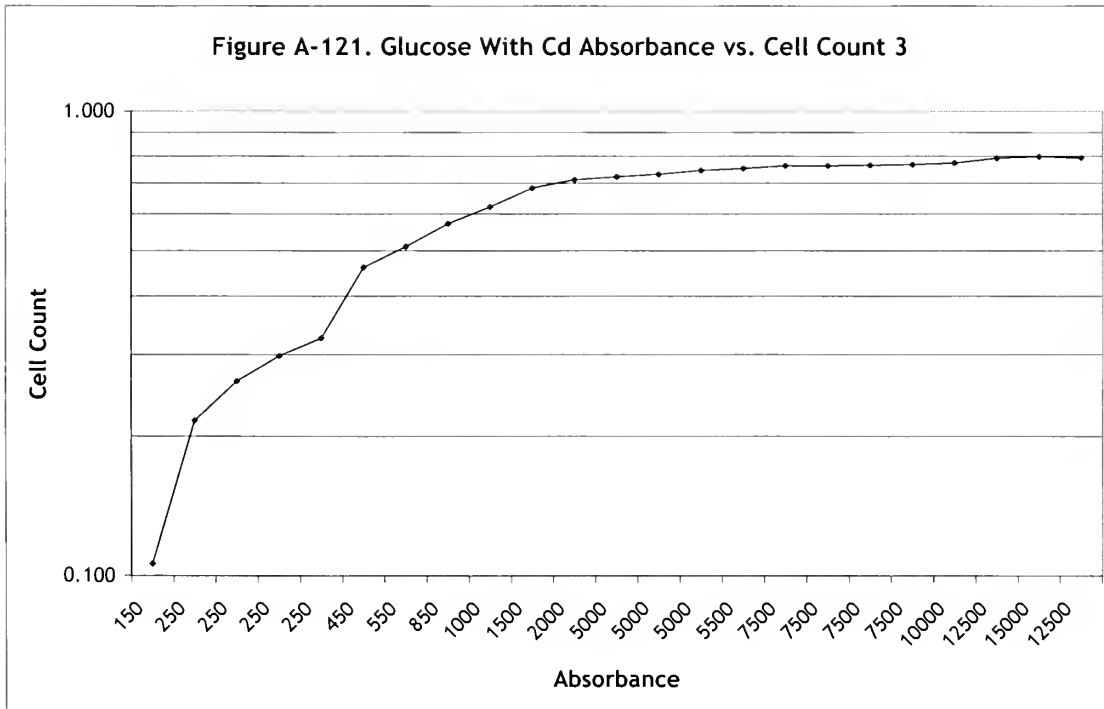
Figure A-120. Glucose With Cd Absorbance vs. Cell Count 2



Cell Count increases from 5000 to 10000

Difference in absorbance = $(0.774 - 0.751) = 0.023$

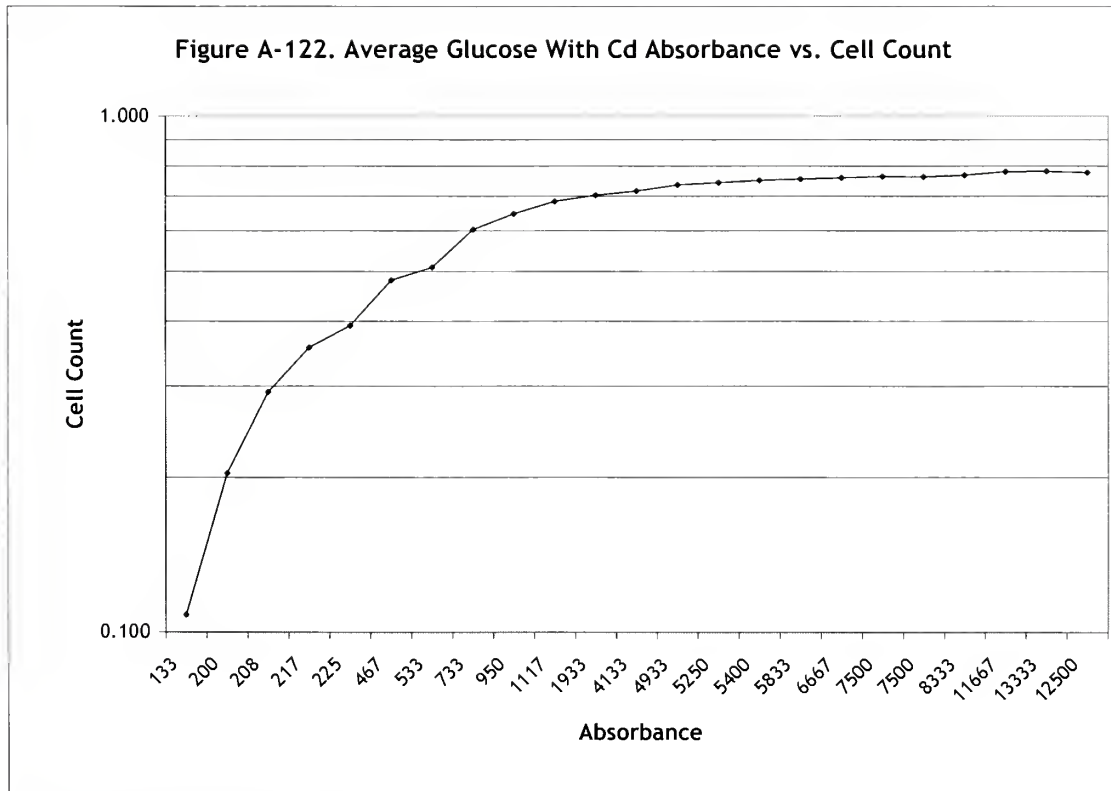
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	150	0.106	1.28	0.783	150	0.106
360	250	0.216	1.64	0.608	250	0.216
420	250	0.263	1.83	0.546	250	0.263
480	250	0.298	1.99	0.504	250	0.298
540	250	0.325	2.11	0.473	250	0.325
600	450	0.461	2.89	0.346	450	0.461
660	550	0.511	3.24	0.308	550	0.511
720	850	0.572	3.73	0.268	850	0.572
750	1000	0.622	4.19	0.239	1000	0.622
780	1500	0.683	4.82	0.207	1500	0.683
840	2000	0.712	5.15	0.194	2000	0.712
900	5000	0.722	5.27	0.190	5000	0.722
960	5000	0.731	5.38	0.186	5000	0.731
1020	5000	0.746	5.57	0.179	5000	0.746
1080	5500	0.753	5.66	0.177	5500	0.753
1140	7500	0.762	5.78	0.173	7500	0.762
1170	7500	0.763	5.79	0.173	7500	0.763
1200	7500	0.765	5.82	0.172	7500	0.765
1260	7500	0.767	5.85	0.171	7500	0.767
1320	10000	0.773	5.93	0.169	10000	0.773
1380	12500	0.792	6.19	0.161	12500	0.792
1440	15000	0.799	6.30	0.159	15000	0.799
1500	12500	0.794	6.22	0.161	12500	0.794



Cell Count increases from 5000 to 10000

Difference in absorbance = (0.773 - 0.746) = 0.027

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	133	0.108	1.28	0.780	133	0.108
360	200	0.204	1.60	0.626	200	0.204
420	208	0.292	1.96	0.510	208	0.292
480	217	0.356	2.27	0.440	217	0.356
540	225	0.393	2.47	0.405	225	0.393
600	467	0.481	3.02	0.331	467	0.481
660	533	0.510	3.23	0.309	533	0.510
720	733	0.603	4.01	0.249	733	0.603
750	950	0.647	4.44	0.225	950	0.647
780	1117	0.684	4.83	0.207	1117	0.684
840	1933	0.704	5.05	0.198	1933	0.704
900	4133	0.717	5.21	0.192	4133	0.717
960	4933	0.736	5.45	0.184	4933	0.736
1020	5250	0.744	5.55	0.180	5250	0.744
1080	5400	0.751	5.64	0.177	5400	0.751
1140	5833	0.756	5.70	0.175	5833	0.756
1170	6667	0.760	5.76	0.174	6667	0.760
1200	7500	0.764	5.81	0.172	7500	0.764
1260	7500	0.764	5.81	0.172	7500	0.764
1320	8333	0.769	5.87	0.170	8333	0.769
1380	11667	0.781	6.04	0.165	11667	0.781
1440	13333	0.782	6.06	0.165	13333	0.782
1500	12500	0.778	6.00	0.167	12500	0.778

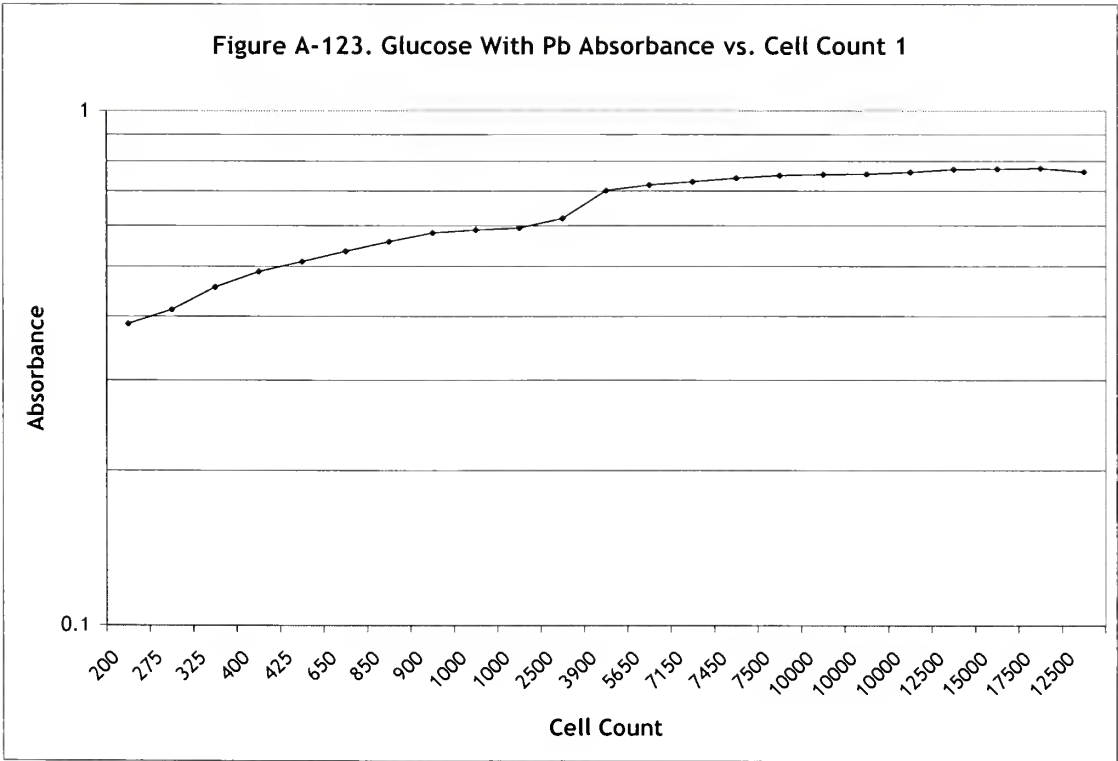


Cell Count increases from 6667 to 13333

Difference in absorbance = (0.782 - 0.760) = 0.022

GLUCOSE WITH Pb CELL COUNT

Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.387	2.44	0.410	200	0.387
360	275	0.412	2.58	0.387	275	0.412
420	325	0.456	2.86	0.350	325	0.456
480	400	0.489	3.08	0.324	400	0.489
540	425	0.511	3.24	0.308	425	0.511
600	650	0.535	3.43	0.292	650	0.535
660	850	0.559	3.62	0.276	850	0.559
720	900	0.581	3.81	0.262	900	0.581
750	1000	0.589	3.88	0.258	1000	0.589
780	1000	0.594	3.93	0.255	1000	0.594
840	2500	0.620	4.17	0.240	2500	0.620
900	3900	0.702	5.04	0.198	3900	0.702
960	5650	0.719	5.24	0.191	5650	0.719
1020	7150	0.730	5.37	0.186	7150	0.730
1080	7450	0.742	5.52	0.181	7450	0.742
1140	7500	0.751	5.63	0.178	7500	0.751
1170	10000	0.754	5.68	0.176	10000	0.754
1200	10000	0.755	5.69	0.176	10000	0.755
1260	10000	0.761	5.76	0.174	10000	0.761
1320	12500	0.771	5.90	0.170	12500	0.771
1380	15000	0.772	5.92	0.169	15000	0.772
1440	17500	0.774	5.94	0.168	17500	0.774
1500	12500	0.762	5.78	0.173	12500	0.762

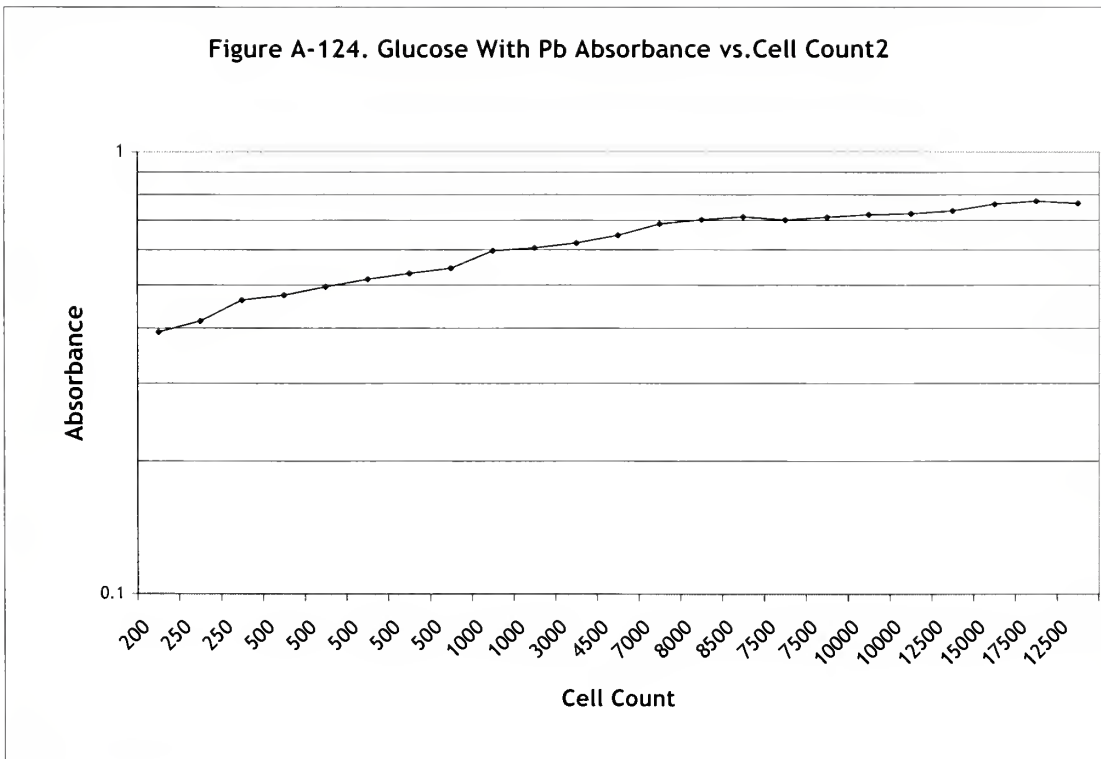


Cell Count increases from 425 to 850

Difference in absorbance = (0.559 - 0.511) = 0.048

Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.392	2.47	0.406	200	0.392
360	250	0.415	2.60	0.385	250	0.415
420	250	0.463	2.90	0.344	250	0.463
480	500	0.475	2.99	0.335	500	0.475
540	500	0.496	3.13	0.319	500	0.496
600	500	0.515	3.27	0.305	500	0.515
660	500	0.531	3.40	0.294	500	0.531
720	500	0.546	3.52	0.284	500	0.546
750	1000	0.597	3.95	0.253	1000	0.597
780	1000	0.606	4.04	0.248	1000	0.606
840	3000	0.622	4.19	0.239	3000	0.622
900	4500	0.648	4.45	0.225	4500	0.648
960	7000	0.687	4.86	0.206	7000	0.687
1020	8000	0.702	5.04	0.199	8000	0.702
1080	8500	0.712	5.15	0.194	8500	0.712
1140	7500	0.701	5.02	0.199	7500	0.701
1170	7500	0.711	5.14	0.195	7500	0.711
1200	10000	0.721	5.26	0.190	10000	0.721
1260	10000	0.724	5.30	0.189	10000	0.724
1320	12500	0.736	5.45	0.184	12500	0.736
1380	15000	0.762	5.78	0.173	15000	0.762
1440	17500	0.774	5.94	0.168	17500	0.774
1500	12500	0.765	5.82	0.172	12500	0.765

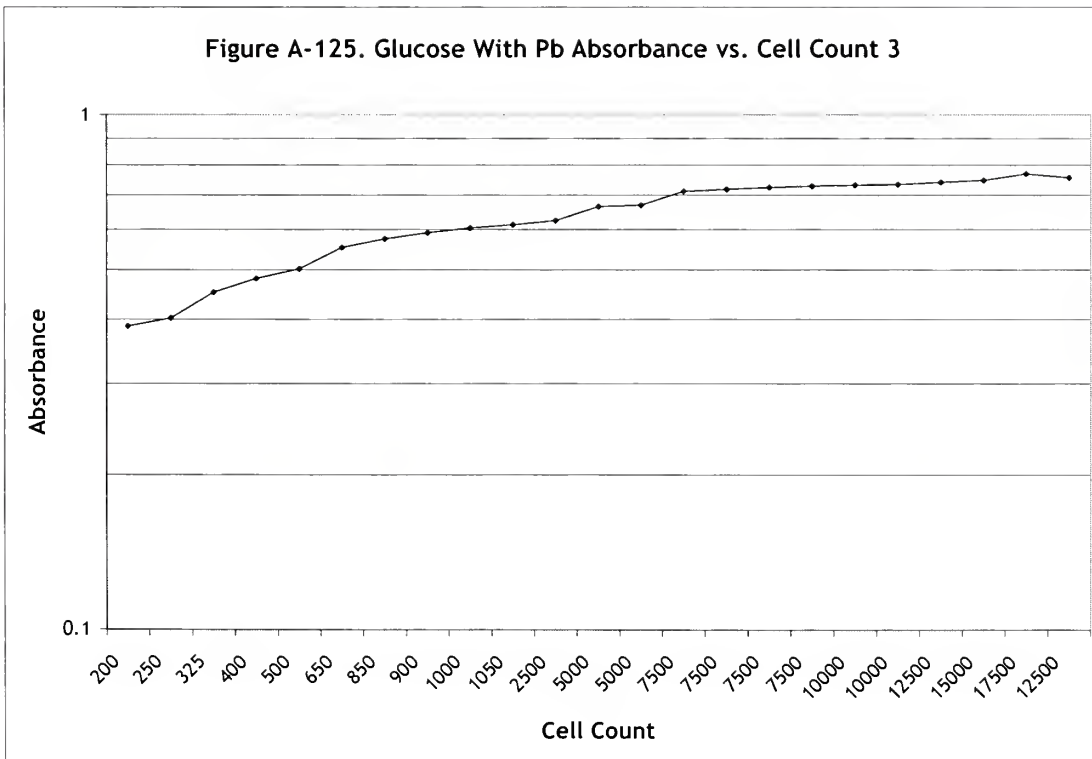
Figure A-124. Glucose With Pb Absorbance vs. Cell Count2



Cell Count increases from 7500 to 15000

Difference in absorbance = (0.762 - 0.711) = 0.051

Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.388	2.44	0.409	200	0.388
360	250	0.402	2.52	0.396	250	0.402
420	325	0.452	2.83	0.353	325	0.452
480	400	0.481	3.03	0.330	400	0.481
540	500	0.502	3.18	0.315	500	0.502
600	650	0.553	3.57	0.280	650	0.553
660	850	0.575	3.76	0.266	850	0.575
720	900	0.591	3.90	0.256	900	0.591
750	1000	0.604	4.02	0.249	1000	0.604
780	1050	0.613	4.10	0.244	1050	0.613
840	2500	0.624	4.21	0.238	2500	0.624
900	5000	0.665	4.62	0.216	5000	0.665
960	5000	0.669	4.67	0.214	5000	0.669
1020	7500	0.712	5.15	0.194	7500	0.712
1080	7500	0.718	5.22	0.191	7500	0.718
1140	7500	0.724	5.30	0.189	7500	0.724
1170	7500	0.728	5.35	0.187	7500	0.728
1200	10000	0.732	5.40	0.185	10000	0.732
1260	10000	0.734	5.42	0.185	10000	0.734
1320	12500	0.741	5.51	0.182	12500	0.741
1380	15000	0.748	5.60	0.179	15000	0.748
1440	17500	0.769	5.87	0.170	17500	0.769
1500	12500	0.756	5.70	0.175	12500	0.756

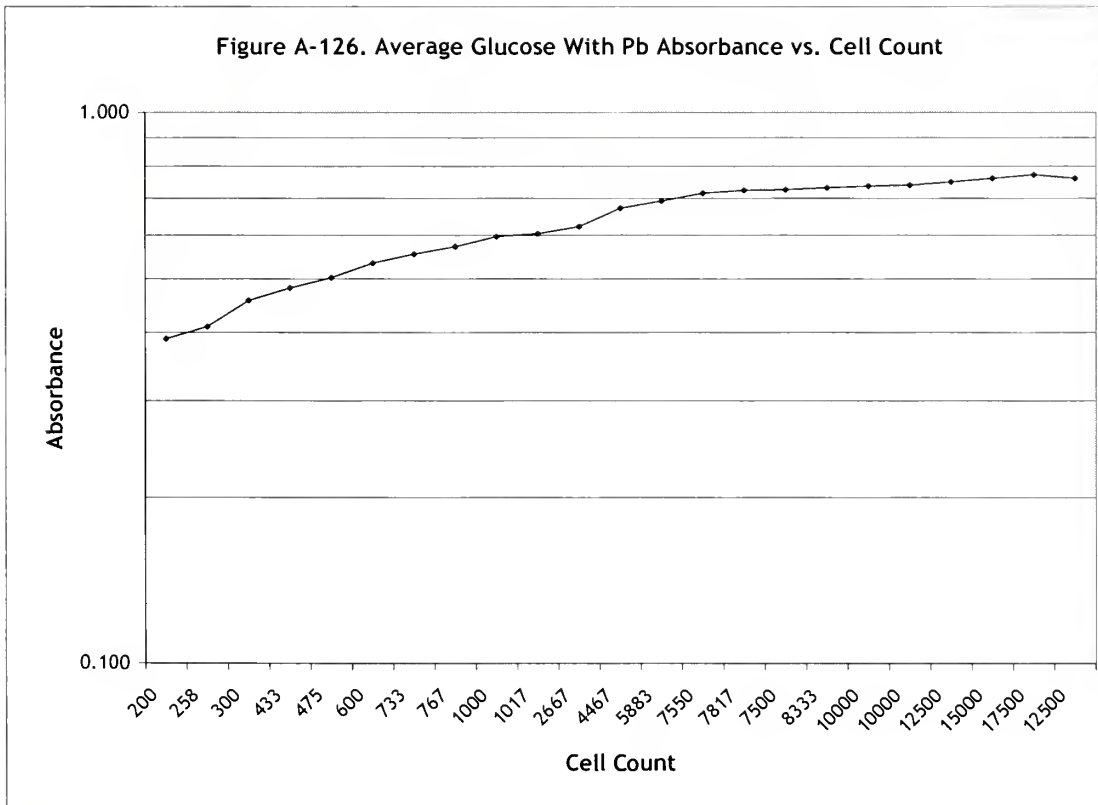


Cell Count increases from 5000 to 10000

Difference in absorbance = (0.732 - 0.669) = 0.063

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
300	200	0.389	2.45	0.408	200	0.389
360	258	0.410	2.57	0.389	258	0.410
420	300	0.457	2.86	0.349	300	0.457
480	433	0.482	3.03	0.330	433	0.482
540	475	0.503	3.18	0.314	475	0.503
600	600	0.534	3.42	0.292	600	0.534
660	733	0.555	3.59	0.279	733	0.555
720	767	0.573	3.74	0.268	767	0.573
750	1000	0.597	3.95	0.253	1000	0.597
780	1017	0.604	4.02	0.249	1017	0.604
840	2667	0.622	4.19	0.239	2667	0.622
900	4467	0.672	4.70	0.213	4467	0.672
960	5883	0.692	4.92	0.203	5883	0.692
1020	7550	0.715	5.19	0.193	7550	0.715
1080	7817	0.724	5.30	0.189	7817	0.724
1140	7500	0.725	5.31	0.188	7500	0.725
1170	8333	0.731	5.38	0.186	8333	0.731
1200	10000	0.736	5.45	0.184	10000	0.736
1260	10000	0.740	5.49	0.182	10000	0.740
1320	12500	0.749	5.61	0.178	12500	0.749
1380	15000	0.761	5.76	0.173	15000	0.761
1440	17500	0.772	5.92	0.169	17500	0.772
1500	12500	0.761	5.77	0.173	12500	0.761

Figure A-126. Average Glucose With Pb Absorbance vs. Cell Count



Cell Count increases from 300 to 600

Difference in absorbance = (0.534 - 0.457) = 0.077

APPENDIX B

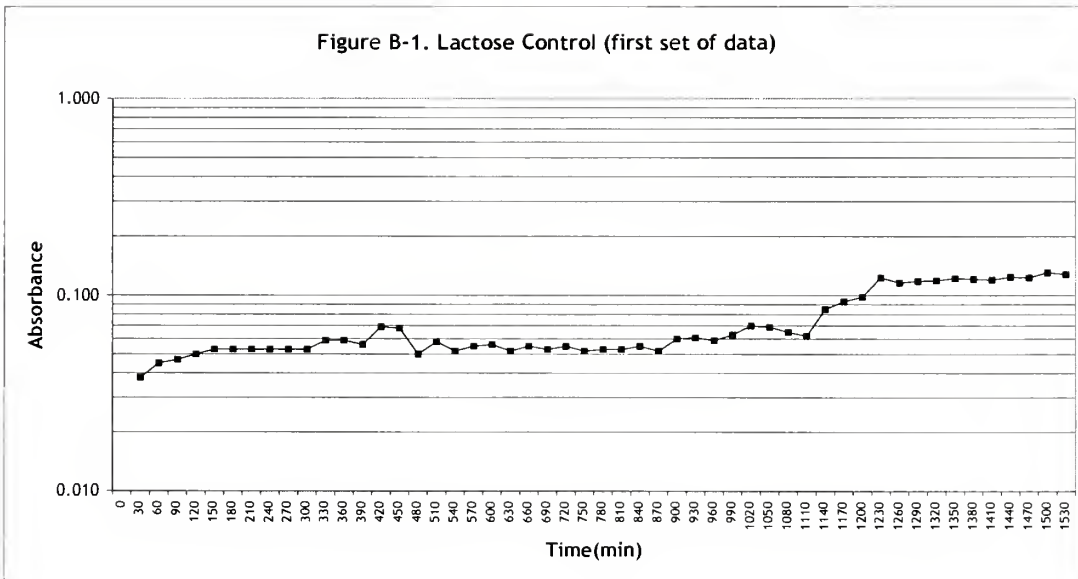
All of the doubling times for *M.roseus* are presented in APPENDIX B

Micrococcus roseus

Lactose control

Time	Run1	Run2	Run3	Average
0	0.000	0.000	0.000	0.000
30	0.038	0.039	0.037	0.038
60	0.045	0.043	0.046	0.045
90	0.047	0.043	0.043	0.044
120	0.050	0.043	0.046	0.046
150	0.053	0.044	0.045	0.047
180	0.053	0.049	0.048	0.050
210	0.053	0.065	0.049	0.056
240	0.053	0.066	0.051	0.057
270	0.053	0.065	0.053	0.057
300	0.053	0.065	0.052	0.057
330	0.059	0.066	0.056	0.060
360	0.059	0.065	0.055	0.060
390	0.056	0.071	0.059	0.062
420	0.069	0.076	0.058	0.068
450	0.068	0.075	0.061	0.068
480	0.050	0.072	0.058	0.060
510	0.058	0.057	0.054	0.056
540	0.052	0.053	0.056	0.054
570	0.055	0.059	0.059	0.058
600	0.056	0.068	0.061	0.062
630	0.052	0.065	0.063	0.060
660	0.055	0.066	0.064	0.062
690	0.053	0.069	0.059	0.060
720	0.055	0.065	0.062	0.061
750	0.052	0.060	0.060	0.057
780	0.053	0.059	0.059	0.057
810	0.053	0.060	0.058	0.057
840	0.055	0.061	0.061	0.059
870	0.052	0.060	0.060	0.057
900	0.060	0.065	0.063	0.063
930	0.061	0.060	0.061	0.061
960	0.059	0.067	0.058	0.061
990	0.063	0.065	0.059	0.062
1020	0.070	0.068	0.055	0.064
1050	0.069	0.067	0.058	0.065
1080	0.065	0.073	0.061	0.066
1110	0.062	0.066	0.065	0.064
1140	0.085	0.063	0.069	0.072
1170	0.093	0.086	0.076	0.085
1200	0.098	0.097	0.088	0.094
1230	0.123	0.128	0.126	0.126
1260	0.116	0.127	0.130	0.124
1290	0.118	0.121	0.132	0.124
1320	0.119	0.127	0.134	0.127
1350	0.122	0.125	0.135	0.127
1380	0.121	0.123	0.137	0.127
1410	0.120	0.125	0.136	0.127
1440	0.124	0.135	0.135	0.131
1470	0.123	0.131	0.139	0.131
1500	0.131	0.133	0.140	0.135
1530	0.128	0.129	0.142	0.133

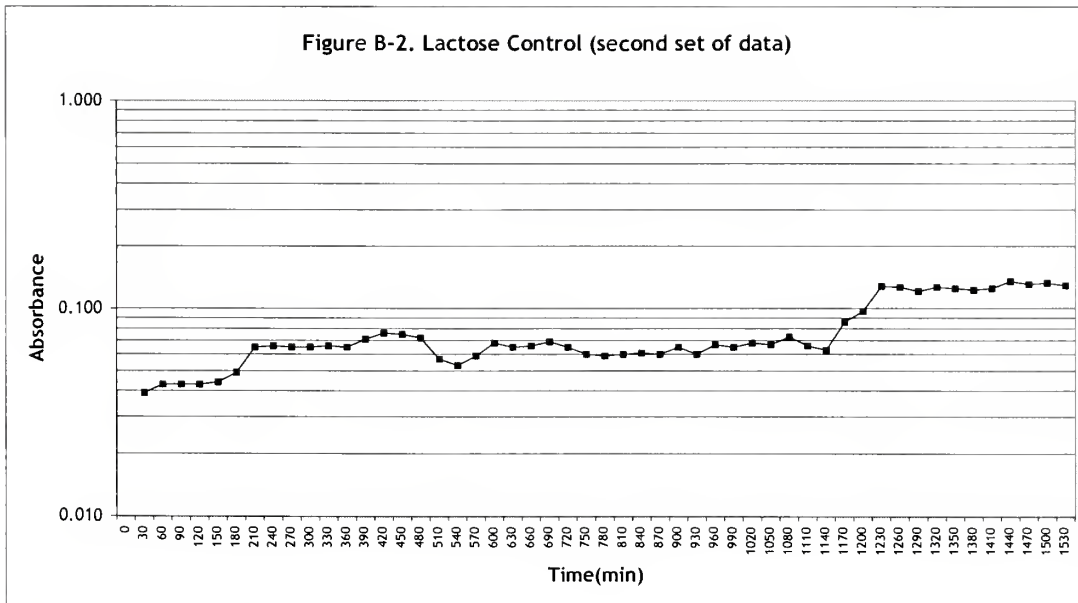
Figure B-1. Lactose Control (first set of data)



Absorbance increases from 0.062 to 0.123

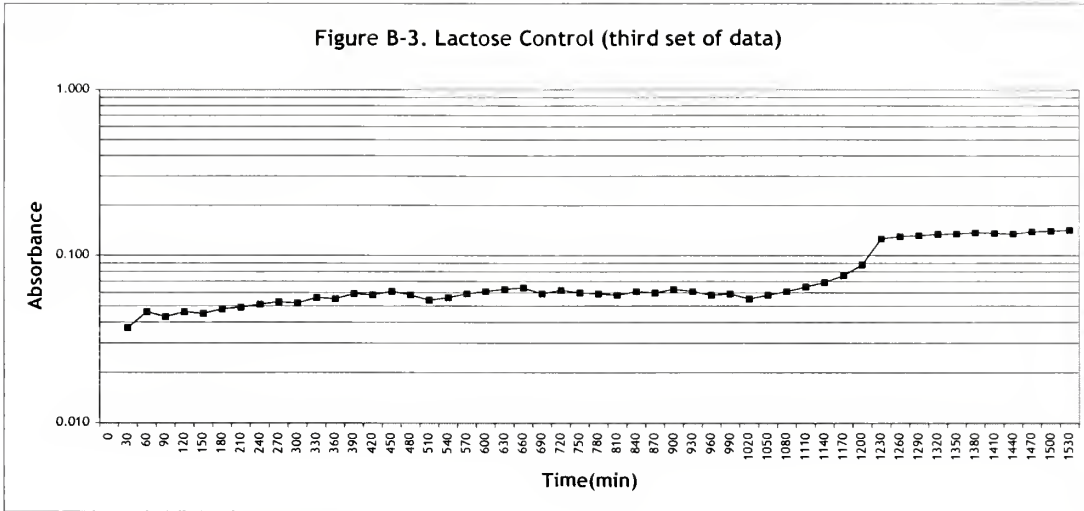
Actual growth rate = (1230 min - 1100 min) = 120 min

Figure B-2. Lactose Control (second set of data)



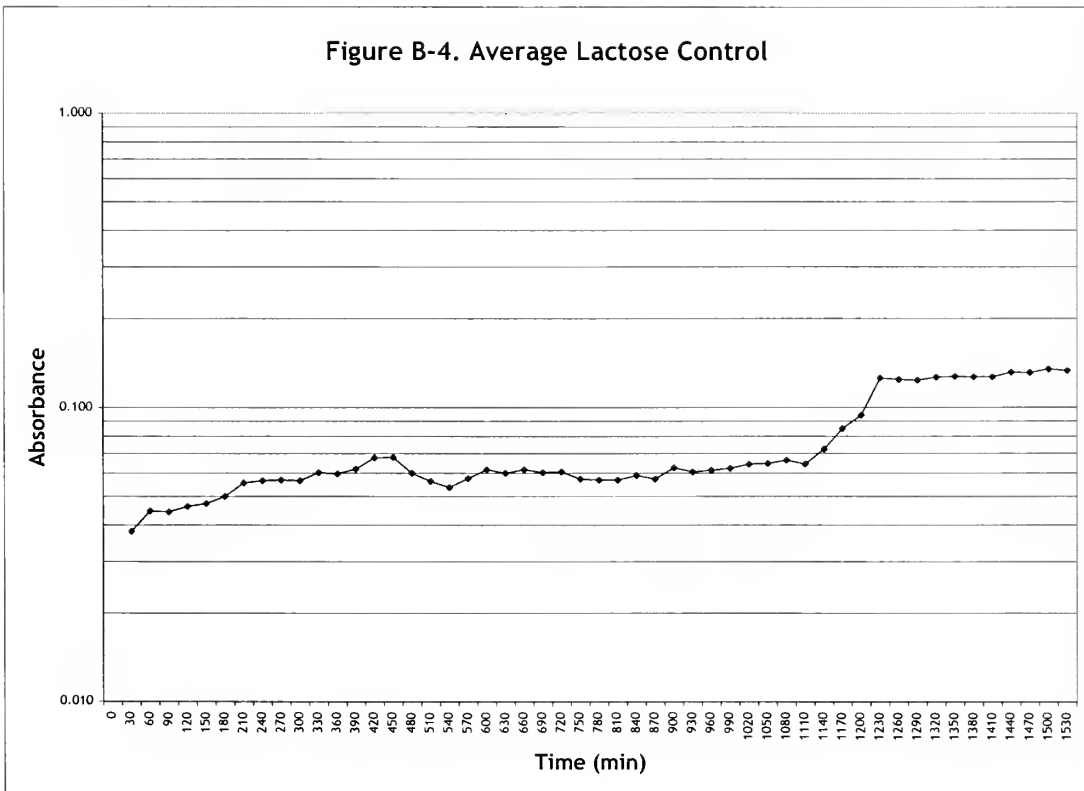
Absorbance increases from 0.063 to 0.127

Actual growth rate = (1260 min - 1140 min) = 120 min



Absorbance increases from 0.065 to 0.130

Actual growth rate = (1260 min - 1110 min) = 150 min



Absorbance increases from 0.064 to 0.126

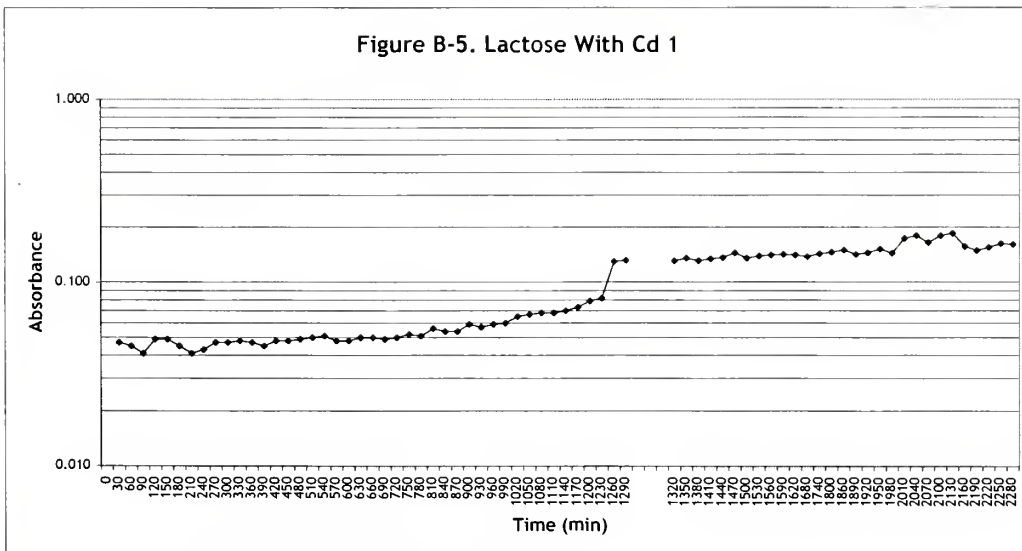
Actual growth rate = (1230 min - 1110 min) = 120 min

Micrococcus roseus

Lactose With Cadmium

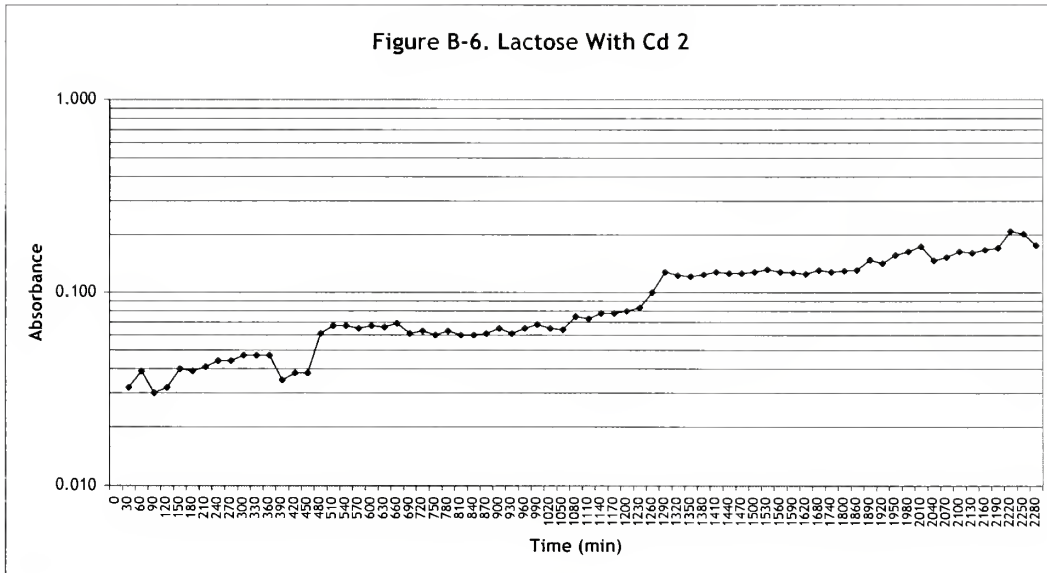
Time (min)	Run 1	Run 2	Average
0	0.000	0.000	0.000
30	0.047	0.032	0.040
60	0.045	0.039	0.042
90	0.041	0.030	0.036
120	0.049	0.032	0.041
150	0.049	0.040	0.045
180	0.045	0.039	0.042
210	0.041	0.041	0.041
240	0.043	0.044	0.044
270	0.047	0.044	0.046
300	0.047	0.047	0.047
330	0.048	0.047	0.048
360	0.047	0.047	0.047
390	0.045	0.035	0.040
420	0.048	0.038	0.043
450	0.048	0.038	0.043
480	0.049	0.061	0.055
510	0.050	0.067	0.059
540	0.051	0.067	0.059
570	0.048	0.065	0.057
600	0.048	0.067	0.058
630	0.050	0.066	0.058
660	0.050	0.069	0.060
690	0.049	0.061	0.055
720	0.050	0.063	0.057
750	0.052	0.060	0.056
780	0.051	0.063	0.057
810	0.056	0.060	0.058
840	0.054	0.060	0.057
870	0.054	0.061	0.058
900	0.059	0.065	0.062
930	0.057	0.061	0.059
960	0.059	0.065	0.062
990	0.060	0.068	0.064
1020	0.065	0.065	0.065
1050	0.067	0.064	0.066
1080	0.068	0.075	0.072
1110	0.068	0.073	0.071
1140	0.070	0.078	0.074
1170	0.073	0.078	0.076
1200	0.079	0.080	0.080
1230	0.082	0.083	0.083
1260	0.130	0.100	0.115
1290	0.132	0.128	0.130

1320	0.131	0.123	0.127
1350	0.135	0.121	0.128
1380	0.131	0.124	0.128
1410	0.134	0.128	0.131
1440	0.136	0.126	0.131
1470	0.145	0.126	0.136
1500	0.135	0.128	0.132
1530	0.139	0.132	0.136
1560	0.141	0.128	0.135
1590	0.142	0.127	0.135
1620	0.141	0.125	0.133
1680	0.138	0.131	0.135
1740	0.143	0.128	0.136
1800	0.146	0.130	0.138
1860	0.150	0.131	0.141
1890	0.142	0.148	0.145
1920	0.145	0.142	0.144
1950	0.152	0.157	0.155
1980	0.144	0.164	0.154
2010	0.174	0.174	0.174
2040	0.180	0.147	0.164
2070	0.165	0.153	0.159
2100	0.180	0.164	0.172
2130	0.185	0.161	0.173
2160	0.157	0.167	0.162
2190	0.149	0.171	0.160
2220	0.155	0.209	0.182
2250	0.163	0.202	0.183
2280	0.161	0.176	0.169



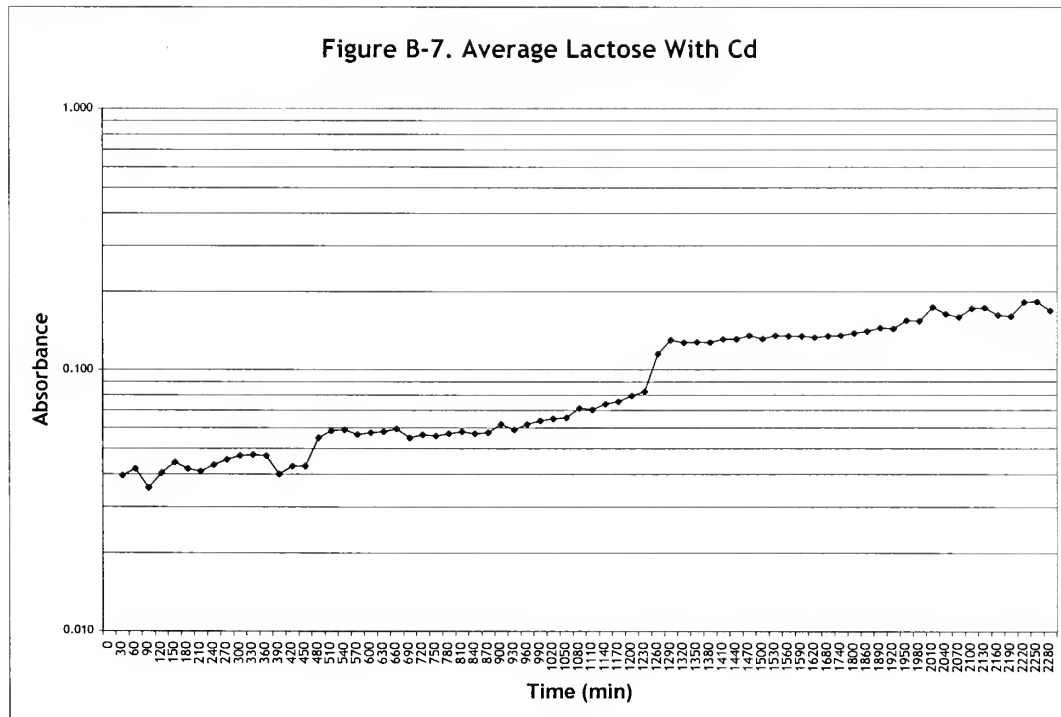
Absorbance increases from 0.065 to 0.130

Actual growth = (1260 min - 1020 min) = 240 min



Absorbance increases from 0.064 to 0.128

Actual growth = (1290 min - 1050 min) = 240 min

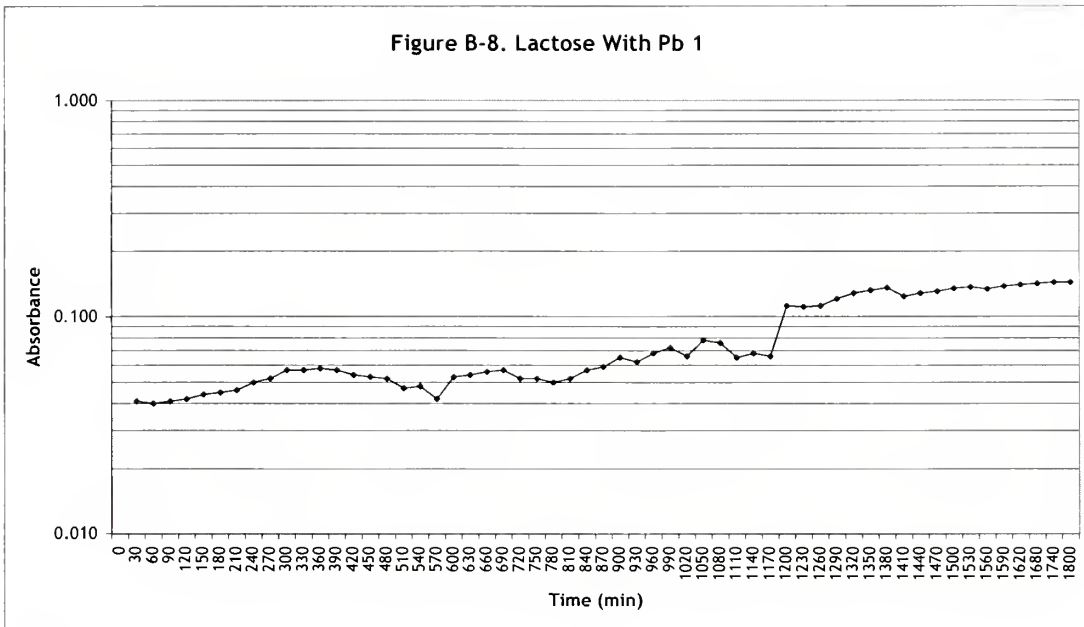


Absorbance increases from 0.065 to 0.130

Actual growth = (1290 min - 1020 min) = 270 min

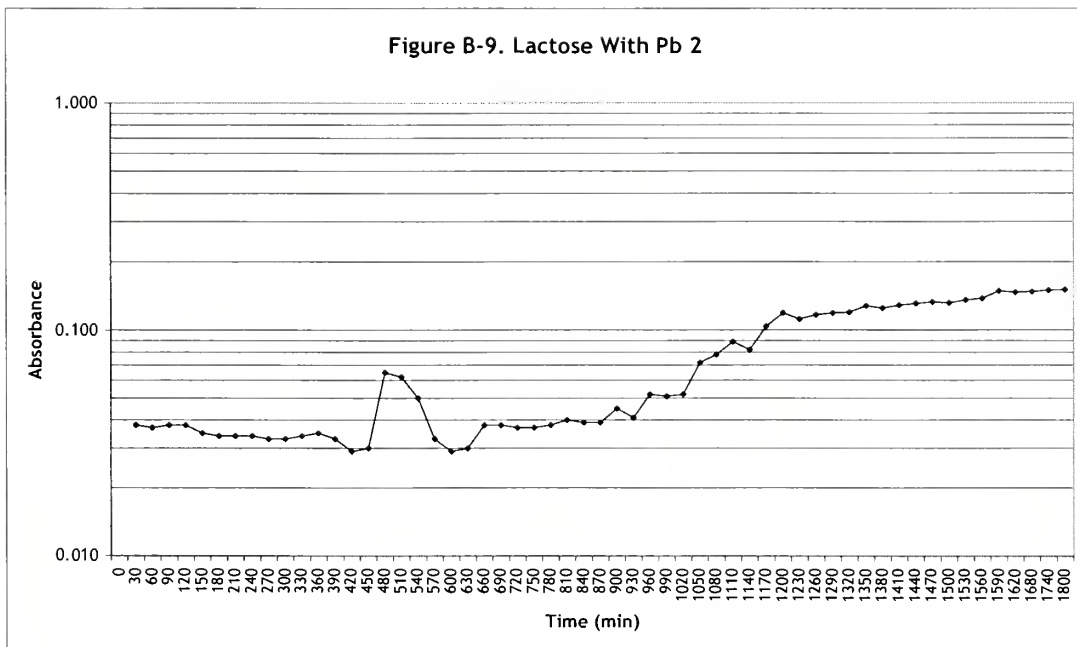
Lactose With Lead (*Micrococcus roseus*)

Time (min)	Run 1	Run 2	Run 3	Average
0	0.000	0.000	0.000	0.000
30	0.041	0.038	0.036	0.038
60	0.040	0.037	0.037	0.038
90	0.041	0.038	0.038	0.039
120	0.042	0.038	0.036	0.039
150	0.044	0.035	0.035	0.038
180	0.045	0.034	0.033	0.037
210	0.046	0.034	0.034	0.038
240	0.050	0.034	0.036	0.040
270	0.052	0.033	0.038	0.041
300	0.057	0.033	0.035	0.042
330	0.057	0.034	0.033	0.041
360	0.058	0.035	0.035	0.043
390	0.057	0.033	0.033	0.041
420	0.054	0.029	0.038	0.040
450	0.053	0.030	0.046	0.043
480	0.052	0.065	0.053	0.057
510	0.047	0.062	0.059	0.056
540	0.048	0.050	0.051	0.050
570	0.042	0.033	0.048	0.041
600	0.053	0.029	0.045	0.042
630	0.054	0.030	0.043	0.042
660	0.056	0.038	0.041	0.045
690	0.057	0.038	0.048	0.048
720	0.052	0.037	0.045	0.045
750	0.052	0.037	0.041	0.043
780	0.050	0.038	0.043	0.044
810	0.052	0.040	0.044	0.045
840	0.057	0.039	0.046	0.047
870	0.059	0.039	0.041	0.046
900	0.065	0.045	0.045	0.052
930	0.062	0.041	0.049	0.051
960	0.068	0.052	0.051	0.057
990	0.072	0.051	0.052	0.058
1020	0.066	0.052	0.051	0.056
1050	0.078	0.072	0.073	0.074
1080	0.076	0.078	0.075	0.076
1110	0.065	0.089	0.084	0.079
1140	0.068	0.082	0.091	0.080
1170	0.066	0.104	0.102	0.091
1200	0.112	0.119	0.116	0.116
1230	0.111	0.112	0.118	0.114
1260	0.112	0.117	0.117	0.115
1290	0.121	0.119	0.119	0.120
1320	0.128	0.120	0.120	0.123
1350	0.132	0.128	0.122	0.127
1380	0.136	0.125	0.124	0.128
1410	0.124	0.129	0.129	0.127
1440	0.128	0.131	0.131	0.130
1470	0.131	0.133	0.133	0.132
1500	0.135	0.132	0.132	0.133
1530	0.137	0.136	0.135	0.136
1560	0.134	0.138	0.138	0.137
1590	0.138	0.149	0.141	0.143
1620	0.140	0.147	0.144	0.144
1680	0.142	0.148	0.146	0.145
1740	0.144	0.150	0.149	0.148
1800	0.144	0.151	0.151	0.149



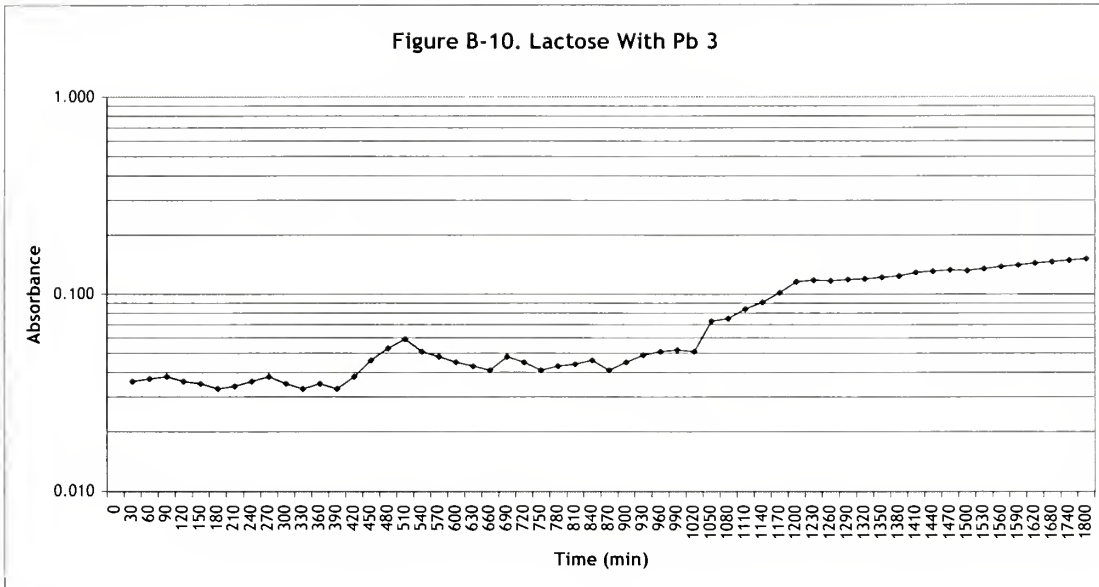
Absorbance increases from 0.068 to 0.136

Actual growth rate = (1380 min - 1140 min) = 240 min



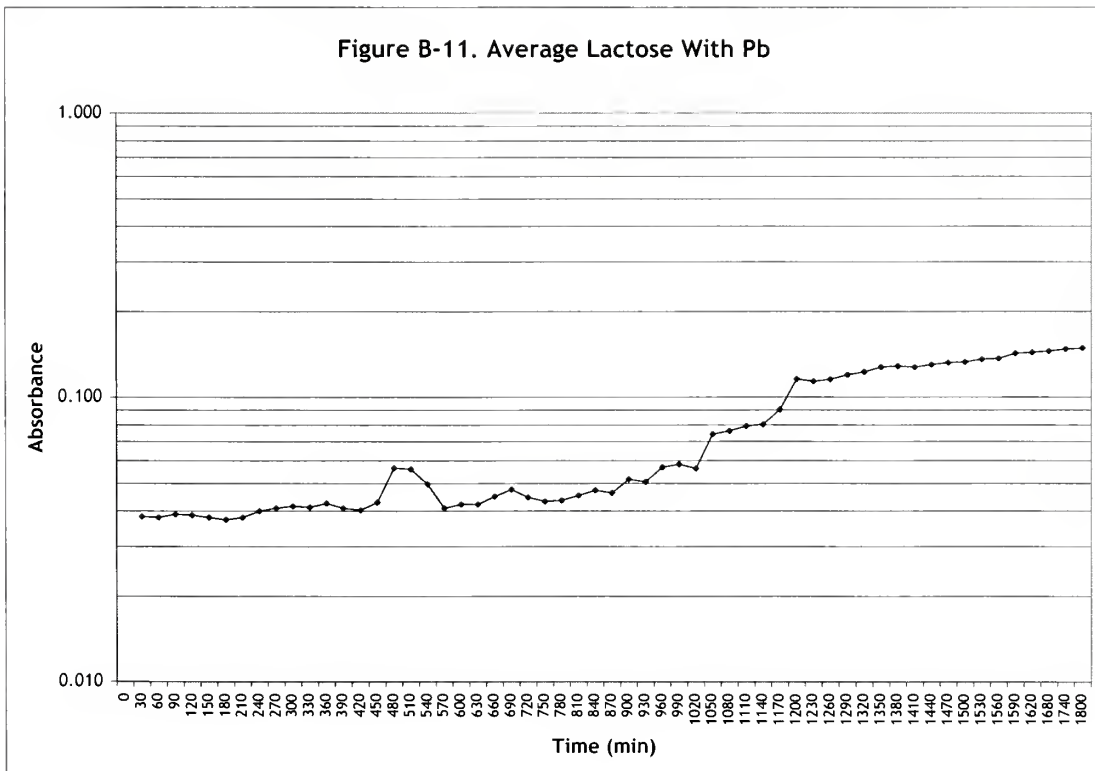
Absorbance increases from 0.052 to 0.104

Actual growth rate = (1170 min - 960 min) = 210 min



Absorbance increases from 0.051 to 0.102

Actual growth rate = (1170 min - 960 min) = 210 min



Absorbance increases from 0.058 to 0.116

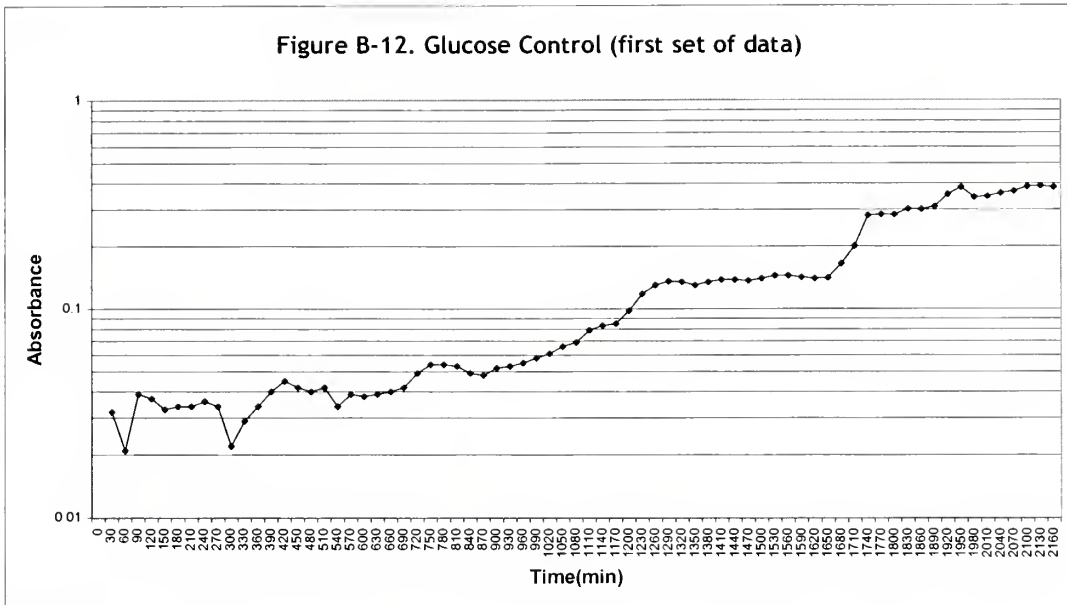
Actual growth rate = (1200 min - 990 min) = 210 min

Micrococcus roseus

Glucose control

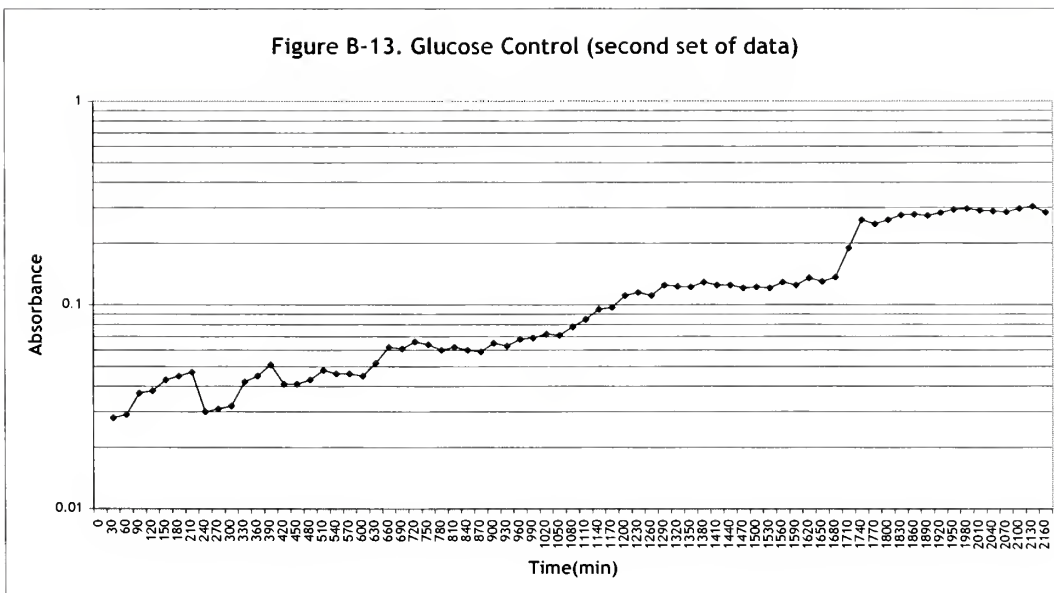
Time(min)	Run1	Run2	Run3	Average
0	0	0	0	0.000
30	0.032	0.028	0.026	0.029
60	0.021	0.029	0.038	0.029
90	0.039	0.037	0.041	0.039
120	0.037	0.038	0.044	0.040
150	0.033	0.043	0.046	0.041
180	0.034	0.045	0.044	0.041
210	0.034	0.047	0.049	0.043
240	0.036	0.030	0.047	0.038
270	0.034	0.031	0.042	0.036
300	0.022	0.032	0.041	0.032
330	0.029	0.042	0.044	0.038
360	0.034	0.045	0.046	0.042
390	0.040	0.051	0.049	0.047
420	0.045	0.041	0.051	0.046
450	0.042	0.041	0.05	0.044
480	0.040	0.043	0.052	0.045
510	0.042	0.048	0.049	0.046
540	0.034	0.046	0.053	0.044
570	0.039	0.046	0.051	0.045
600	0.038	0.045	0.049	0.044
630	0.039	0.052	0.054	0.048
660	0.040	0.062	0.056	0.053
690	0.042	0.061	0.059	0.054
720	0.049	0.066	0.066	0.060
750	0.054	0.064	0.068	0.062
780	0.054	0.060	0.066	0.060
810	0.053	0.062	0.067	0.061
840	0.049	0.060	0.071	0.060
870	0.048	0.059	0.068	0.058
900	0.052	0.065	0.072	0.063
930	0.053	0.063	0.071	0.062
960	0.055	0.068	0.074	0.066
990	0.058	0.069	0.076	0.068
1020	0.061	0.072	0.078	0.070
1050	0.066	0.071	0.08	0.072
1080	0.069	0.078	0.081	0.076
1110	0.079	0.085	0.085	0.083
1140	0.083	0.095	0.096	0.091
1170	0.085	0.097	0.092	0.091
1200	0.098	0.111	0.108	0.106
1230	0.118	0.115	0.119	0.117
1260	0.130	0.111	0.124	0.122
1290	0.136	0.125	0.131	0.131
1320	0.135	0.123	0.134	0.131
1350	0.130	0.122	0.138	0.130
1380	0.135	0.129	0.141	0.135
1410	0.138	0.125	0.143	0.135
1440	0.138	0.125	0.144	0.136
1470	0.137	0.121	0.148	0.135

1500	0.140	0.122	0.146	0.136
1530	0.145	0.121	0.151	0.139
1560	0.145	0.129	0.152	0.142
1590	0.142	0.125	0.154	0.140
1620	0.140	0.135	0.156	0.144
1650	0.141	0.130	0.152	0.141
1680	0.165	0.136	0.158	0.153
1710	0.201	0.190	0.211	0.201
1740	0.251	0.222	0.248	0.240
1770	0.265	0.249	0.267	0.260
1800	0.282	0.261	0.302	0.282
1830	0.303	0.276	0.314	0.298
1860	0.301	0.278	0.321	0.300
1890	0.310	0.274	0.326	0.303
1920	0.355	0.283	0.348	0.329
1950	0.384	0.294	0.361	0.346
1980	0.344	0.297	0.365	0.335
2010	0.347	0.290	0.361	0.333
2040	0.360	0.288	0.369	0.339
2070	0.368	0.285	0.372	0.342
2100	0.386	0.297	0.383	0.355
2130	0.390	0.304	0.392	0.362
2160	0.384	0.284	0.391	0.353



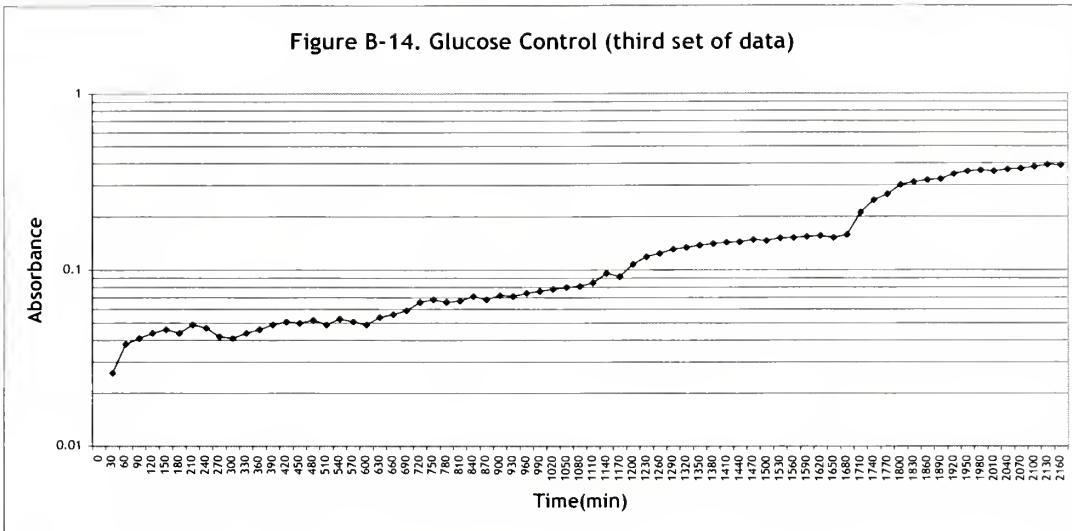
Absorbance increases from 0.141 to 0.282

Actual growth rate = (1800 min - 1650 min) = 150 min



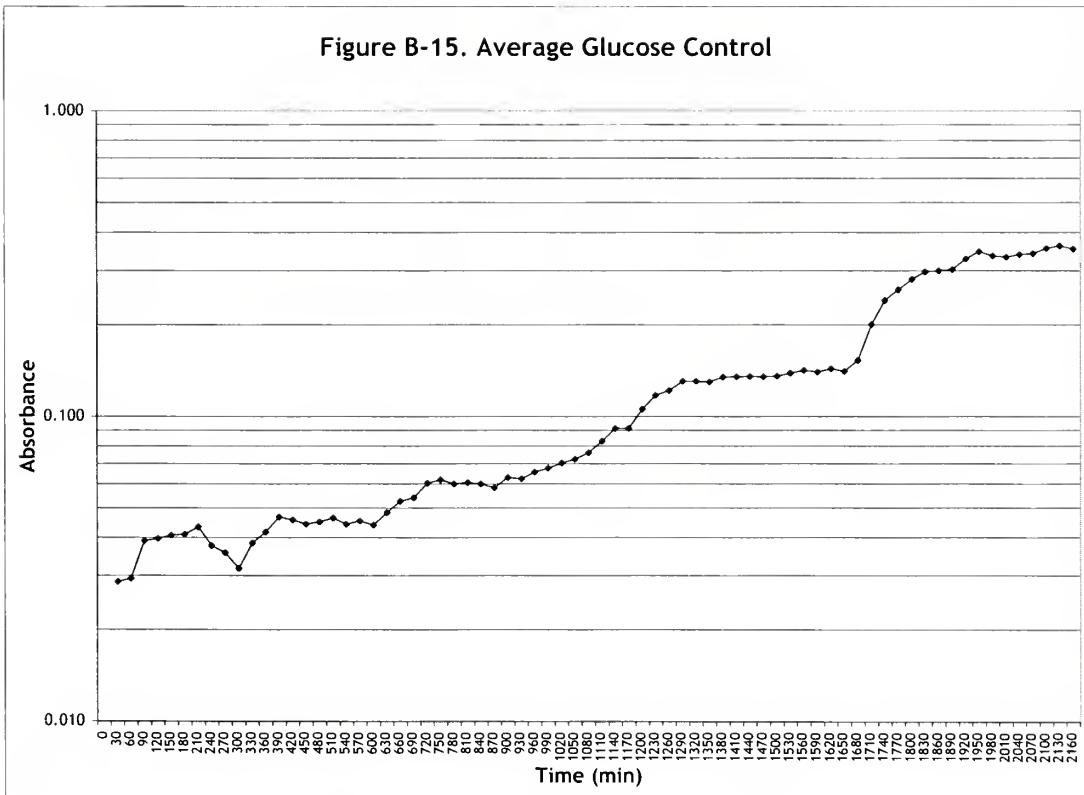
Absorbance increases from 0.130 to 0.261

Actual growth rate = (1800 min - 1650 min) = 150 min



Absorbance increases from 0.158 to 0.314

Actual growth rate = (1830 min - 1680 min) = 150 min



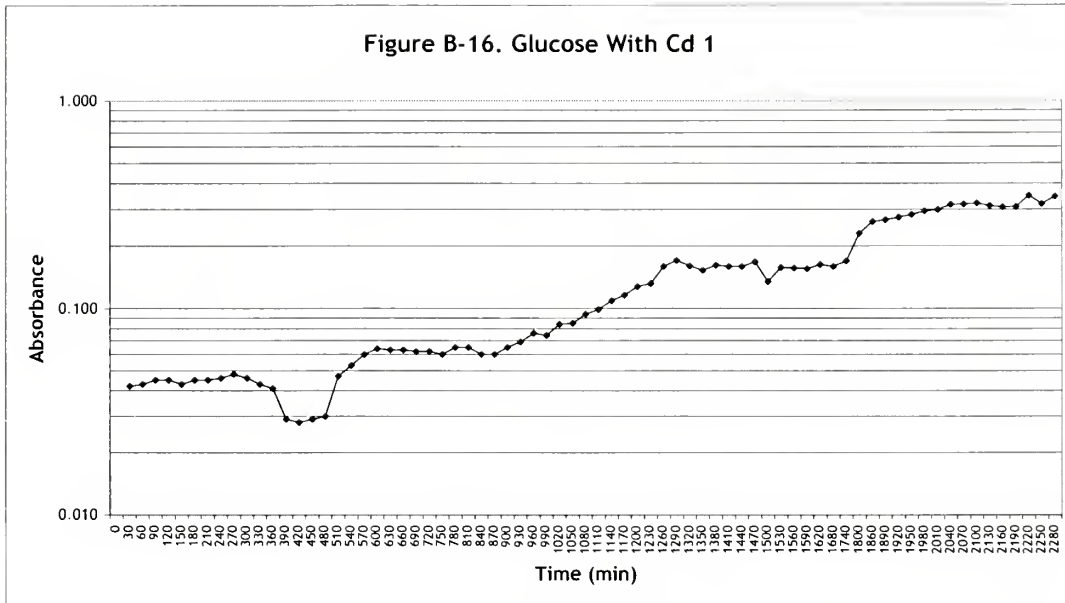
Absorbance increases from 0.141 to 0.282

Actual growth rate = (1800 min - 1650 min) = 150 min

Glucose and Cd at concentration of $1 \cdot 10^{-3} \text{M}$

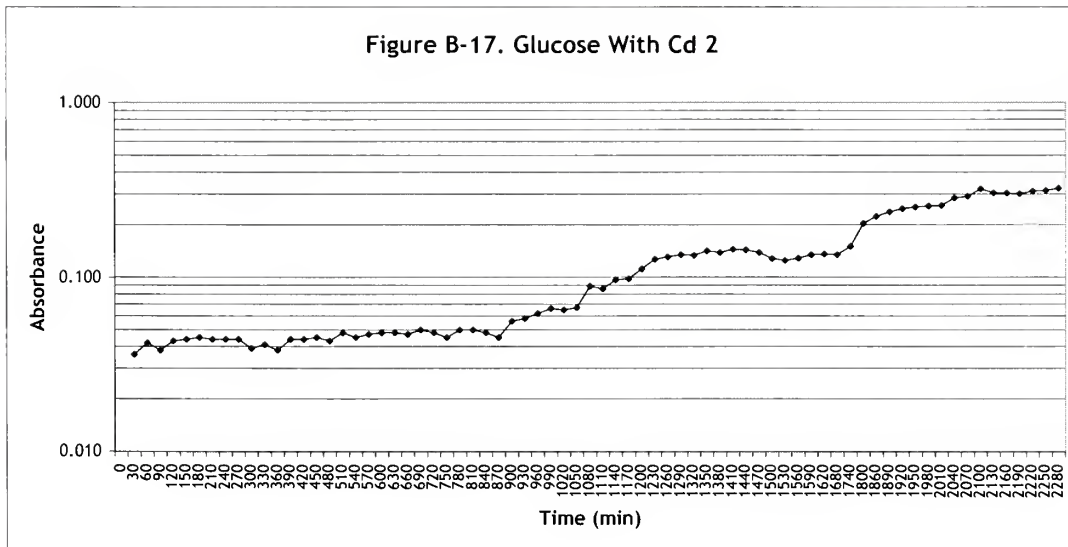
Time (min)	Run 1	Run 2	Run3	Average
0	0.000	0.000	0.000	0.000
30	0.042	0.036	0.032	0.037
60	0.043	0.042	0.038	0.041
90	0.045	0.038	0.041	0.041
120	0.045	0.043	0.044	0.044
150	0.043	0.044	0.046	0.044
180	0.045	0.045	0.047	0.046
210	0.045	0.044	0.045	0.045
240	0.046	0.044	0.046	0.045
270	0.048	0.044	0.043	0.045
300	0.046	0.039	0.049	0.045
330	0.043	0.041	0.046	0.043
360	0.041	0.038	0.044	0.041
390	0.029	0.044	0.042	0.038
420	0.028	0.044	0.046	0.039
450	0.029	0.045	0.048	0.041
480	0.030	0.043	0.045	0.039
510	0.047	0.048	0.051	0.049
540	0.053	0.045	0.053	0.050
570	0.060	0.047	0.056	0.054
600	0.064	0.048	0.059	0.057
630	0.063	0.048	0.061	0.057
660	0.063	0.047	0.063	0.058
690	0.062	0.050	0.065	0.059
720	0.062	0.048	0.068	0.059
750	0.060	0.045	0.071	0.059
780	0.065	0.050	0.069	0.061
810	0.065	0.050	0.073	0.063
840	0.060	0.048	0.071	0.060
870	0.060	0.045	0.075	0.060
900	0.065	0.056	0.078	0.066
930	0.069	0.058	0.075	0.067
960	0.076	0.062	0.078	0.072
990	0.074	0.066	0.076	0.072
1020	0.084	0.065	0.078	0.076
1050	0.085	0.067	0.081	0.078
1080	0.094	0.089	0.088	0.090
1110	0.099	0.086	0.097	0.094
1140	0.109	0.097	0.106	0.104
1170	0.116	0.098	0.112	0.109
1200	0.128	0.112	0.122	0.121
1230	0.132	0.127	0.128	0.129
1260	0.160	0.131	0.148	0.146
1290	0.171	0.135	0.161	0.156
1320	0.161	0.134	0.168	0.154
1350	0.153	0.142	0.171	0.155
1380	0.162	0.139	0.174	0.158
1410	0.160	0.145	0.176	0.160
1440	0.160	0.144	0.174	0.159
1470	0.168	0.139	0.178	0.162
1500	0.135	0.128	0.174	0.146
1530	0.158	0.125	0.176	0.153
1560	0.157	0.129	0.178	0.155

1590	0.156	0.135	0.181	0.157
1620	0.163	0.136	0.183	0.161
1680	0.160	0.135	0.185	0.160
1740	0.170	0.151	0.181	0.167
1800	0.230	0.204	0.218	0.217
1860	0.263	0.224	0.242	0.243
1890	0.268	0.238	0.261	0.256
1920	0.276	0.248	0.272	0.265
1950	0.284	0.253	0.281	0.273
1980	0.295	0.256	0.294	0.282
2010	0.299	0.259	0.296	0.285
2040	0.317	0.285	0.312	0.305
2070	0.318	0.291	0.317	0.309
2100	0.322	0.321	0.324	0.322
2130	0.312	0.305	0.321	0.313
2160	0.308	0.304	0.319	0.310
2190	0.309	0.301	0.325	0.312
2220	0.351	0.312	0.341	0.335
2250	0.319	0.315	0.348	0.327
2280	0.347	0.324	0.346	0.339



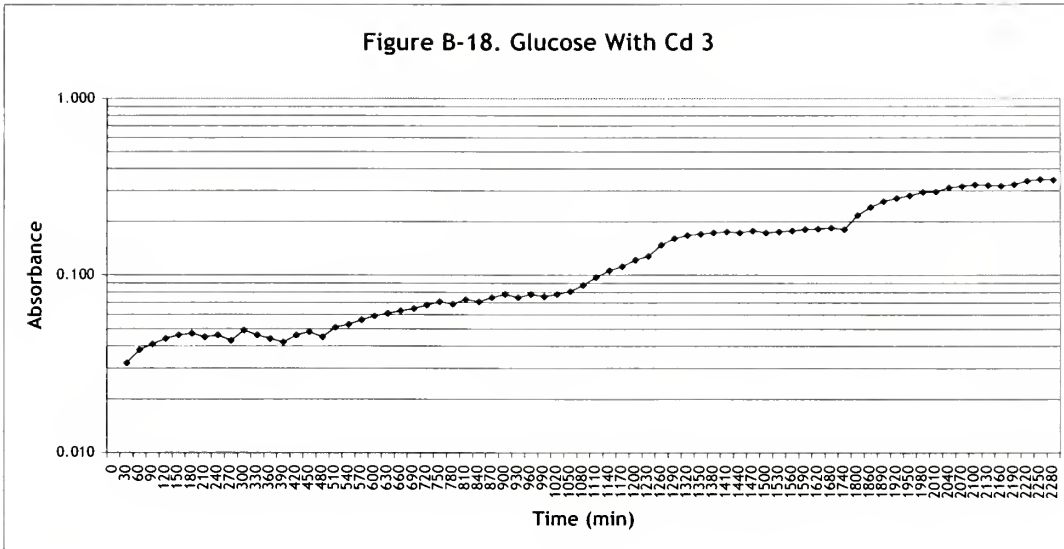
Absorbance increases from 0.085 to 0.171

Actual growth rate = (1290 min - 1050 min) = 240 min



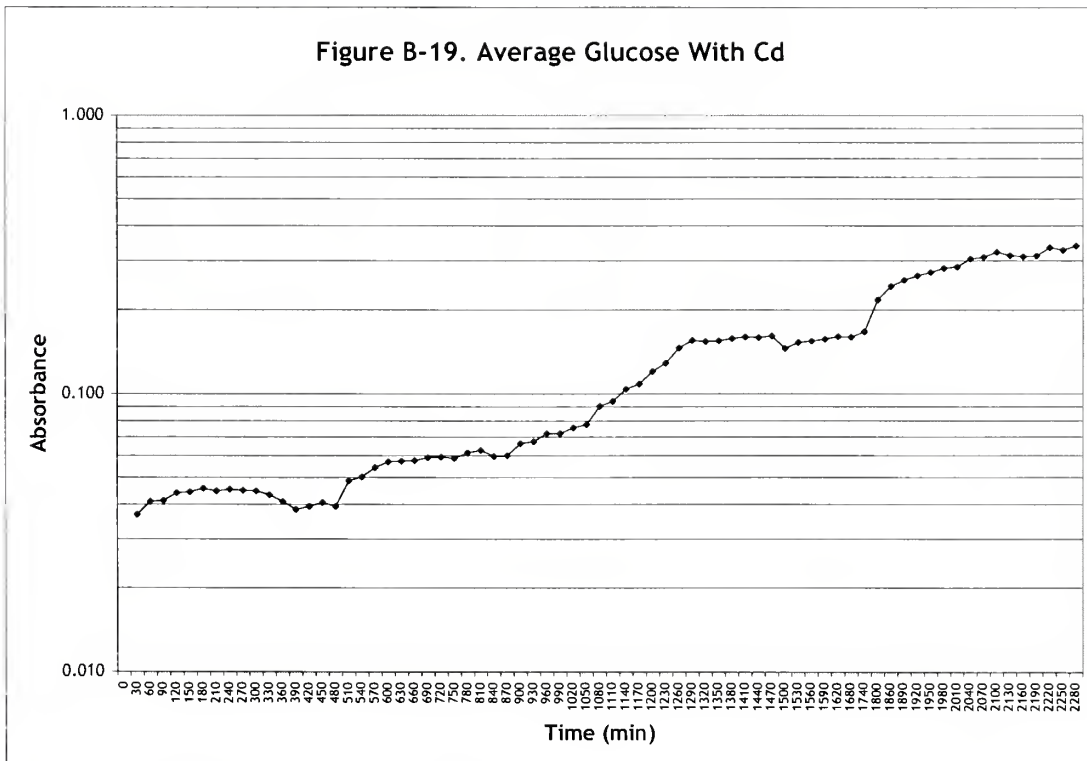
Absorbance increases from 0.065 to 0.131

Actual growth rate = (1260 min - 1020 min) = 240 min



Absorbance increases from 0.081 to 0.161

Actual growth rate = (1290 min - 1050 min) = 240 min



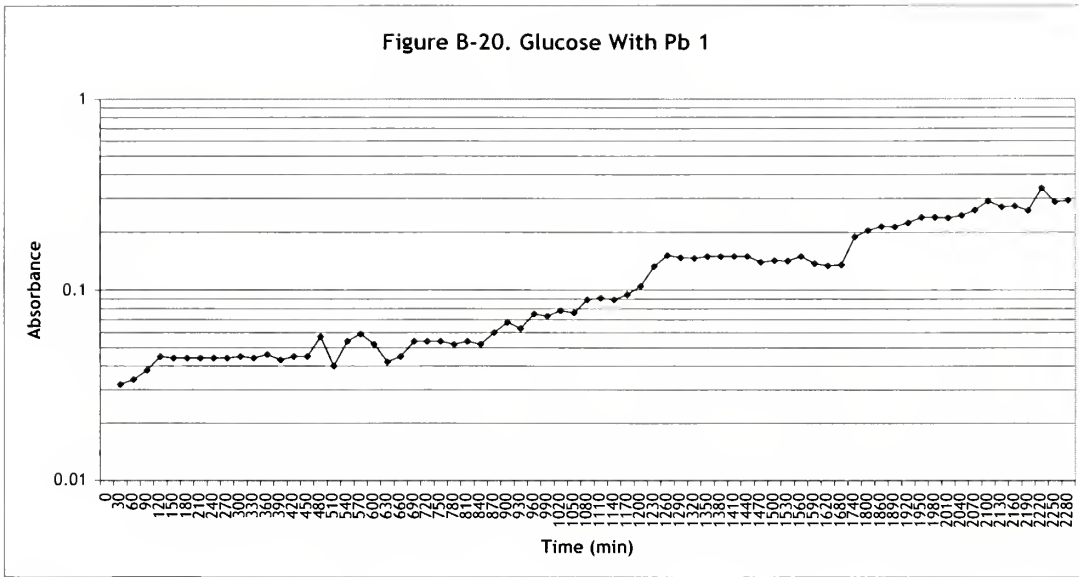
Absorbance increases from 0.078 to 0.156

Actual growth rate = (1290 min - 1050 min) = 240 min

Glucose and Pb at concentration of $1 \times 10^{-3} \text{M}$

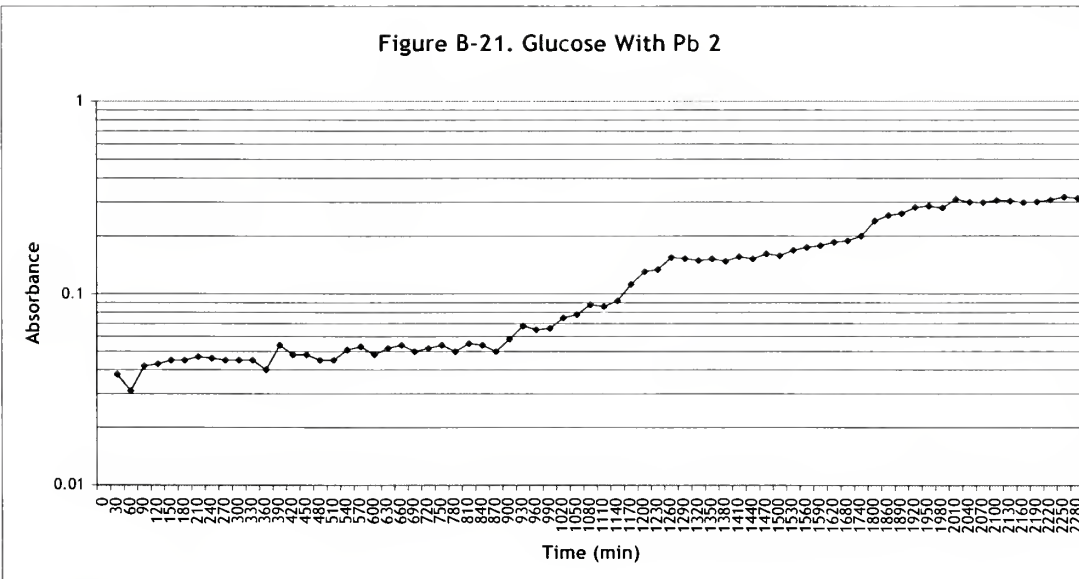
Time (min)	Run 1	Run 2	Run3	Average
0	0	0	0	0.000
30	0.032	0.038	0.032	0.034
60	0.034	0.031	0.036	0.034
90	0.038	0.042	0.041	0.040
120	0.045	0.043	0.043	0.044
150	0.044	0.045	0.045	0.045
180	0.044	0.045	0.044	0.044
210	0.044	0.047	0.046	0.046
240	0.044	0.046	0.043	0.044
270	0.044	0.045	0.044	0.044
300	0.045	0.045	0.045	0.045
330	0.044	0.045	0.044	0.044
360	0.046	0.040	0.046	0.044
390	0.043	0.054	0.043	0.047
420	0.045	0.048	0.047	0.047
450	0.045	0.048	0.051	0.048
480	0.057	0.045	0.057	0.053
510	0.040	0.045	0.053	0.046
540	0.054	0.051	0.056	0.054
570	0.059	0.053	0.059	0.057
600	0.052	0.048	0.054	0.051
630	0.042	0.052	0.056	0.050
660	0.045	0.054	0.057	0.052
690	0.054	0.050	0.054	0.053
720	0.054	0.052	0.056	0.054
750	0.054	0.054	0.057	0.055
780	0.052	0.050	0.059	0.054
810	0.054	0.055	0.058	0.056
840	0.052	0.054	0.057	0.054
870	0.060	0.050	0.060	0.057
900	0.068	0.058	0.068	0.065
930	0.063	0.068	0.067	0.066
960	0.075	0.065	0.075	0.072
990	0.073	0.066	0.078	0.072
1020	0.078	0.075	0.079	0.077
1050	0.076	0.078	0.081	0.078
1080	0.089	0.088	0.089	0.089
1110	0.091	0.086	0.091	0.089
1140	0.089	0.092	0.092	0.091
1170	0.095	0.112	0.095	0.101
1200	0.105	0.131	0.108	0.115
1230	0.133	0.134	0.141	0.136
1260	0.152	0.155	0.161	0.156
1290	0.148	0.153	0.162	0.154
1320	0.147	0.149	0.159	0.152
1350	0.150	0.152	0.163	0.155
1380	0.150	0.148	0.165	0.154
1410	0.150	0.156	0.161	0.156
1440	0.150	0.152	0.167	0.156
1470	0.140	0.162	0.165	0.156
1500	0.143	0.158	0.163	0.155
1530	0.142	0.169	0.171	0.161

1560	0.150	0.175	0.169	0.165
1590	0.138	0.178	0.173	0.163
1620	0.134	0.186	0.182	0.167
1680	0.136	0.189	0.186	0.170
1740	0.190	0.200	0.191	0.194
1800	0.205	0.240	0.211	0.219
1860	0.215	0.256	0.215	0.229
1890	0.214	0.262	0.217	0.231
1920	0.224	0.282	0.218	0.241
1950	0.240	0.287	0.242	0.256
1980	0.240	0.280	0.261	0.260
2010	0.238	0.311	0.293	0.281
2040	0.245	0.300	0.301	0.282
2070	0.261	0.299	0.308	0.289
2100	0.291	0.308	0.311	0.303
2130	0.271	0.305	0.312	0.296
2160	0.275	0.299	0.308	0.294
2190	0.260	0.302	0.311	0.291
2220	0.340	0.309	0.326	0.325
2250	0.288	0.320	0.336	0.315
2280	0.293	0.315	0.332	0.313



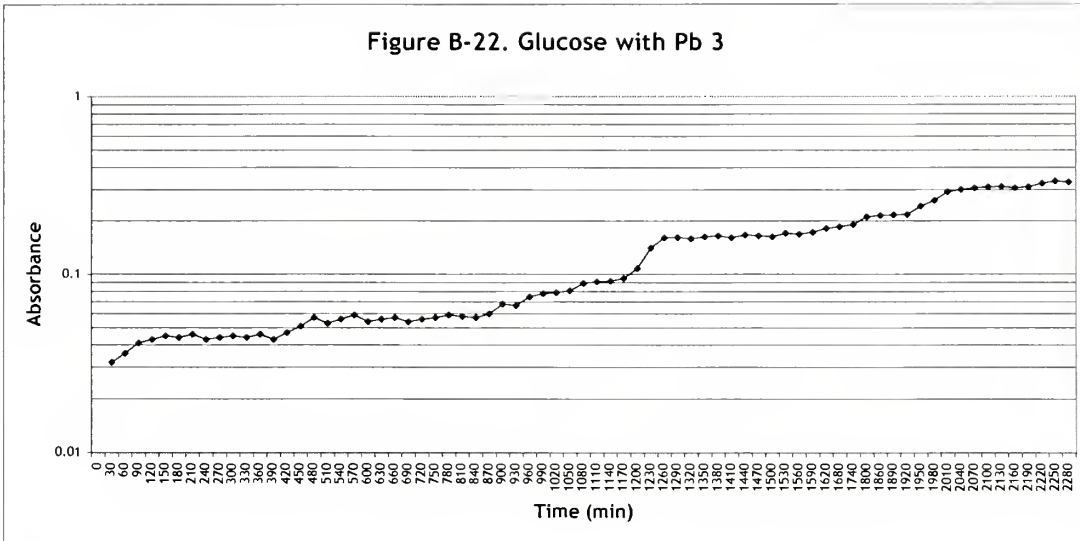
Absorbance increases from 0.076 to 0.152

Actual growth rate = (1260 min - 1050 min) = 210 min



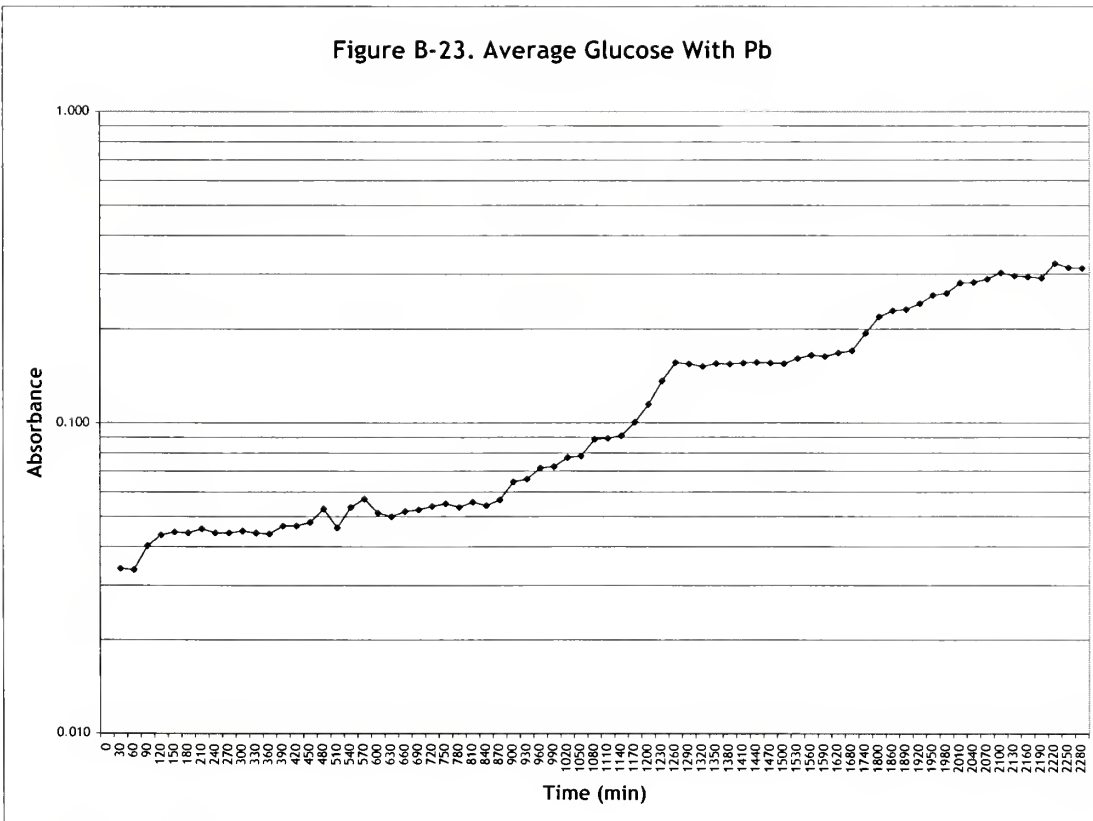
Absorbance increases from 0.076 to 0.152

Actual growth rate = (1260 min - 1050 min) = 210 min



Absorbance increases from 0.081 to 0.161

Actual growth rate = (1260 min - 1050 min) = 210 min



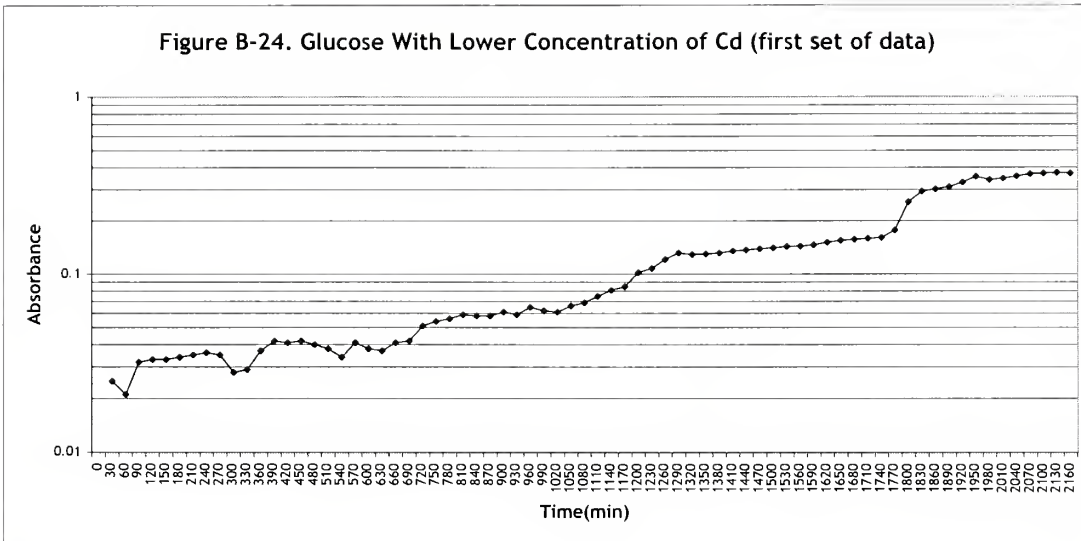
Absorbance increases from 0.078 to 0.156

Actual growth rate = (1260 min - 1050 min) = 210 min

Glucose and Cd at concentration of $1 \times 10^{-5} \text{M}$

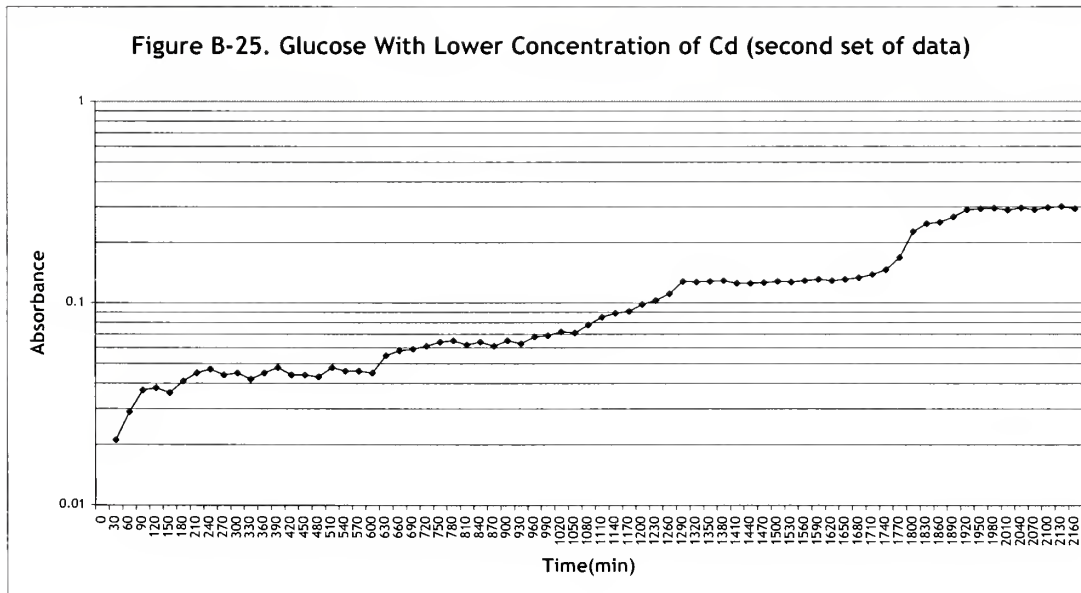
Time(min)	Run1	Run2	Run3	Average
0	0	0	0	0.000
30	0.025	0.021	0.024	0.023
60	0.021	0.029	0.023	0.024
90	0.032	0.037	0.028	0.032
120	0.033	0.038	0.033	0.035
150	0.033	0.036	0.032	0.034
180	0.034	0.041	0.034	0.036
210	0.035	0.045	0.036	0.039
240	0.036	0.047	0.038	0.040
270	0.035	0.044	0.041	0.040
300	0.028	0.045	0.046	0.040
330	0.029	0.042	0.048	0.040
360	0.037	0.045	0.044	0.042
390	0.042	0.048	0.045	0.045
420	0.041	0.044	0.046	0.044
450	0.042	0.044	0.048	0.045
480	0.040	0.043	0.044	0.042
510	0.038	0.048	0.042	0.043
540	0.034	0.046	0.046	0.042
570	0.041	0.046	0.048	0.045
600	0.038	0.045	0.046	0.043
630	0.037	0.055	0.051	0.048
660	0.041	0.058	0.054	0.051
690	0.042	0.059	0.051	0.051
720	0.051	0.061	0.052	0.055
750	0.054	0.064	0.054	0.057
780	0.056	0.065	0.056	0.059
810	0.059	0.062	0.058	0.060
840	0.058	0.064	0.059	0.060
870	0.058	0.061	0.058	0.059
900	0.061	0.065	0.062	0.063
930	0.059	0.063	0.063	0.062
960	0.065	0.068	0.065	0.066
990	0.062	0.069	0.064	0.065
1020	0.061	0.072	0.065	0.066
1050	0.066	0.071	0.066	0.068
1080	0.069	0.078	0.068	0.072
1110	0.075	0.085	0.078	0.079
1140	0.081	0.089	0.081	0.084
1170	0.085	0.091	0.092	0.089
1200	0.102	0.098	0.104	0.101
1230	0.108	0.103	0.114	0.108
1260	0.121	0.111	0.123	0.118
1290	0.132	0.128	0.128	0.129
1320	0.129	0.127	0.129	0.128
1350	0.130	0.128	0.130	0.129
1380	0.132	0.129	0.134	0.132
1410	0.135	0.125	0.138	0.133
1440	0.137	0.125	0.141	0.134
1470	0.139	0.126	0.144	0.136
1500	0.141	0.128	0.146	0.138

1530	0.143	0.127	0.148	0.139
1560	0.144	0.129	0.151	0.141
1590	0.146	0.131	0.154	0.144
1620	0.151	0.129	0.158	0.146
1650	0.155	0.131	0.156	0.147
1680	0.157	0.133	0.161	0.150
1710	0.159	0.138	0.163	0.153
1740	0.161	0.146	0.168	0.158
1770	0.177	0.168	0.186	0.177
1800	0.256	0.226	0.248	0.243
1830	0.294	0.248	0.297	0.280
1860	0.303	0.252	0.312	0.289
1890	0.311	0.268	0.325	0.301
1920	0.331	0.291	0.333	0.318
1950	0.356	0.294	0.361	0.337
1980	0.342	0.296	0.363	0.334
2010	0.347	0.290	0.368	0.335
2040	0.357	0.297	0.371	0.342
2070	0.368	0.291	0.374	0.344
2100	0.371	0.298	0.379	0.349
2130	0.375	0.302	0.381	0.353
2160	0.372	0.294	0.379	0.348



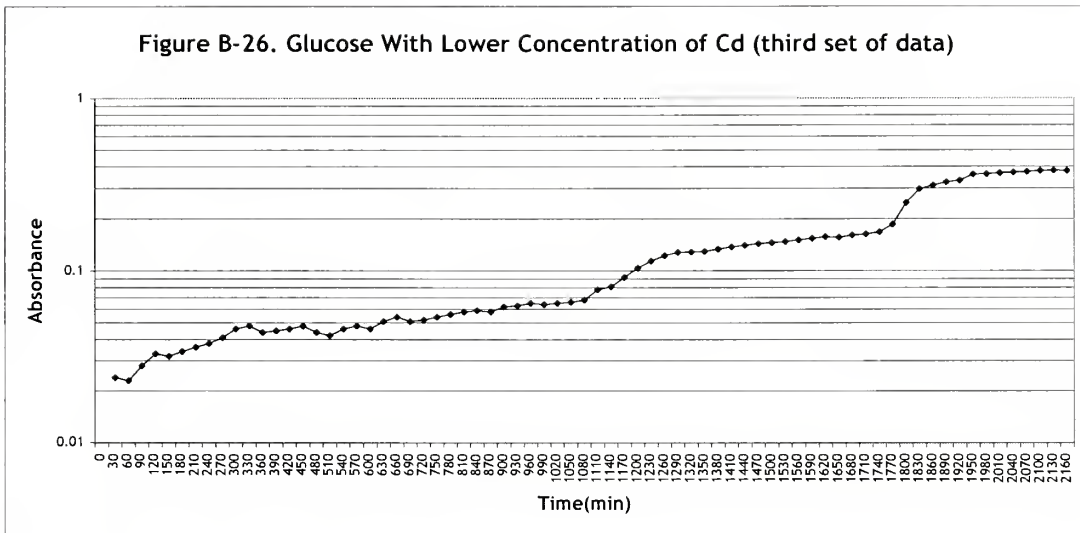
Absorbance increases from 0.177 to 0.356

Actual growth rate = (1950 min - 1770 min) = 180 min



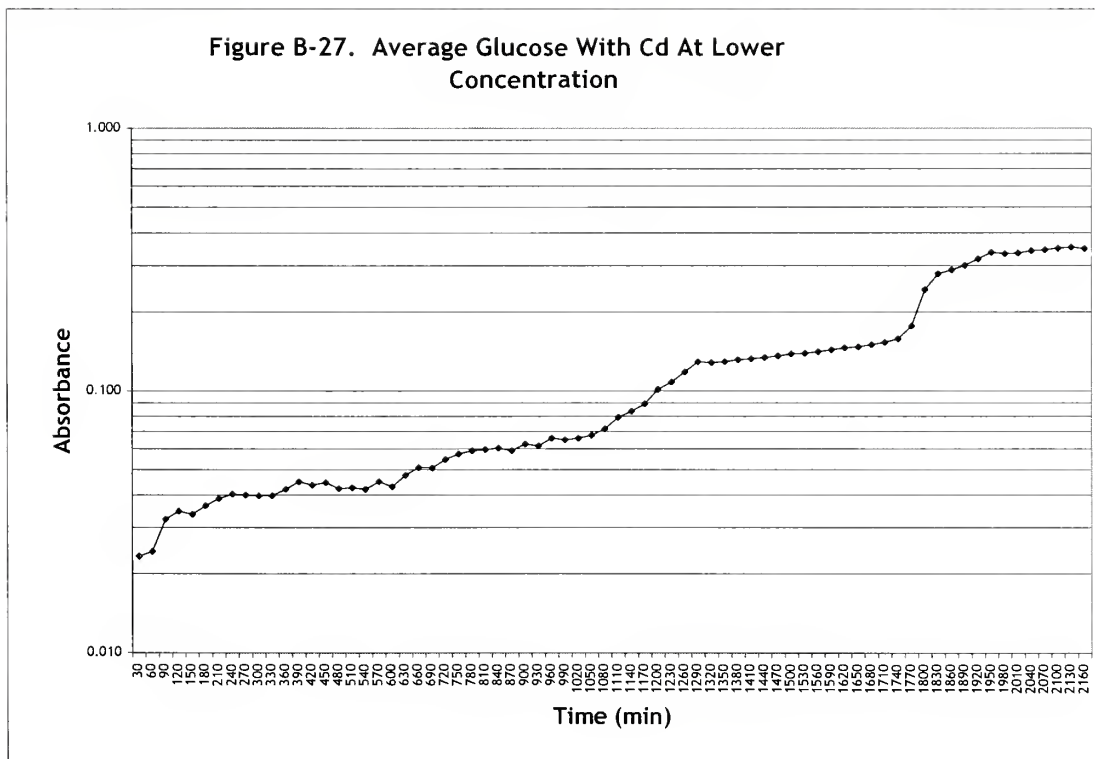
Absorbance increases from 0.146 to 0.291

Actual growth rate = (1920 min - 1740 min) = 180 min



Absorbance increases from 0.156 to 0.312

Actual growth rate = (1860 min - 1650 min) = 210 min



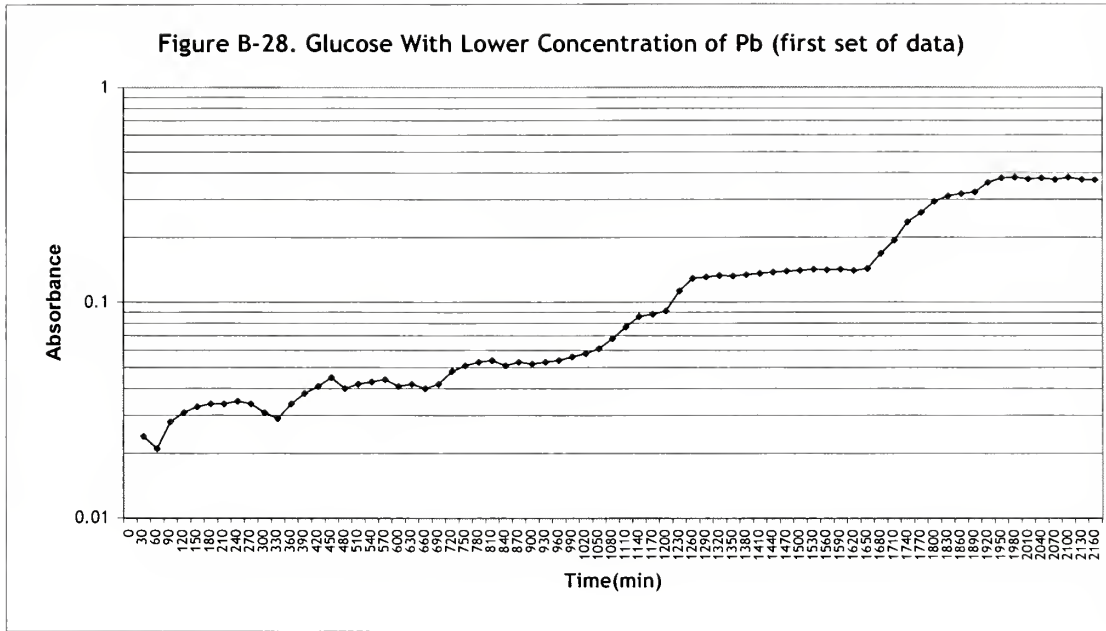
Absorbance increases from 0.158 to 0.318

Actual growth rate = (1920 min - 1740 min) = 180 min

Glucose and Pb at concentration of $1 \times 10^{-5} \text{M}$

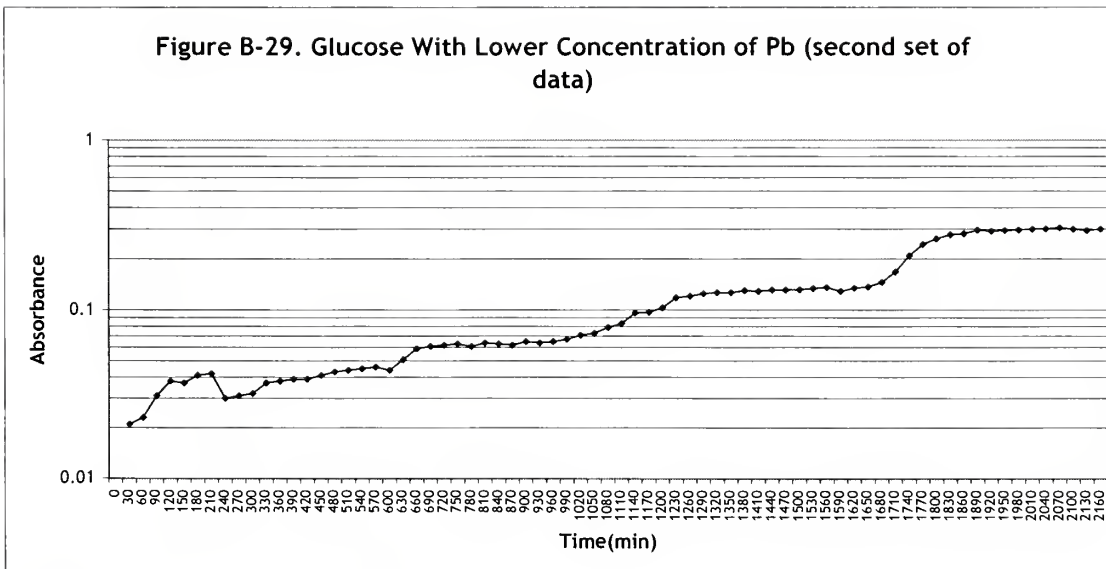
Time(min)	Run1	Run2	Run3	Average
0	0	0	0	0.000
30	0.024	0.021	0.025	0.023
60	0.021	0.023	0.021	0.022
90	0.028	0.031	0.032	0.030
120	0.031	0.038	0.033	0.034
150	0.033	0.037	0.034	0.035
180	0.034	0.041	0.033	0.036
210	0.034	0.042	0.035	0.037
240	0.035	0.030	0.036	0.034
270	0.034	0.031	0.035	0.033
300	0.031	0.032	0.032	0.032
330	0.029	0.037	0.036	0.034
360	0.034	0.038	0.037	0.036
390	0.038	0.039	0.042	0.040
420	0.041	0.039	0.041	0.040
450	0.045	0.041	0.042	0.043
480	0.040	0.043	0.041	0.041
510	0.042	0.044	0.043	0.043
540	0.043	0.045	0.046	0.045
570	0.044	0.046	0.045	0.045
600	0.041	0.044	0.047	0.044
630	0.042	0.051	0.05	0.048
660	0.040	0.059	0.052	0.050
690	0.042	0.061	0.058	0.054
720	0.048	0.062	0.061	0.057
750	0.051	0.063	0.062	0.059
780	0.053	0.061	0.064	0.059
810	0.054	0.064	0.065	0.061
840	0.051	0.063	0.067	0.060
870	0.053	0.062	0.065	0.060
900	0.052	0.065	0.061	0.059
930	0.053	0.064	0.064	0.060
960	0.054	0.065	0.063	0.061
990	0.056	0.067	0.065	0.063
1020	0.058	0.071	0.068	0.066
1050	0.061	0.073	0.071	0.068
1080	0.068	0.079	0.075	0.074
1110	0.077	0.083	0.078	0.079
1140	0.086	0.096	0.081	0.088
1170	0.088	0.097	0.089	0.091
1200	0.091	0.103	0.102	0.099
1230	0.113	0.118	0.114	0.115
1260	0.129	0.121	0.126	0.125
1290	0.131	0.125	0.132	0.129
1320	0.133	0.127	0.134	0.131
1350	0.132	0.127	0.135	0.131
1380	0.134	0.13	0.134	0.133
1410	0.136	0.129	0.137	0.134

1440	0.138	0.131	0.137	0.135
1470	0.139	0.131	0.139	0.136
1500	0.140	0.132	0.141	0.138
1530	0.142	0.134	0.143	0.140
1560	0.141	0.136	0.144	0.140
1590	0.142	0.129	0.146	0.139
1620	0.140	0.135	0.149	0.141
1650	0.143	0.137	0.146	0.142
1680	0.168	0.146	0.157	0.157
1710	0.194	0.168	0.172	0.178
1740	0.236	0.209	0.211	0.219
1770	0.261	0.244	0.251	0.252
1800	0.294	0.263	0.291	0.283
1830	0.312	0.278	0.301	0.297
1860	0.319	0.281	0.312	0.304
1890	0.325	0.295	0.323	0.314
1920	0.361	0.291	0.358	0.337
1950	0.378	0.294	0.372	0.348
1980	0.381	0.297	0.379	0.352
2010	0.375	0.299	0.381	0.352
2040	0.378	0.301	0.382	0.354
2070	0.372	0.305	0.379	0.352
2100	0.381	0.299	0.385	0.355
2130	0.372	0.294	0.381	0.349
2160	0.371	0.299	0.382	0.351



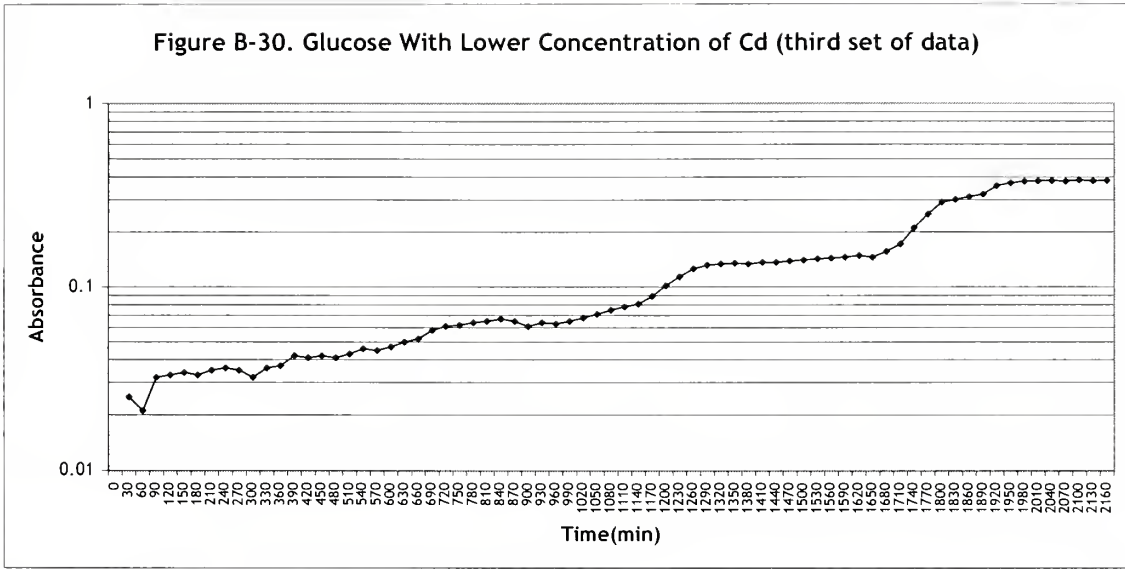
Absorbance increases from 0.143 to 0.294

Actual growth rate = (1800 min - 1650 min) = 150 min



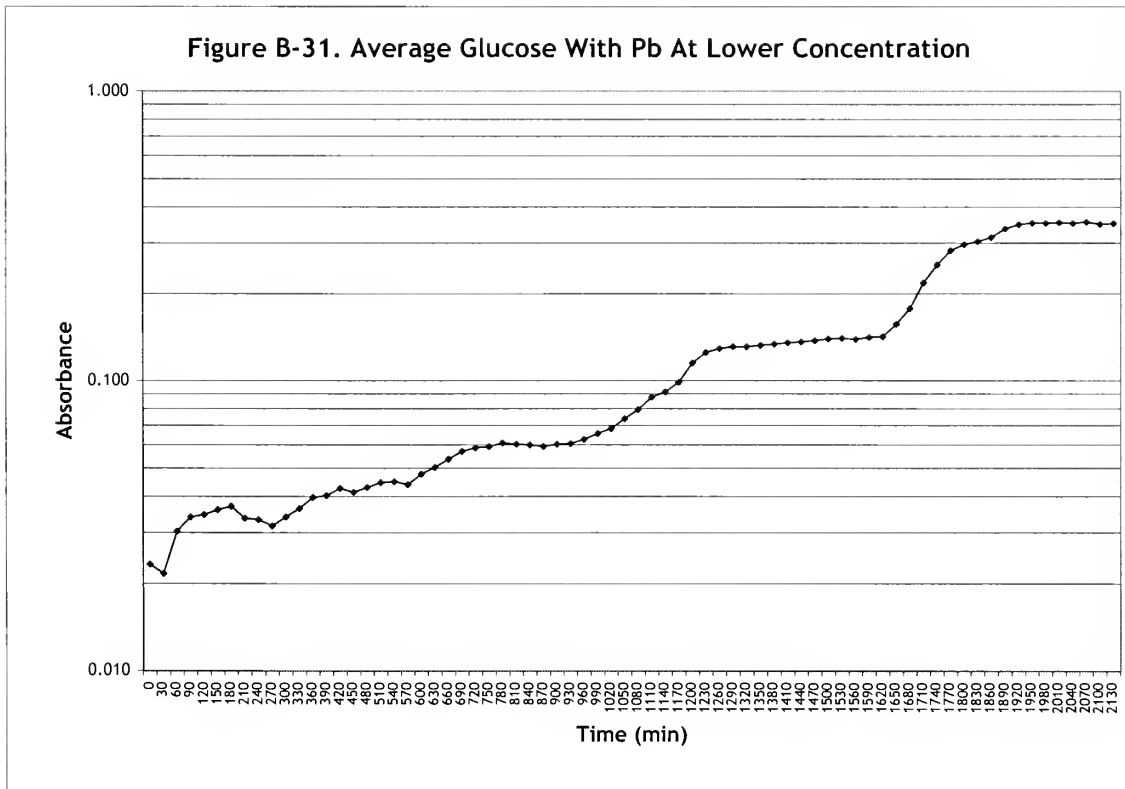
Absorbance increases from 0.146 to 0.278

Actual growth rate = (1830 min - 1680 min) = 150 min



Absorbance increases from 0.157 to 0.312

Actual growth rate = (1860 min - 1680 min) = 180 min

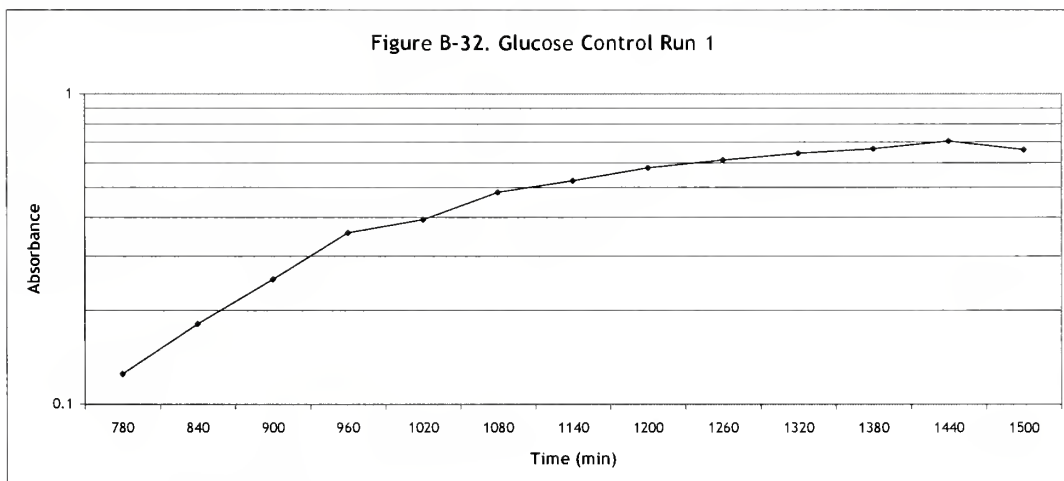


Absorbance increases from 0.142 to 0.283

Actual growth rate = (1800 min - 1650 min) = 150 min

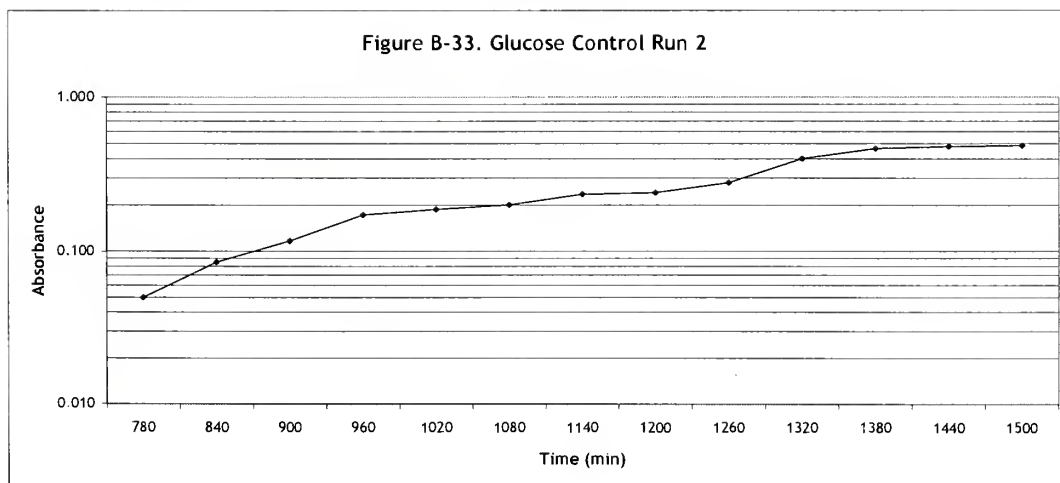
Data for *M.roseus* from absorbance vs. time in LB Broth- Rich Media

Glucose control				
Time(min)	Run1	Run2	Run3	Average
0				
780	0.125	0.050	0.091	0.089
840	0.181	0.085	0.132	0.133
900	0.252	0.117	0.175	0.181
960	0.357	0.173	0.266	0.265
1020	0.394	0.188	0.301	0.294
1080	0.482	0.201	0.358	0.347
1140	0.525	0.236	0.412	0.391
1200	0.578	0.242	0.475	0.432
1260	0.612	0.281	0.512	0.468
1320	0.644	0.401	0.556	0.534
1380	0.666	0.466	0.608	0.580
1440	0.706	0.479	0.644	0.610
1500	0.661	0.487	0.689	0.612



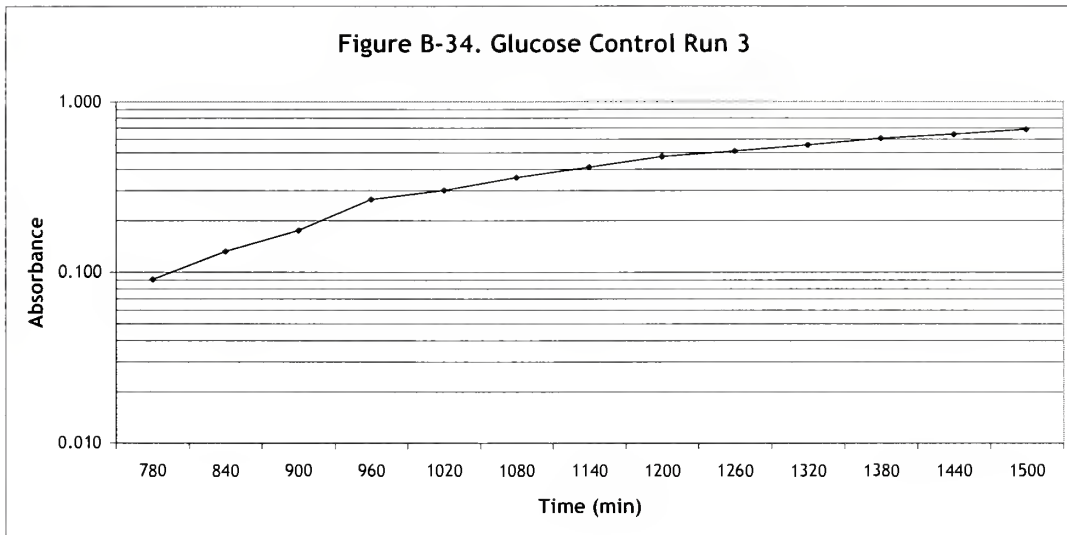
Absorbance increases from 0.125 to .0.252

Actual growth rate = (900min -780 min) = 120 min



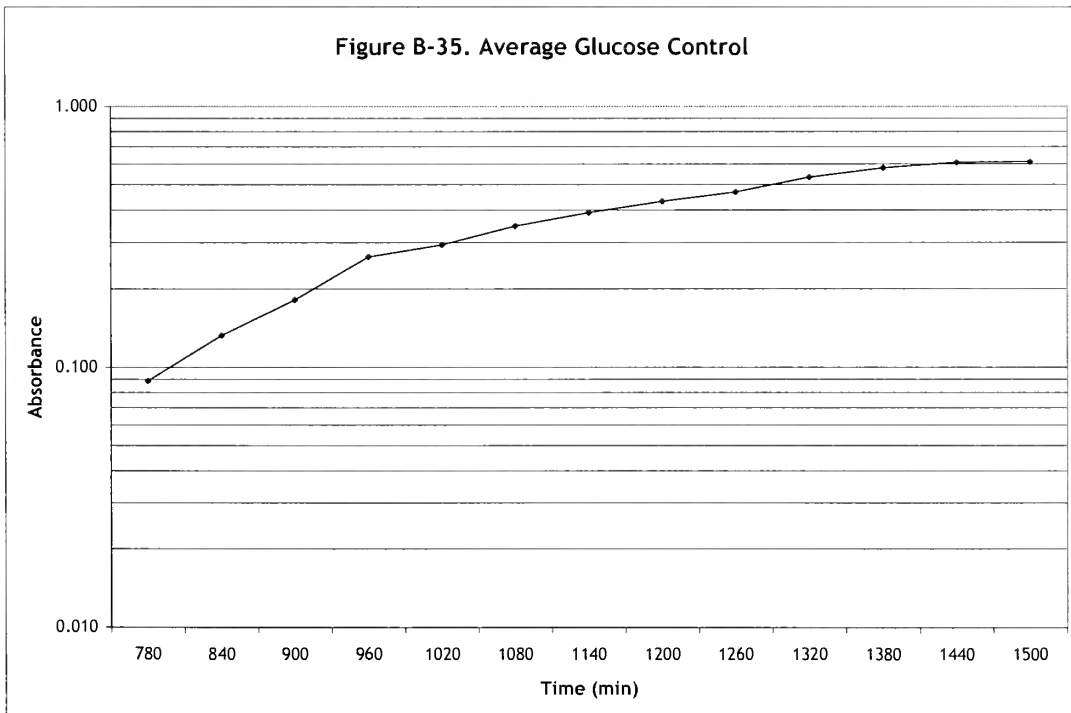
Absorbance increases from 0.085 to .0.173

Actual growth rate = (960min -840 min) = 120 min



Absorbance increases from 0.132 to .0.266

Actual growth rate = (960min -840 min) = 120 min

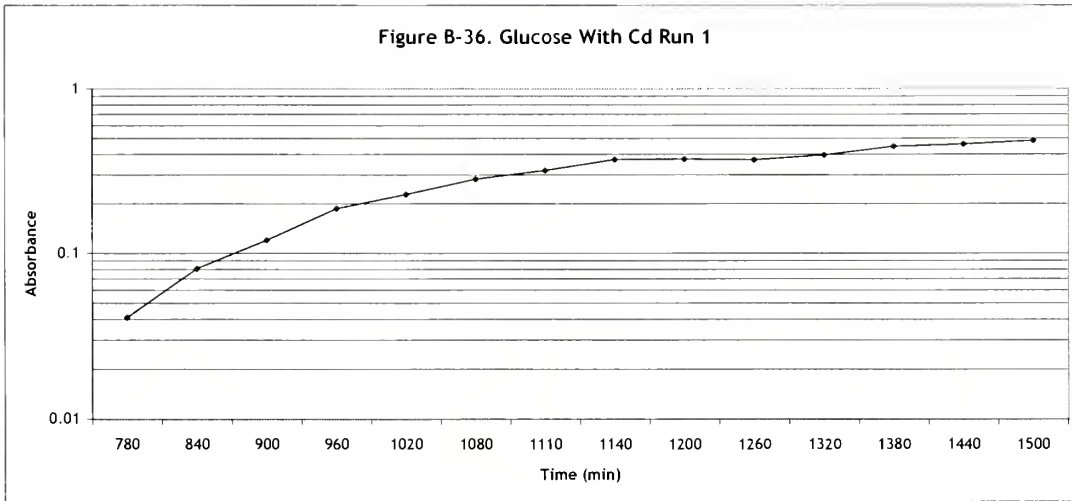


Absorbance increases from 0.133 to .0.265

Actual growth rate = (960min -840 min) = 120 min

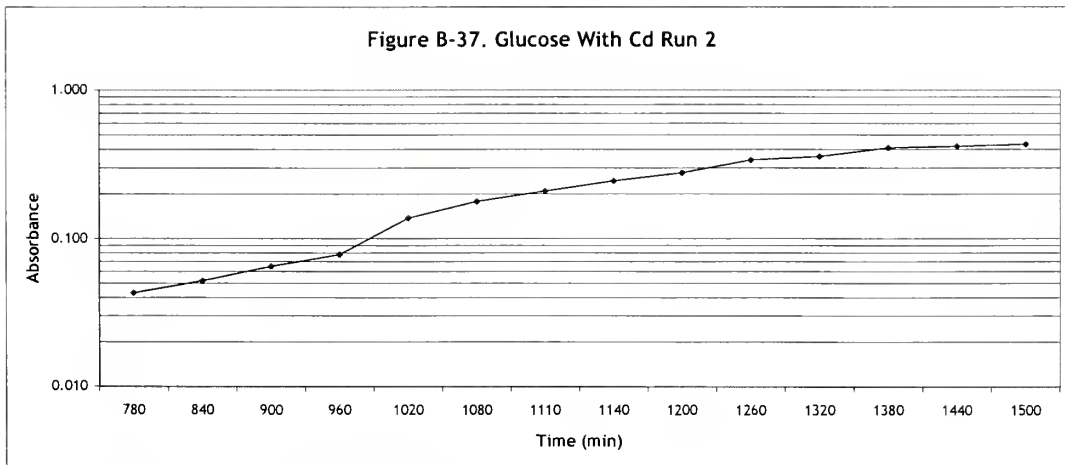
Data for *M. roseus* from absorbance vs. time in LB Broth- Rich Media

Glucose With 1×10^{-5} M Cd				
Time(min)	Run1	Run2	Run3	Average
0				
780	0.041	0.043	0.032	0.039
840	0.081	0.052	0.059	0.064
900	0.120	0.065	0.063	0.083
960	0.187	0.078	0.092	0.119
1020	0.228	0.138	0.107	0.158
1080	0.282	0.179	0.158	0.206
1110	0.319	0.209	0.228	0.252
1140	0.371	0.246	0.264	0.294
1200	0.374	0.278	0.298	0.317
1260	0.370	0.339	0.378	0.362
1320	0.396	0.358	0.454	0.403
1380	0.448	0.41	0.474	0.444
1440	0.462	0.421	0.492	0.458
1500	0.484	0.434	0.511	0.476



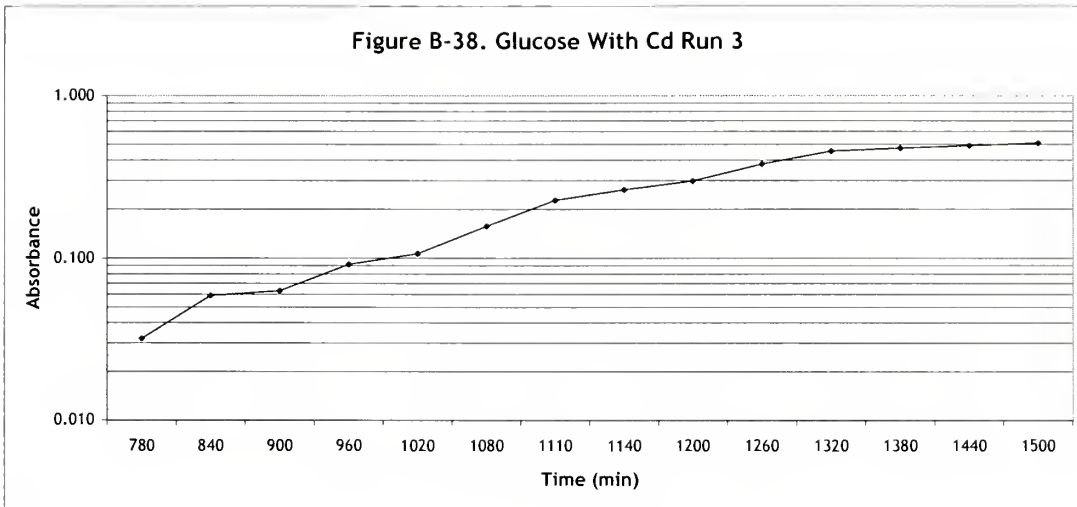
Absorbance increases from 0.187 to .0.371

Actual growth rate = (1140 min -960 min) = 180 min



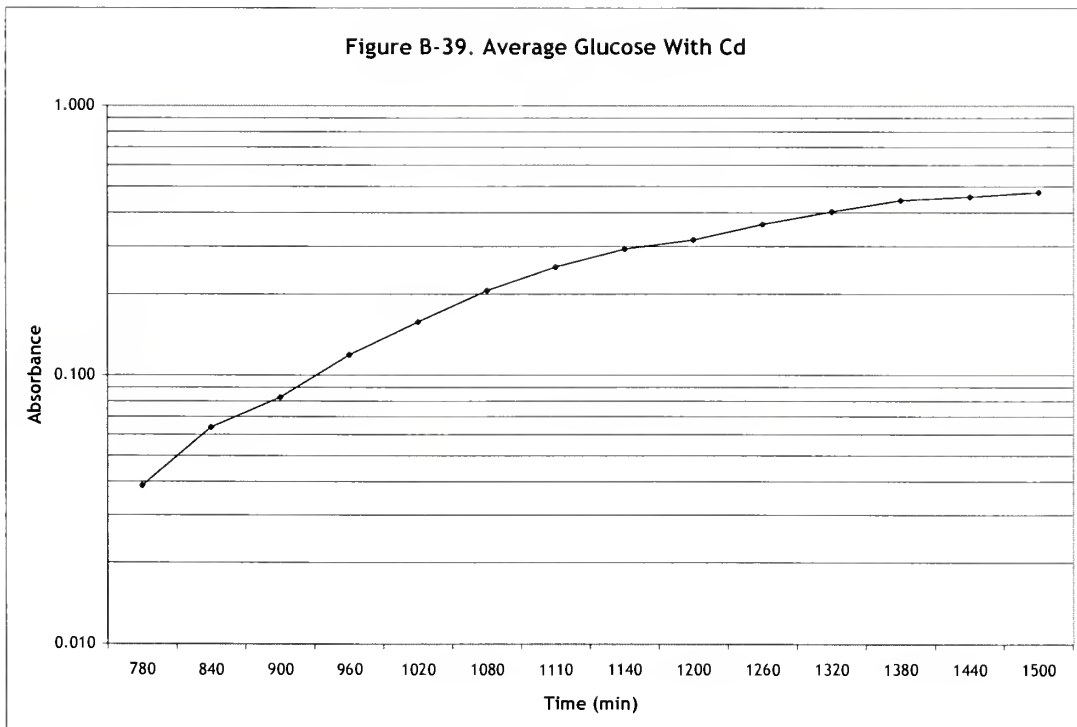
Absorbance increases from 0.138 to .0.278

Actual growth rate = (1200 min -1020 min) = 180 min



Absorbance increases from 0.228 to .0.454

Actual growth rate = (1320 min -1110 min) = 210 min

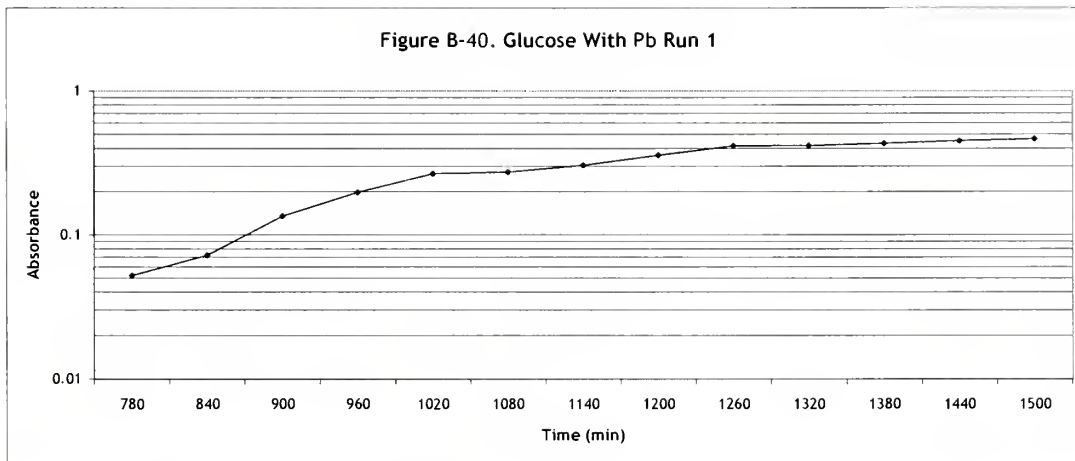


Absorbance increases from 0.158 to .0.317

Actual growth rate = (1200 min -1020 min) = 180 min

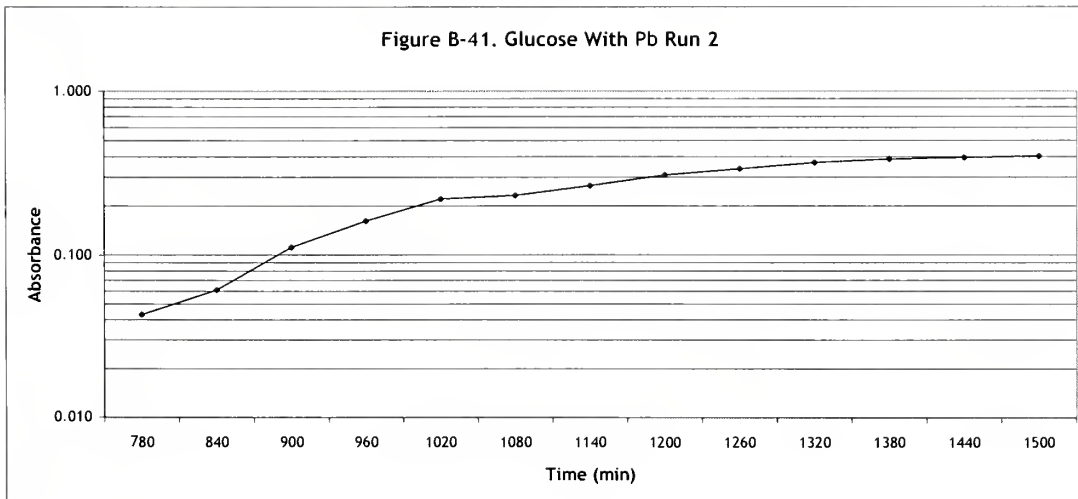
Data for *M.roseus* from absorbance vs. time in LB Broth- Rich Media

Glucose With 1×10^{-5} M Pb				
Time(min)	Run1	Run2	Run3	Average
0				
780	0.052	0.043	0.054	0.050
840	0.072	0.061	0.078	0.070
900	0.135	0.112	0.127	0.125
960	0.198	0.162	0.174	0.178
1020	0.267	0.221	0.235	0.241
1080	0.274	0.232	0.254	0.253
1140	0.305	0.266	0.298	0.290
1200	0.359	0.309	0.342	0.337
1260	0.416	0.337	0.384	0.379
1320	0.418	0.368	0.401	0.396
1380	0.434	0.386	0.422	0.414
1440	0.452	0.395	0.434	0.427
1500	0.466	0.402	0.451	0.440



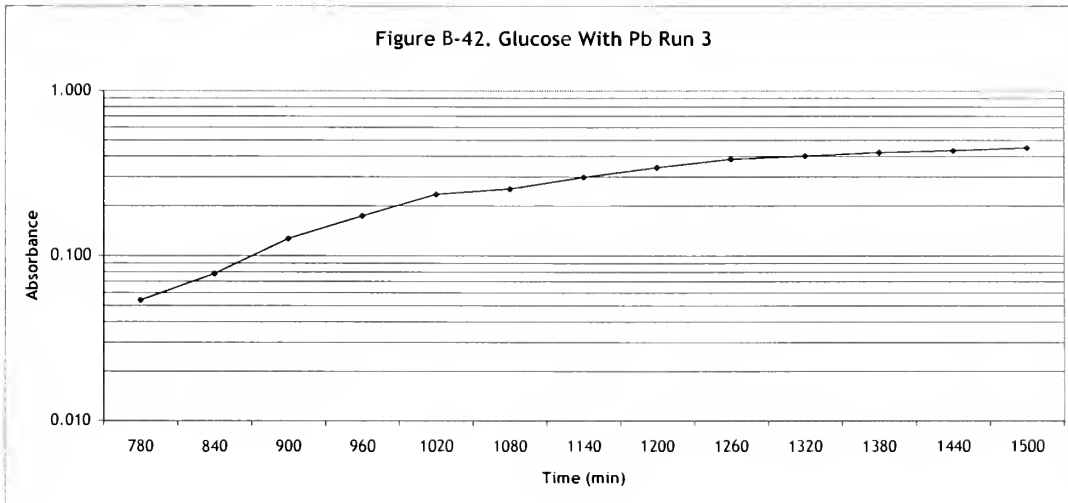
Absorbance increases from 0.135 to .267

Actual growth rate = (1020 min - 900 min) = 120 min



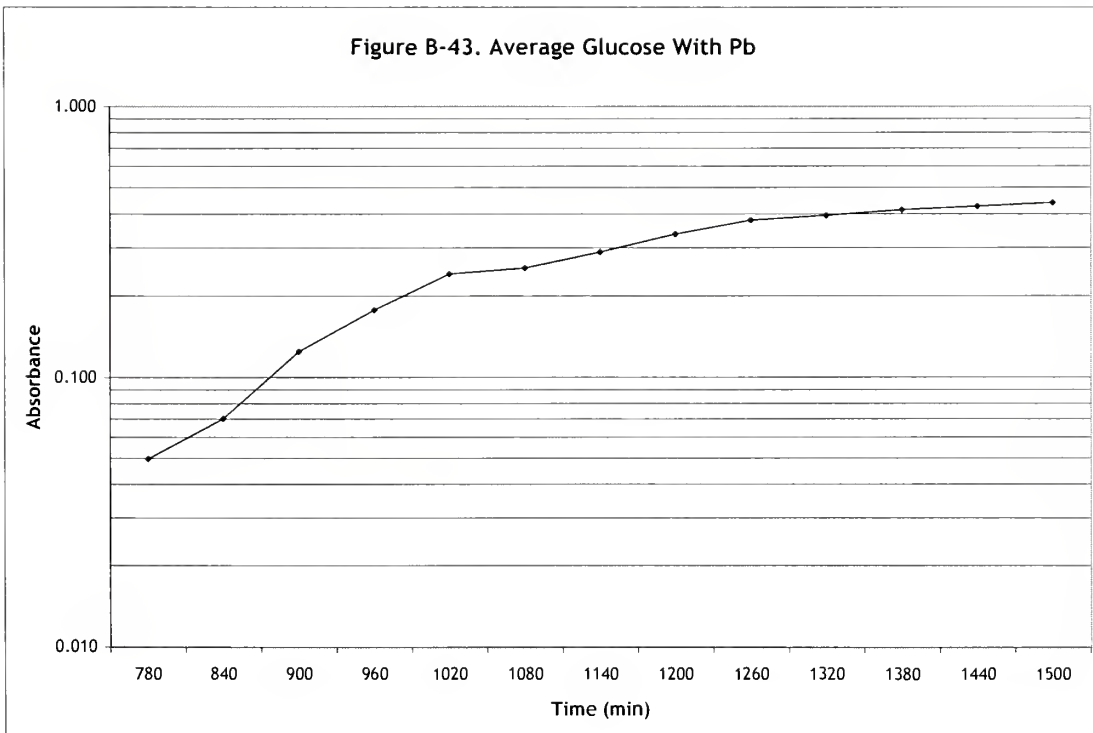
Absorbance increases from 0.112 to .221

Actual growth rate = (1020 min - 900 min) = 120 min



Absorbance increases from 0.127 to .254

Actual growth rate = (1080 min - 900 min) = 180 min

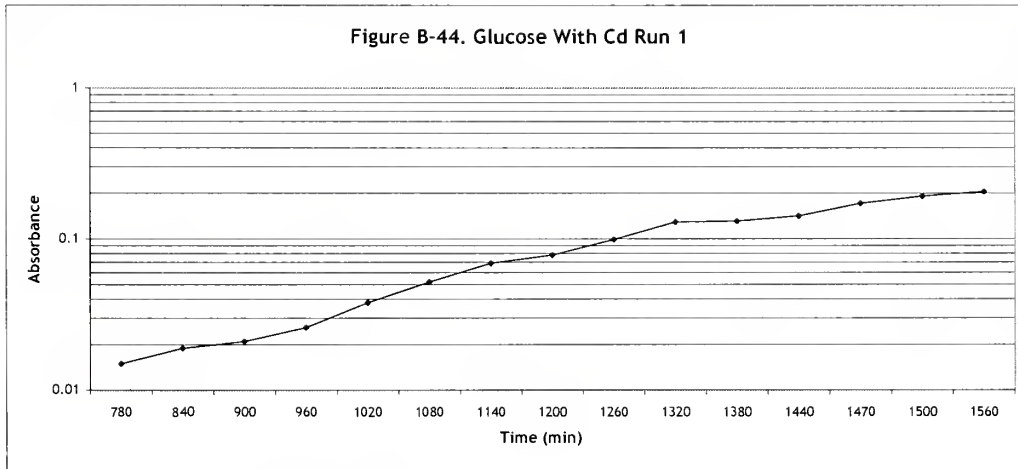


Absorbance increases from 0.125 to .253

Actual growth rate = (1080 min - 900 min) = 180 min

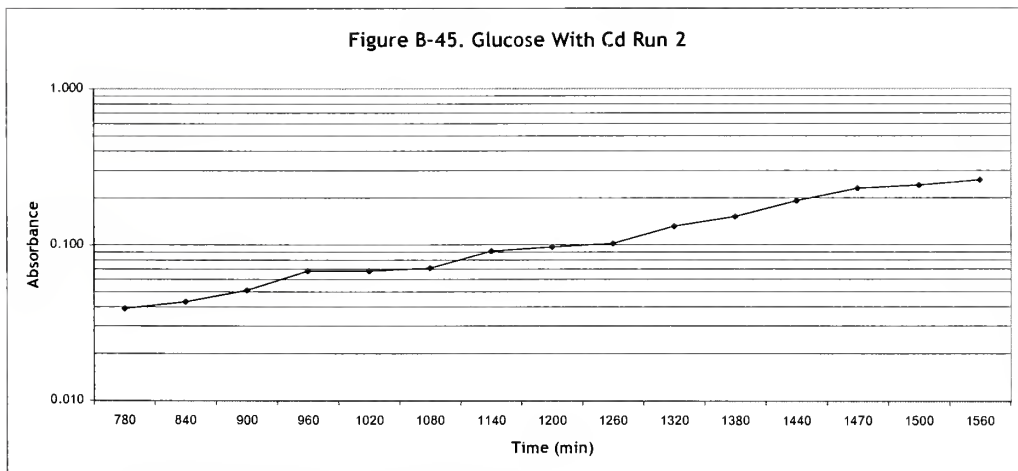
Data for *M.roseus* from absorbance vs. time in LB Broth- Rich Media

Glucose With $1 \cdot 10^{-3}$ M Cd				
Time(min)	Run1	Run2	Run3	Average
0				
780	0.015	0.039	0.021	0.025
840	0.019	0.043	0.028	0.030
900	0.021	0.051	0.033	0.035
960	0.026	0.068	0.034	0.043
1020	0.038	0.068	0.047	0.051
1080	0.052	0.071	0.054	0.059
1140	0.069	0.091	0.071	0.077
1200	0.078	0.097	0.082	0.086
1260	0.099	0.102	0.102	0.101
1320	0.129	0.132	0.123	0.128
1380	0.131	0.152	0.133	0.139
1440	0.142	0.192	0.138	0.157
1470	0.172	0.231	0.161	0.188
1500	0.191	0.242	0.177	0.203
1560	0.205	0.261	0.203	0.223



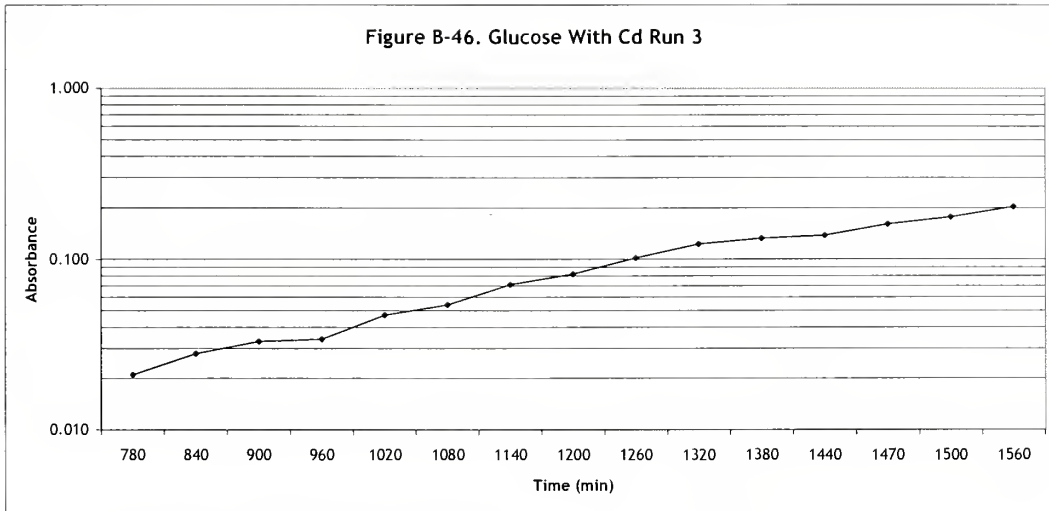
Absorbance increases from 0.099 to .0.198

Actual growth rate = (1500 min - 1260min) = 240 min



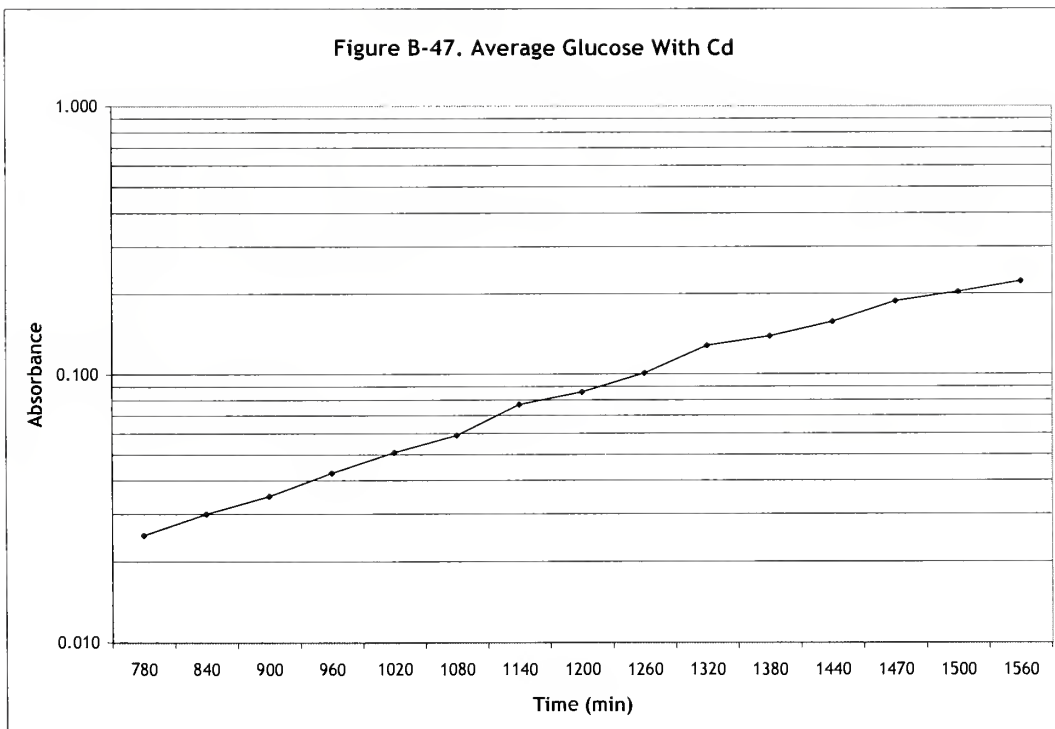
Absorbance increases from 0.132 to .0.261

Actual growth rate = (1560 min - 1320min) = 240 min



Absorbance increases from 0.102 to .0.203

Actual growth rate = (1560 min - 1260min) = 300 min

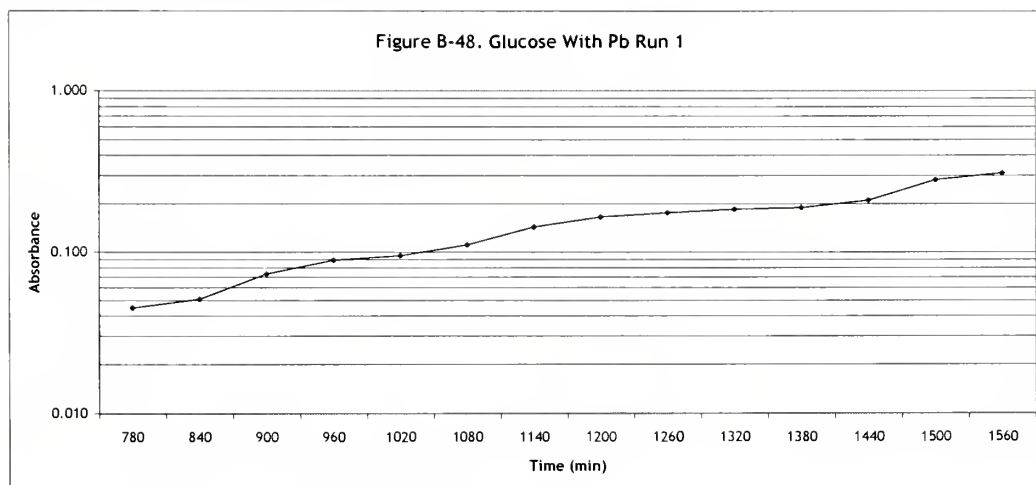


Absorbance increases from 0.101 to .0.203

Actual growth rate = (1500 min - 1260min) = 240 min

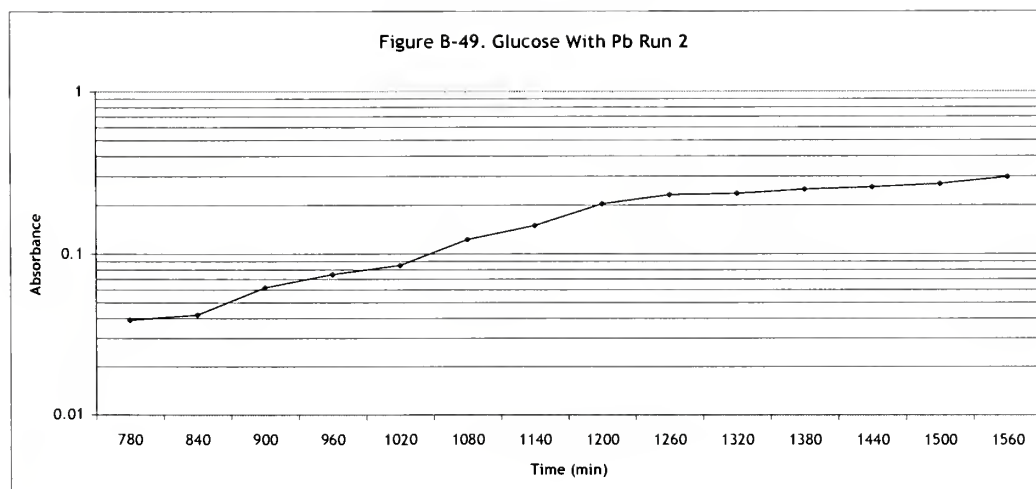
Data for *M. roseus* from absorbance vs. time in LB Broth- Rich Media

Glucose With 1×10^{-3} M Pb				
Time(min)	Run1	Run2	Run3	Average
0				
780	0.045	0.039	0.041	0.042
840	0.051	0.042	0.062	0.052
900	0.073	0.062	0.081	0.072
960	0.089	0.075	0.086	0.083
1020	0.095	0.085	0.089	0.090
1080	0.111	0.123	0.118	0.117
1140	0.144	0.150	0.138	0.144
1200	0.166	0.204	0.164	0.178
1260	0.176	0.231	0.181	0.196
1320	0.185	0.236	0.188	0.203
1380	0.189	0.25	0.192	0.210
1440	0.211	0.259	0.209	0.226
1500	0.282	0.271	0.273	0.275
1560	0.311	0.298	0.302	0.304



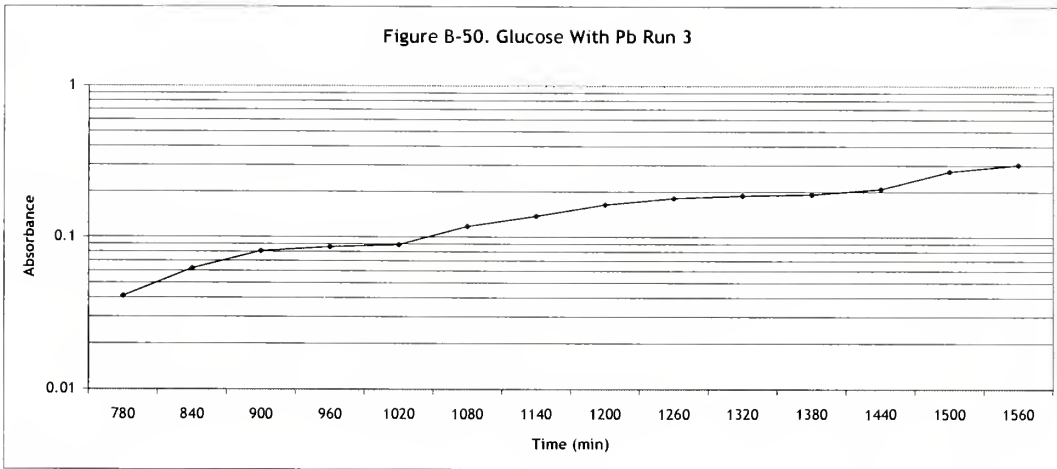
Absorbance increases from 0.073 to 0.144

Actual growth rate = (1140 min - 900 min) = 240 min



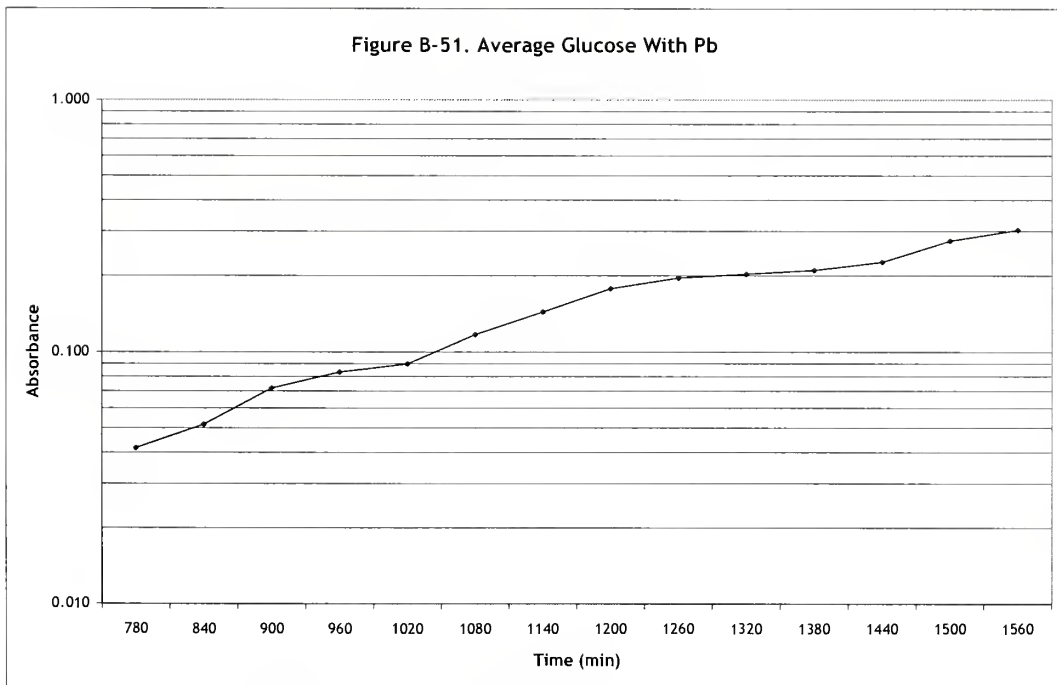
Absorbance increases from 0.075 to 0.150

Actual growth rate = (1140 min - 960min) = 180 min



Absorbance increases from 0.089 to .0.181

Actual growth rate = (1260 min - 1020min) = 240 min



Absorbance increases from 0.090 to 0.178

Actual growth rate = (1200 min - 1020min) = 180 min

M. roseus growth data
Table B-1. M.roseus cell growth over period of time

growth time: 7 hours

	Dilution
	1:5
(twice for all media)	5
	Average
lactose	260
lactose and Cd	110
lactose and Pb	150
glucose	253
glucose and Cd	88
glucose and Pb	232
	1160

growth time: 21 hours

	Dilution
	1:30
	30
	100+
lactose	154
lactose and Cd	148
lactose and Pb	15
glucose	203
glucose and Cd	30
glucose and Pb	90
	1:300
	300
	100+
lactose	54
lactose and Cd	6
lactose and Pb	6
glucose	7
glucose and Cd	1
glucose and Pb	2
	1:9,000
	900
	54
lactose	4
lactose and Cd	0
lactose and Pb	0
glucose	0
glucose and Cd	0
glucose and Pb	0
	Average
	44400
	4940
	4780
	6230
	1000
	2200

growth time: 18 hours

	Dilution
	1:5
	5
	100+
lactose	80
lactose and Cd	127
lactose and Pb	12
glucose	57
glucose and Cd	22
glucose and Pb	54
	1:25
	25
	100+
lactose	126
lactose and Cd	8
lactose and Pb	61
glucose	29
glucose and Cd	0
glucose and Pb	26
	1:250
	250
	126
lactose	31500
lactose and Cd	2017
lactose and Pb	3075
glucose	6
glucose and Cd	0
glucose and Pb	5
	Average
	1374
	570
	1300

growth time: 23 hours

	Dilution	1:30	1:300	1:3,000	1:30,000	Average
		30	300	3,000	30,000	
		(twice la+Cd,lac-Pb,glu,Pb)				
lactose	100+	143	16	2	50300	
lactose and Cd	100+	26	2	0	7200	
lactose and Pb	100+	39	3	0	10800	
glucose	100+	53	5	0	15600	
glucose and Cd	206	19	2	0	5960	
glucose and Pb	100+	65	6	0	19000	

growth time: 22 hours

	Dilution	1:1,000	1:10,000	1:100,000	Average
	100	1,000	10,000	100,000	
		(twice la+Cd,glu+Pb)			
lactose	100+	47	5	49000	
lactose and Cd	68	5	0	6200	
lactose and Pb	95	9	1	9500	
glucose	138	14	1	12600	
glucose and Cd	20	2	0	2000	
glucose and Pb	58	4	0	5200	

growth time: 26 hours

	Dilution	1:30	1:300	1:3,000	1:30,000	1:25,000	Average
		30	300	3,000	30,000	25,000	
		(twice la.,la-Pb,glu+Pb)					
lactose	100+	192	18	0	58067		
lactose and Cd	266	32	2	0	7860		
lactose and Pb	100+	39	4	0	11800		
glucose	100+	102	10	1	30200		
glucose and Cd	249	23	2	0	6790		
glucose and Pb	100+	83	7	0	23600		

growth time: 25 hours

	Dilution	1:300	1:3,000	1:30,000	Average
	30	300	3,000	30,000	
		(twice glu, glu+Pb)			
lactose	100+	184	14	2	52400
lactose and Cd	242	28	0	7220	
lactose and Pb	340	41	4	11500	
glucose	100+	73	6	0	20600
glucose and Cd	231	20	0	6310	
glucose and Pb	100+	75	7	0	21500

		growth time: 28 hours			
Dilution		1:30	1:300	1:3,000	1:30,000
	(twice la., la-Cd, glu+Pb)	30	300	3,000	30,000
	twice(la+Pb)	30	300	3,000	30,000
	1:3,000 1:30,000 1:25,000				
	Average				
	lactose	100+	196	21	0
	lactose and Cd	100+	28	3	0
	lactose and Pb	100+	58	6	0
	glucose	100+	263	28	3
	glucose and Cd	263	24	2	0
	glucose and Pb	100+	90	8	0
					26000

		growth time: 27 hours			
Dilution		1:30	1:300	1:3,000	1:30,000
	(twice la., la+Pb, glu+Pb)	30	300	3,000	30,000
	twice(la., la-Cd, glu+Cd)	30	300	3,000	30,000
	1:3,000 1:30,000 1:25,000				
	Average				
	lactose	100+	192	20	0
	lactose and Cd	280	26	3	0
	lactose and Pb	100+	45	4	0
	glucose	100+	147	14	2
	glucose and Cd	261	24	2	0
	glucose and Pb	100+	83	8	0
					24600

		growth time: 31 hours			
Dilution		1:30	1:300	1:3,000	1:30,000
	(twice la., la-Cd, la+Pb)	30	300	3,000	30,000
	twice (glu +Pb, lac-Pb)	30	300	3,000	30,000
	1:3,000 1:30,000 1:25,000 1:25,000				
	Average				
	lactose	100+	100+	32	3
	lactose and Cd	100+	171	10	0
	lactose and Pb	100+	117	12	1
	glucose	100+	294	36	3
	glucose and Cd	262	29	3	0
	glucose and Pb	100+	98	9	0
					27800

		growth time: 30 hours			
Dilution		1:30	1:300	1:3,000	1:30,000
	(twice glu+Pb)	30	300	3,000	30,000
	twice(la., la-Cd, glu+Cd)	30	300	3,000	30,000
	1:3,000 1:30,000 1:25,000 1:25,000				
	Average				
	lactose	100+	204	23	0
	lactose and Cd	100+	48	4	0
	lactose and Pb	100+	72	8	1
	glucose	100+	285	37	3
	glucose and Cd	240	30	3	0
	glucose and Pb	100+	93	9	0
					27600

32 hours		33 hours	
Dilution	1:300	1:300	1:300
	twice(la+Cd, glu+Cd)	twice(glu-Cd)	twice(glu-Cd)
	3,000	3,000	3,000
	1:3,000	1:3,000	1:3,000
	twice(la, la+Pb, glu)		
	30,000		
	Average		
	6		
	183000		
	lactose		
	100+		
	63		
	lactose and Cd		
	100+		
	144		
	40400		
	lactose and Pb		
	100+		
	39		
	99000		
	glucose		
	100+		
	40		
	140000		
	glucose and Cd		
	100+		
	26		
	86000		
	glucose and Pb		
	100+		
	106		
	306000		

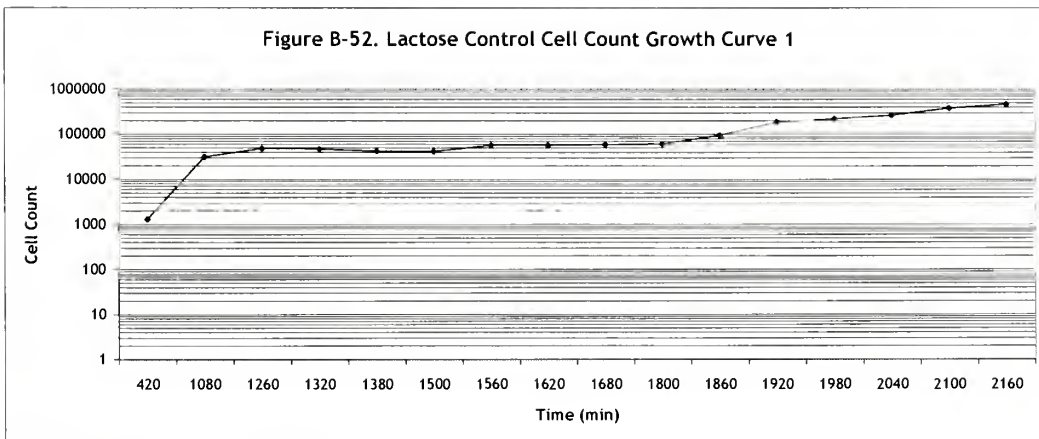
32 hours		33 hours	
Dilution	1:300	1:300	1:300
	twice(la+Cd, glu+Cd)	twice(glu-Cd)	twice(glu-Cd)
	3,000	3,000	3,000
	1:3,000	1:3,000	1:3,000
	twice(la, la+Pb, glu)		
	30,000		
	Average		
	6		
	183000		
	lactose		
	100+		
	63		
	lactose and Cd		
	100+		
	144		
	40400		
	lactose and Pb		
	100+		
	39		
	99000		
	glucose		
	100+		
	40		
	140000		
	glucose and Cd		
	100+		
	26		
	86000		
	glucose and Pb		
	100+		
	106		
	306000		

34 hours		35 hours	
Dilution	1:300	1:300	1:300
	twice(la+Cd, glu+Cd)	twice(glu-Cd)	twice(glu-Cd)
	3,000	3,000	3,000
	1:3,000	1:3,000	1:3,000
	twice(la, la+Pb, glu)		
	30,000		
	Average		
	11		
	273000		
	lactose		
	100+		
	89		
	lactose and Cd		
	100+		
	177		
	52775		
	lactose and Pb		
	100+		
	78		
	216000		
	glucose		
	100+		
	74		
	220500		
	glucose and Cd		
	100+		
	33		
	113000		
	glucose and Pb		
	100+		
	127		
	38525		

36 hours		37 hours	
Dilution	1:300	1:300	1:300
	twice(la+Cd, glu+Cd)	twice(glu-Cd)	twice(glu-Cd)
	3,000	3,000	3,000
	1:3,000	1:3,000	1:3,000
	twice(la, la+Pb, glu)		
	30,000		
	Average		
	16		
	416500		
	lactose		
	100+		
	157		
	lactose and Cd		
	100+		
	176		
	53450		
	lactose and Pb		
	100+		
	86		
	229500		
	glucose		
	100+		
	133		
	306000		
	glucose and Cd		
	100+		
	41		
	12200		
	glucose and Pb		
	100+		
	165		
	44375		

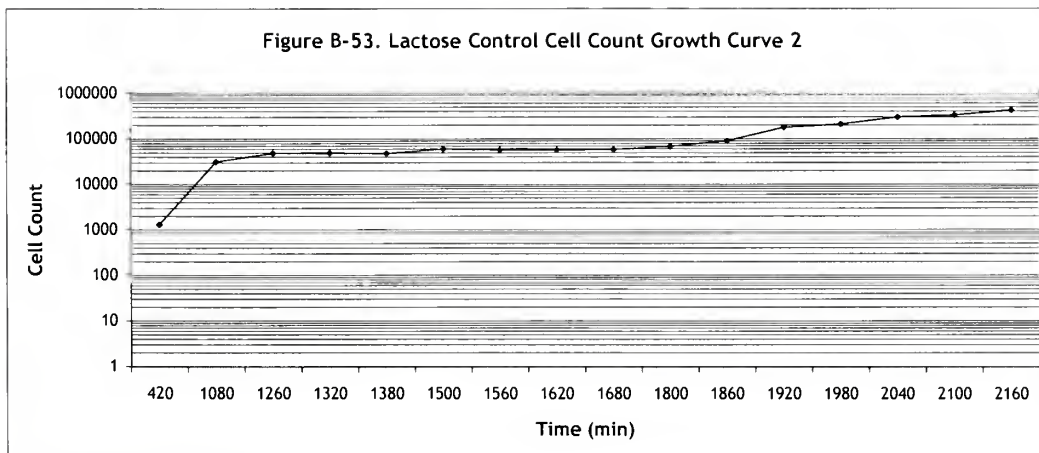
Cell Growth vs. Time
LACTOSE ONLY CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Cell Count 4	Average Cell Count
420	1300	1300	1300		1300
1080	31500	31500	31500		31500
1260	48600	48600	36000		44400
1320	47000	50000	50000		49000
1380	42900	48000	60000		50300
1500	42000	60000	55200		52400
1560	57600	57600	59000		58067
1620	57600	57600	60000		58400
1680	58800	58800	63000		60200
1800	61200	69000	69000		66400
1860	96000	90000	125000	125000	109000
1920	189000	180000	180000		183000
1980	222000	210000	250000	200000	220500
2040	267000	300000	275000	250000	273000
2100	384000	330000	325000	375000	353500
2160	471000	420000	400000	375000	416500



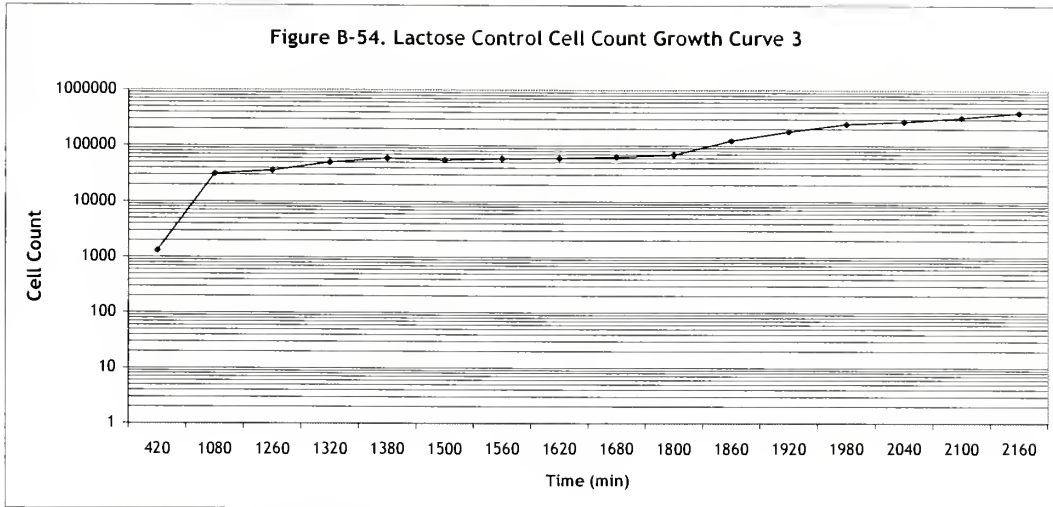
Cell Count increases from 96000 to 189000

Actual growth rate = (1920min - 1860min) = 60 min



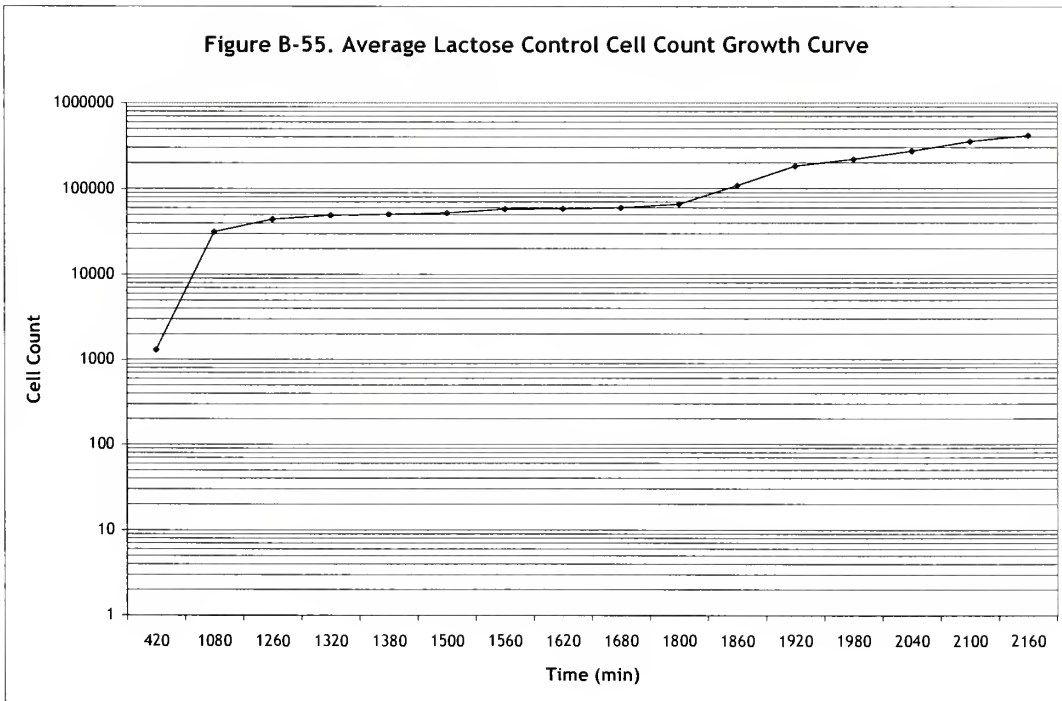
Cell Count increases from 90000 to 180000

Actual growth rate = (1920min - 1860min) = 60 min



Cell Count increases from 125000 to 250000

Actual growth rate = (1980min - 1860min) = 120 min



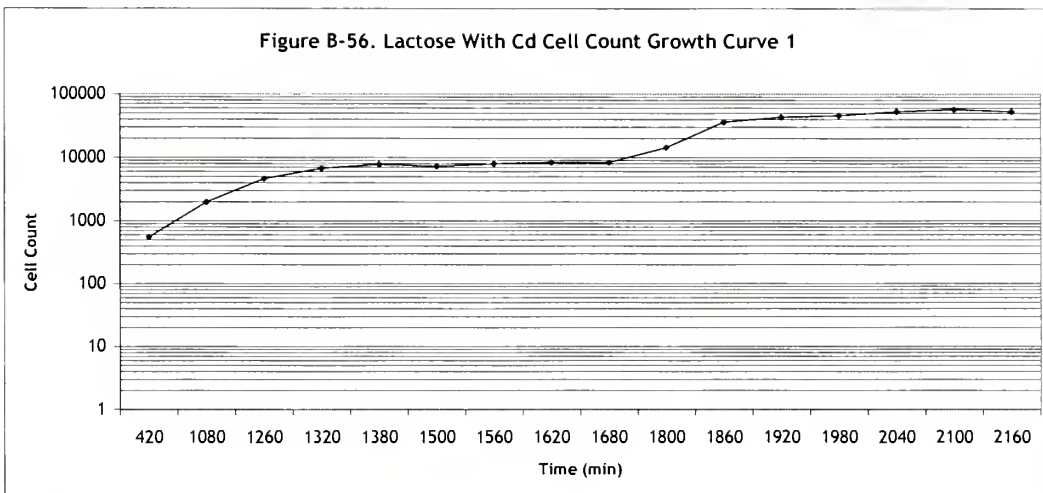
Cell Count increases from 109000 to 220500

Actual growth rate = (1980min - 1860min) = 120 min

LACTOSE WITH Cd CELL COUNT

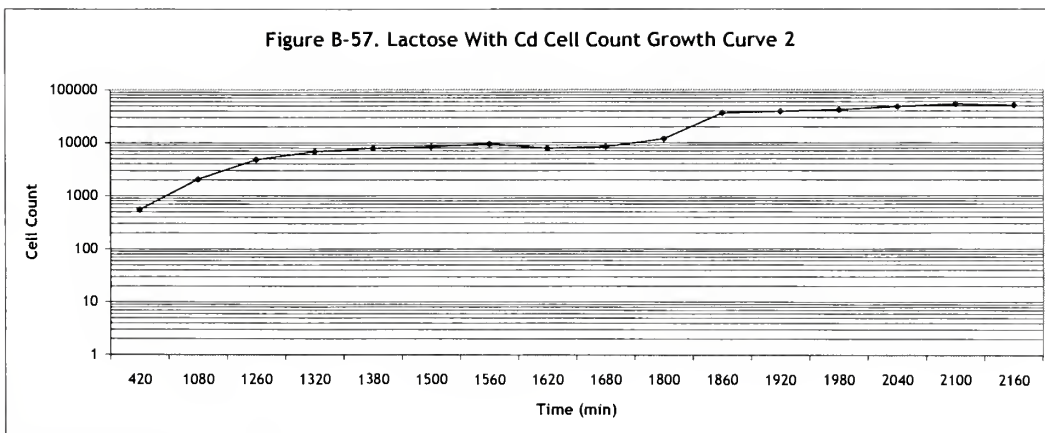
Time(min) Cell Count 1 Cell Count 2 Cell Count 3 Cell Count 4 Average Cell Count

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Cell Count 4	Average Cell Count
420	550	550	550		550
1080	2000	2050	2000		2017
1260	4620	4800	5400		4940
1320	6800	6800	5000		6200
1380	7800	7800	6000		7200
1500	7260	8400	6000		7220
1560	7980	9600	6000		7860
1620	8400	7800	9000		8400
1680	8400	8400	9000		8600
1800	14400	12000	12000		12800
1860	36300	36300	30000		34200
1920	43200	39000	39000		40400
1980	45900	42000	50000	60000	49475
2040	53100	48000	60000	50000	52775
2100	57600	54000	60000	50000	55400
2160	52800	51000	60000	50000	53450



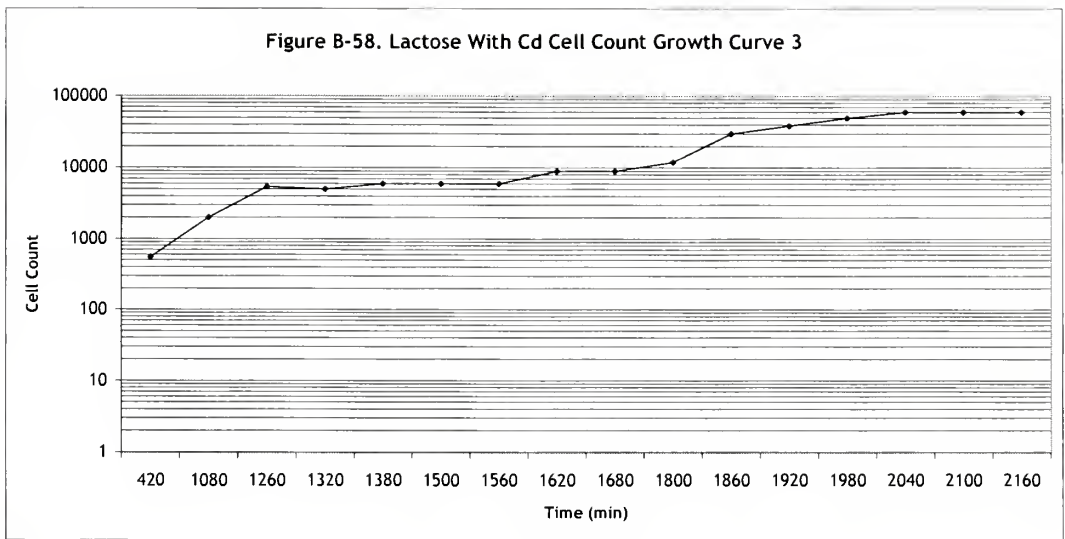
Cell Count increases from 7260 to 14400

Actual growth rate = (1800min - 1500min) = 300 min



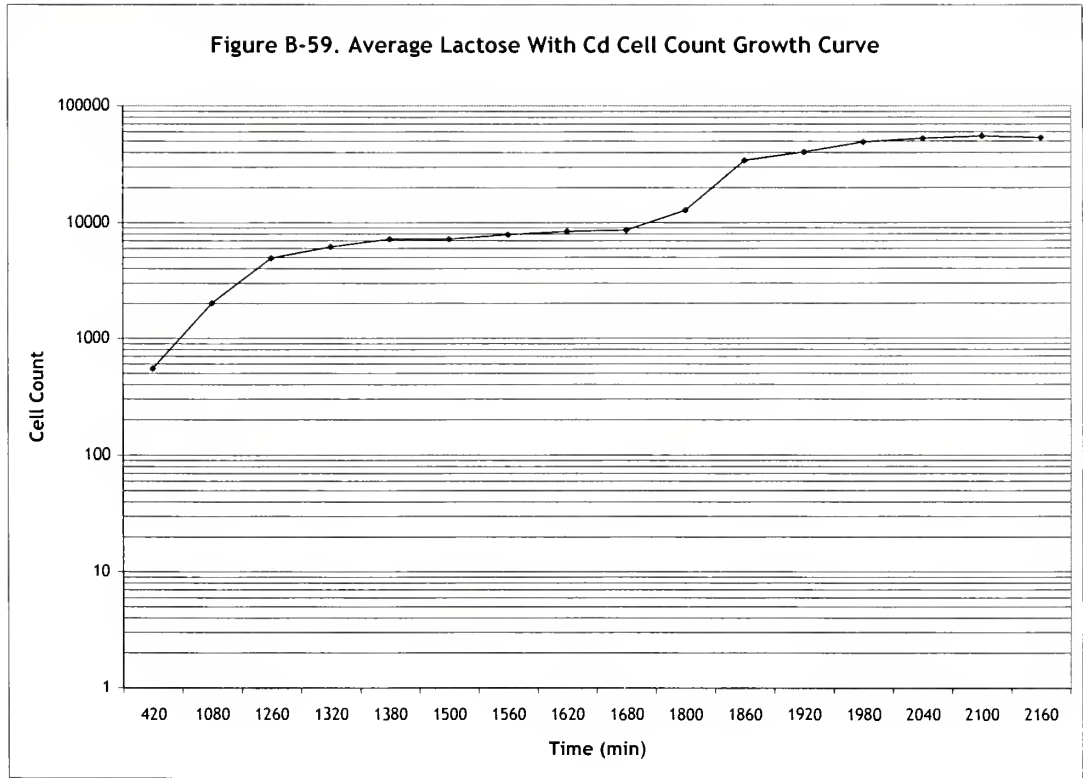
Cell Count increases from 4800 to 9600

Actual growth rate = (1560min - 1260min) = 300 min



Cell Count increases from 30000 to 60000

Actual growth rate = (2040min - 1860min) = 180 min

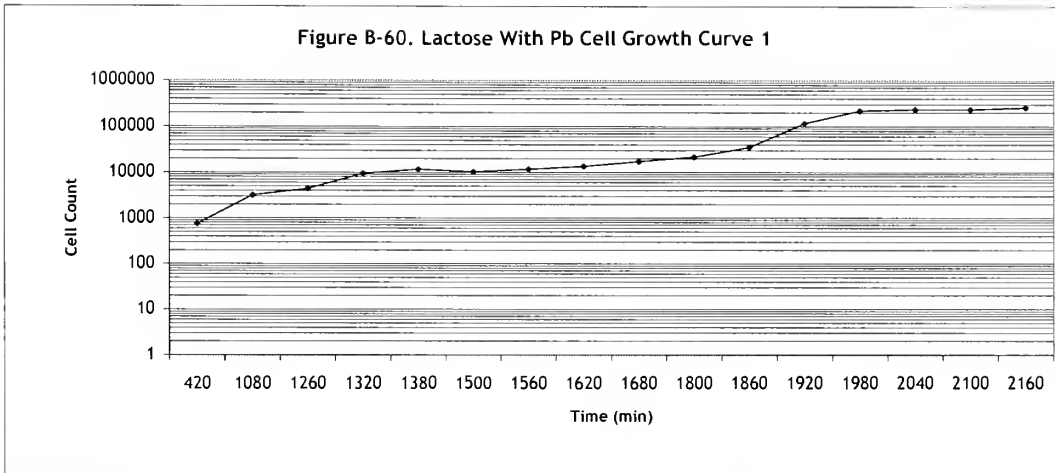


Cell Count increases from 6200 to 12800

Actual growth rate = (1800min - 1380min) = 420 min

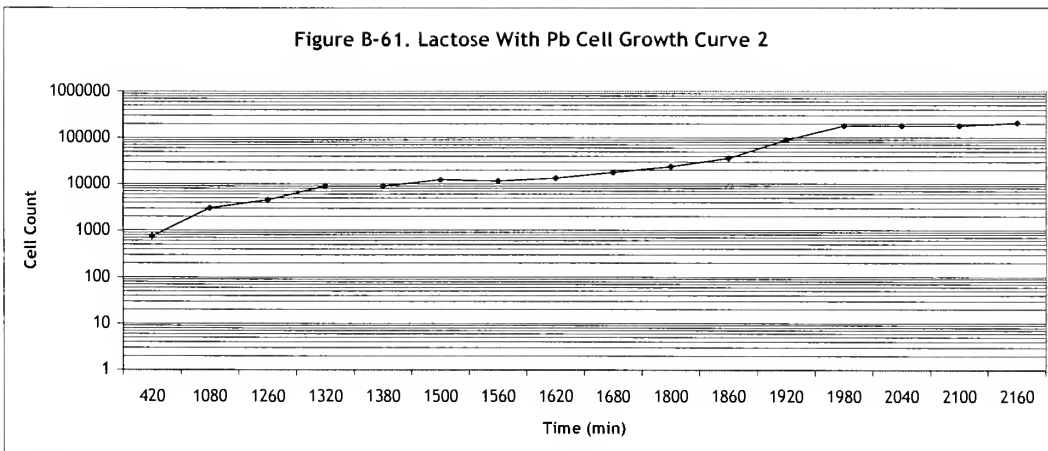
LACTOSE WITH Pb CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Cell Count 4	Average Cell Count
420	750	750	750		750
1080	3175	3050	3000		3075
1260	4440	4500	5400		4780
1320	9500	9000	10000		9500
1380	11700	9000	11700		10800
1500	10200	12300	12000		11500
1560	11700	11700	12000		11800
1620	13500	13500	12000		13000
1680	17400	18000	18000		17800
1800	21600	24000	30000		25200
1860	35100	36000	36000	30000	34275
1920	117000	90000	90000		99000
1980	222000	180000	200000	125000	181750
2040	234000	180000	200000	250000	216000
2100	237000	180000	200000	250000	216750
2160	258000	210000	200000	250000	229500



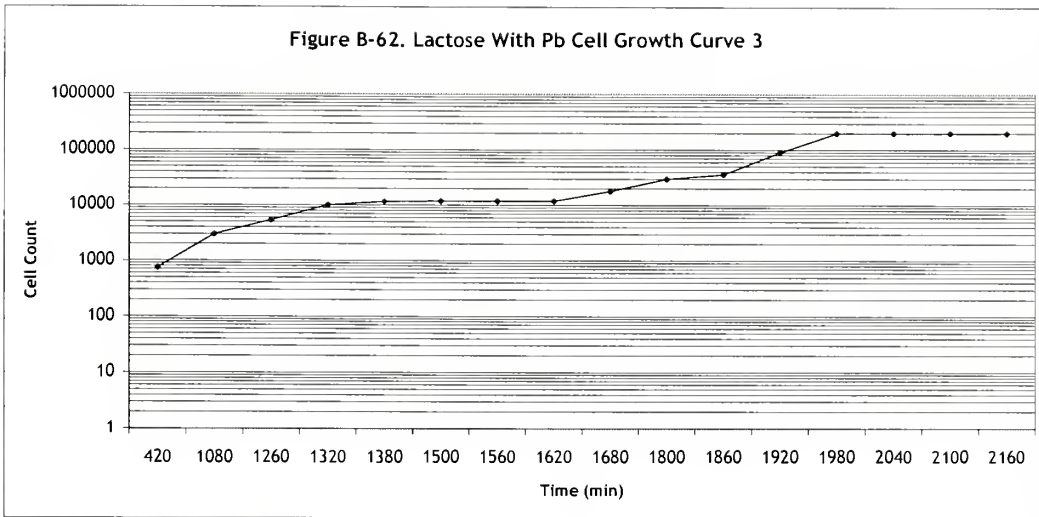
Cell Count increases from 17400 to 35100

Actual growth rate = (1860min - 1680min) = 180 min



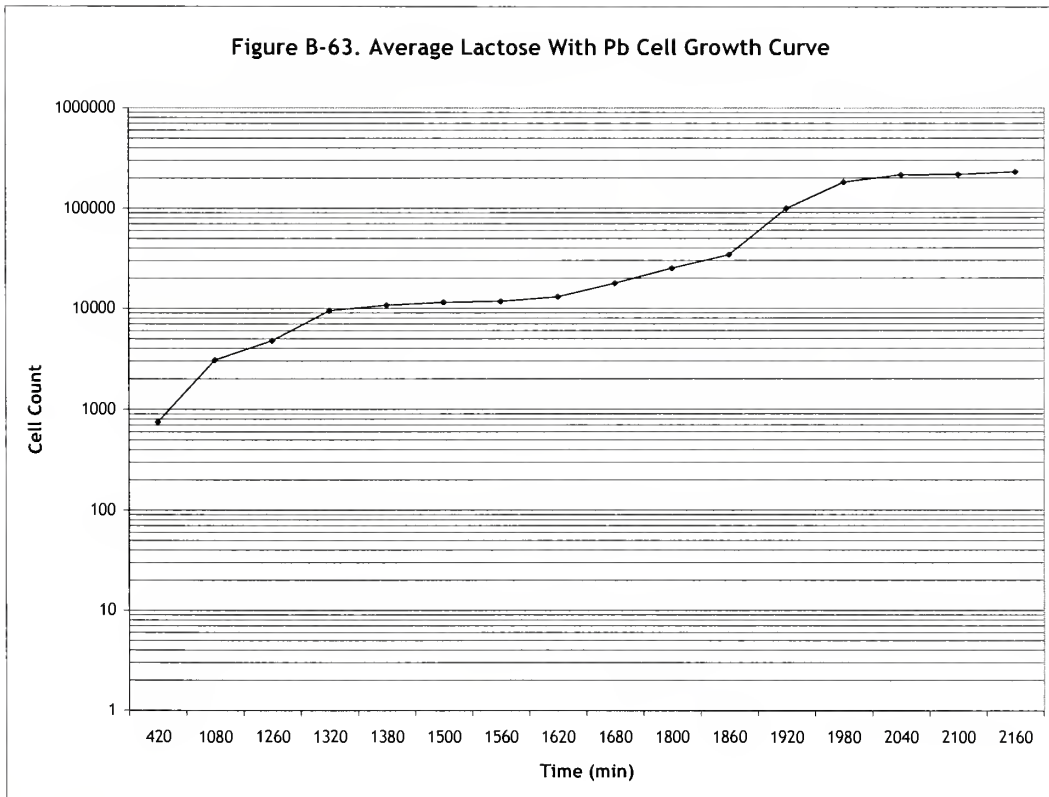
Cell Count increases from 18000 to 36000

Actual growth rate = (1860min - 1680min) = 180 min



Cell Count increases from 18000 to 36000

Actual growth rate = (1860min - 1680min) = 180 min

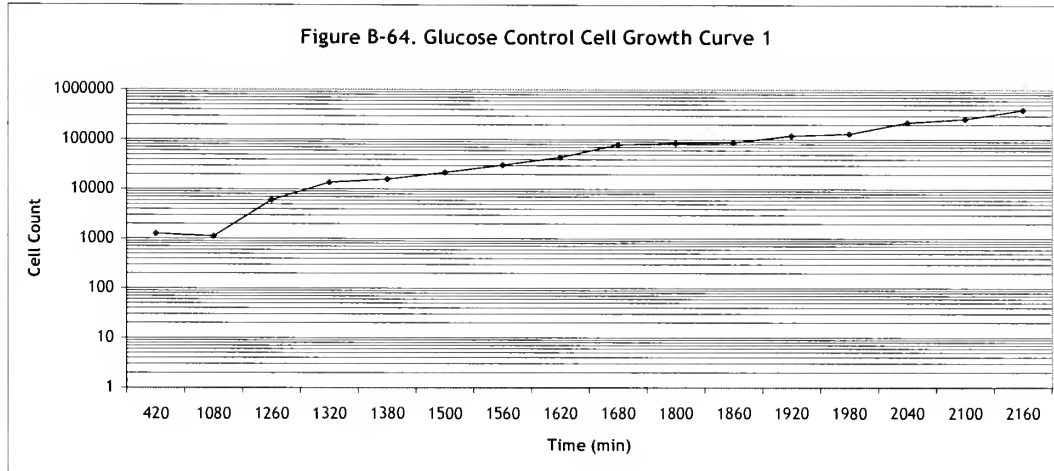


Cell Count increases from 13000 to 25200

Actual growth rate = (1800min - 1620min) = 180 min

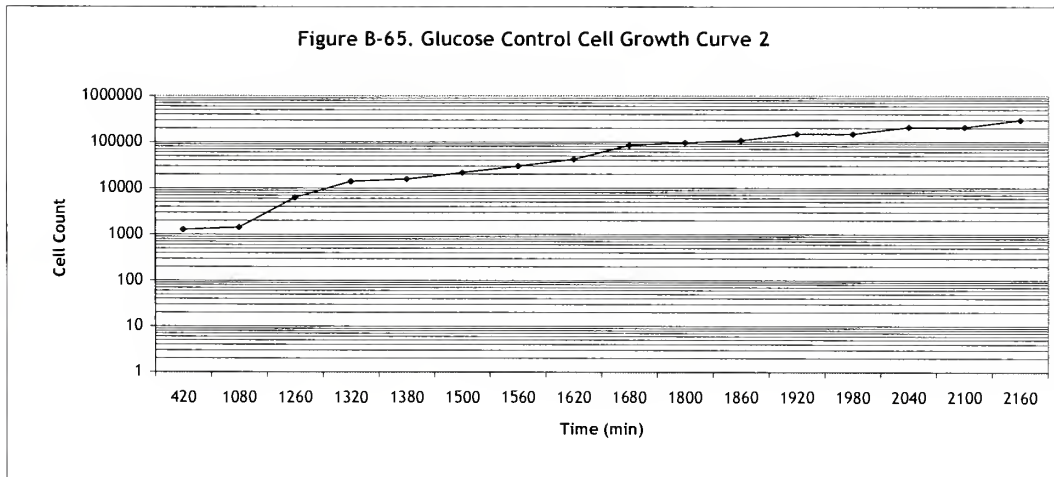
GLUCOSE ONLY CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Cell Count 4	Average Cell Count
420	1265	1265	1265		1265
1080	1120	1425	1450	1500	1374
1260	6090	6300	6300		6230
1320	13800	14000	10000		12600
1380	15900	15900	15000		15600
1500	21900	21900	18000		20600
1560	30600	30000	30000		30200
1620	44100	42000	60000		48700
1680	78900	84000	90000	75000	81975
1800	85500	96000	90000		90500
1860	88200	108000	100000	90000	96550
1920	120000	150000	150000		140000
1980	132000	150000	175000	125000	145500
2040	222000	210000	200000	250000	220500
2100	261000	210000	225000	250000	236500
2160	399000	300000	275000	250000	306000



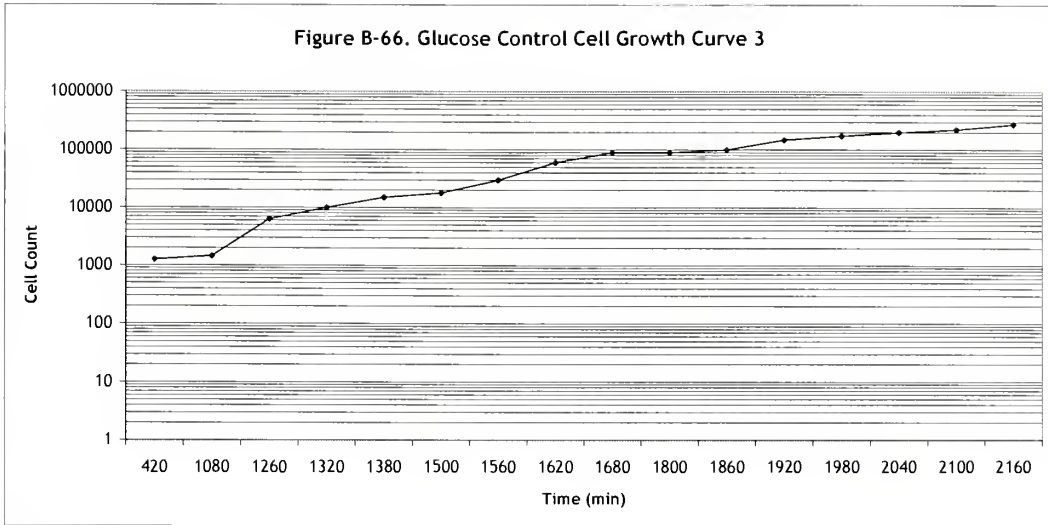
Cell Count increases from 21900 to 44100

Actual growth rate = (1620min - 1500min) = 120 min



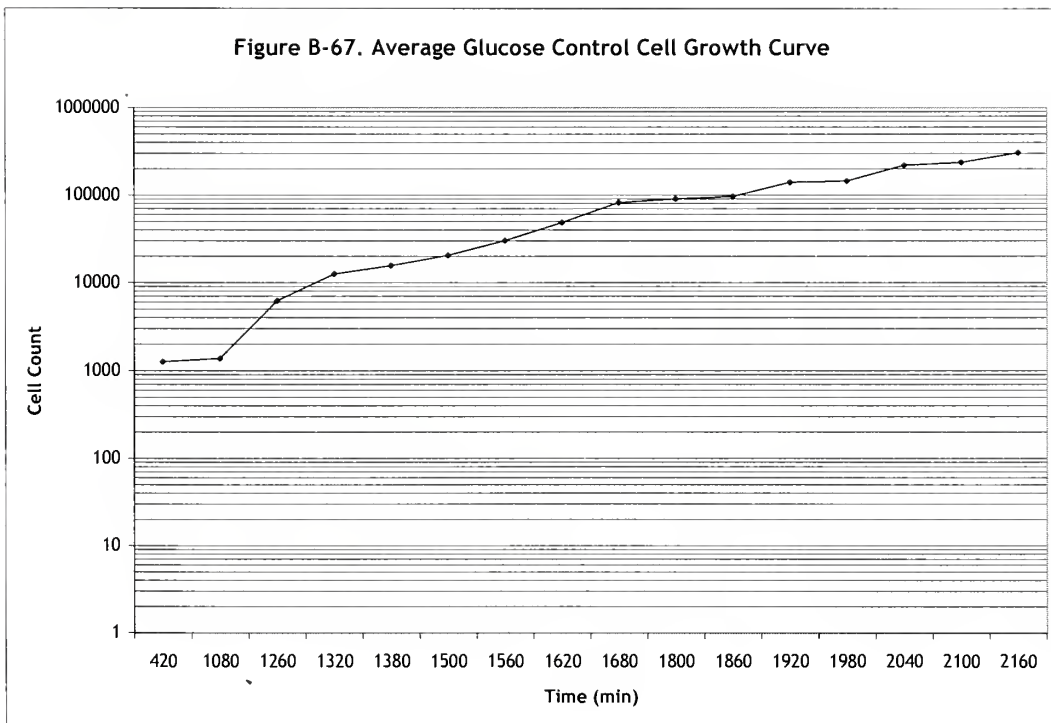
Cell Count increases from 42000 to 84000

Actual growth rate = (1680min - 1620min) = 60 min



Cell Count increases from 30000 to 60000

Actual growth rate = (1620min - 1560min) = 60 min



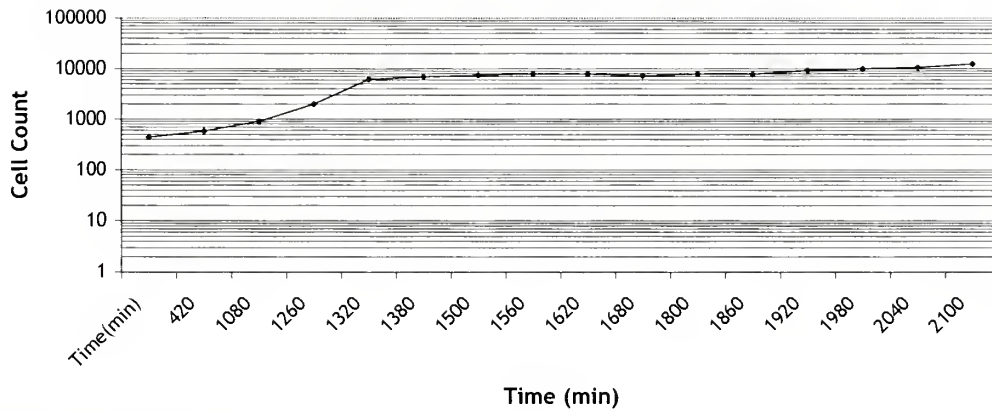
Cell Count increases from 15600 to 30200

Actual growth rate = (1560min - 1380min) = 180 min

GLUCOSE WITH Cd CELL COUNT

Time(min)	Cell Count 1	Cell Count 2	Cell Count 3	Average Cell Count
420	440	440	440	440
1080	580	550	580	570
1260	900	1200	900	1000
1320	2000	2000	2000	2000
1380	6180	5700	6000	5960
1500	6930	6000	6000	6310
1560	7470	6900	6000	6790
1620	7830	7200	6000	7010
1680	7890	7200	6000	7030
1800	7200	9000	9000	8400
1860	7860	8700	9000	8520
1920	7800	9000	9000	8600
1980	9300	9300	9000	9200
2040	9900	12000	12000	11300
2100	10500	12000	12000	11500
2160	12300	12300	12000	12200

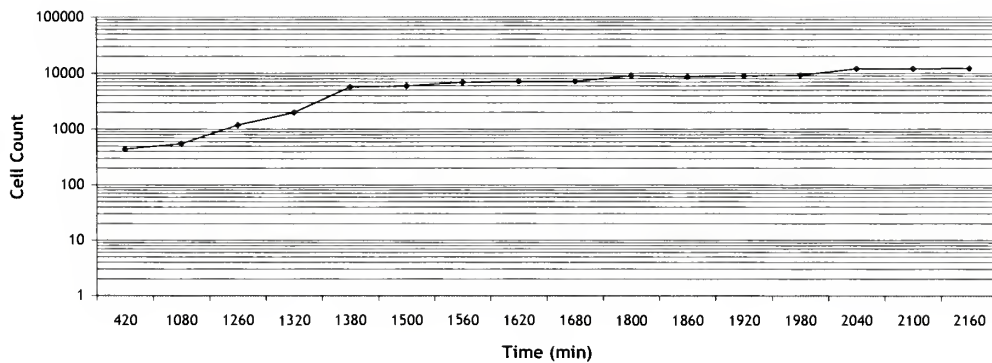
Figure B-68. Glucose With Cd Cell Growth Curve 1



Cell Count increases from 6180 to 12300

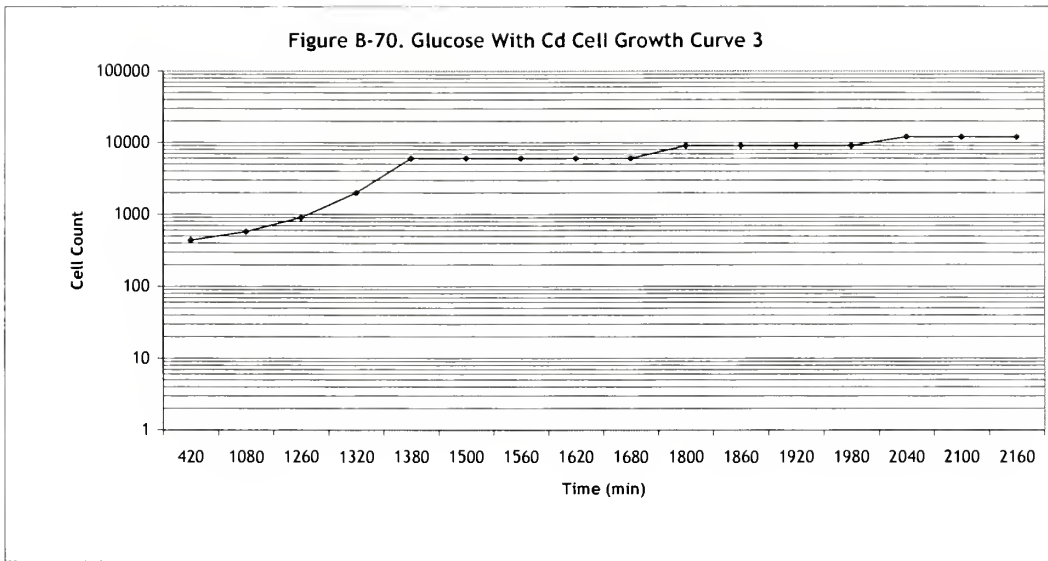
Actual growth rate = (2160min - 1500min) = 660 min

Figure B-69. Glucose With Cd Cell Growth Curve 2



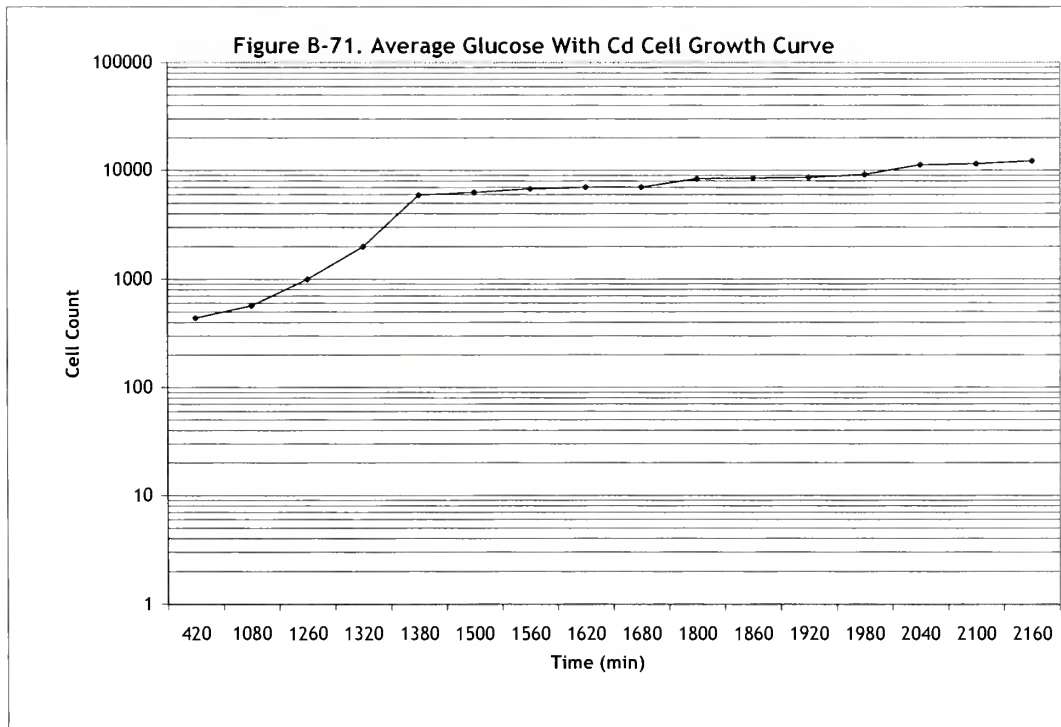
Cell Count increases from 6000 to 12000

Actual growth rate = (2040min - 1500min) = 540 min



Cell Count increases from 6000 to 12000

Actual growth rate = (2040min - 1680min) = 360 min



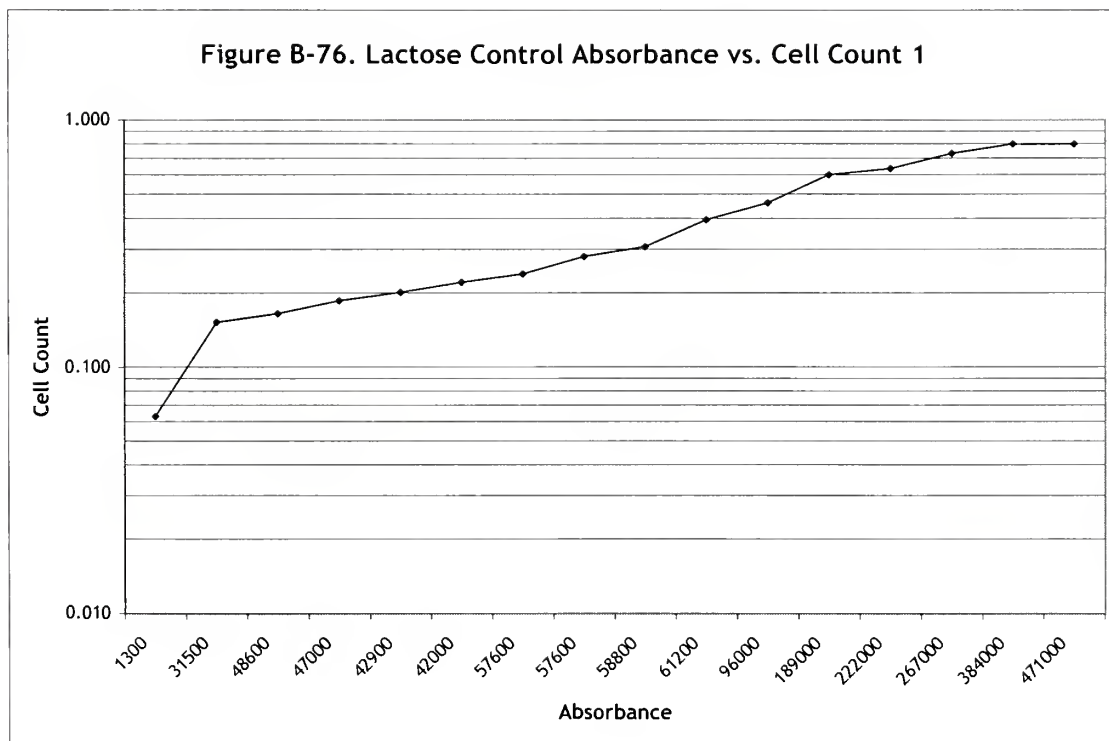
Cell Count increases from 5960 to 11500

Actual growth rate = (2100min - 1380min) = 720 min

M.roseus Cell growth vs. Absorbance

LACTOSE ONLY CELL COUNT

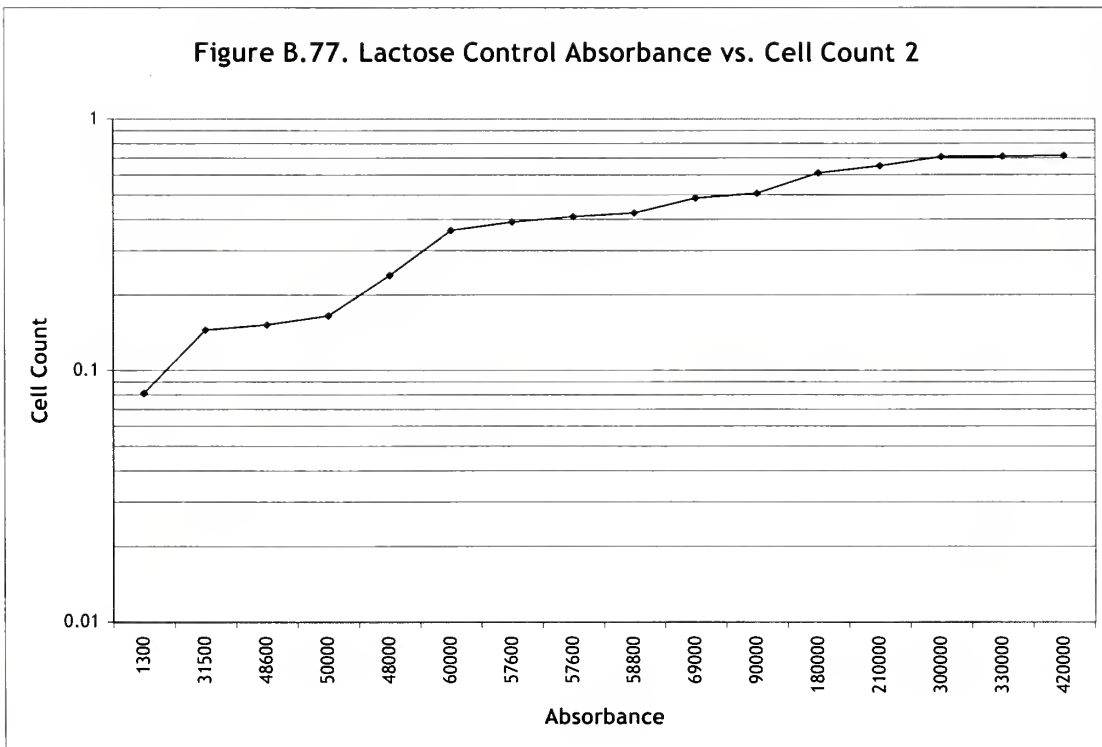
Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1300	0.063	1.16	0.865	1300	0.063
1080	31500	0.152	1.42	0.705	31500	0.152
1260	48600	0.165	1.46	0.684	48600	0.165
1320	47000	0.186	1.53	0.652	47000	0.186
1380	42900	0.201	1.59	0.630	42900	0.201
1500	42000	0.221	1.66	0.601	42000	0.221
1560	57600	0.239	1.73	0.577	57600	0.239
1620	57600	0.281	1.91	0.524	57600	0.281
1680	58800	0.308	2.03	0.492	58800	0.308
1800	61200	0.395	2.48	0.403	61200	0.395
1860	96000	0.461	2.89	0.346	96000	0.461
1920	189000	0.599	3.97	0.252	189000	0.599
1980	222000	0.635	4.32	0.232	222000	0.635
2040	267000	0.732	5.40	0.185	267000	0.732
2100	384000	0.798	6.28	0.159	384000	0.798
2160	471000	0.798	6.28	0.159	471000	0.798



Cell Count increases from 96000 to 189000

Difference in absorbance = (0.599 - 0.461) = 0.138

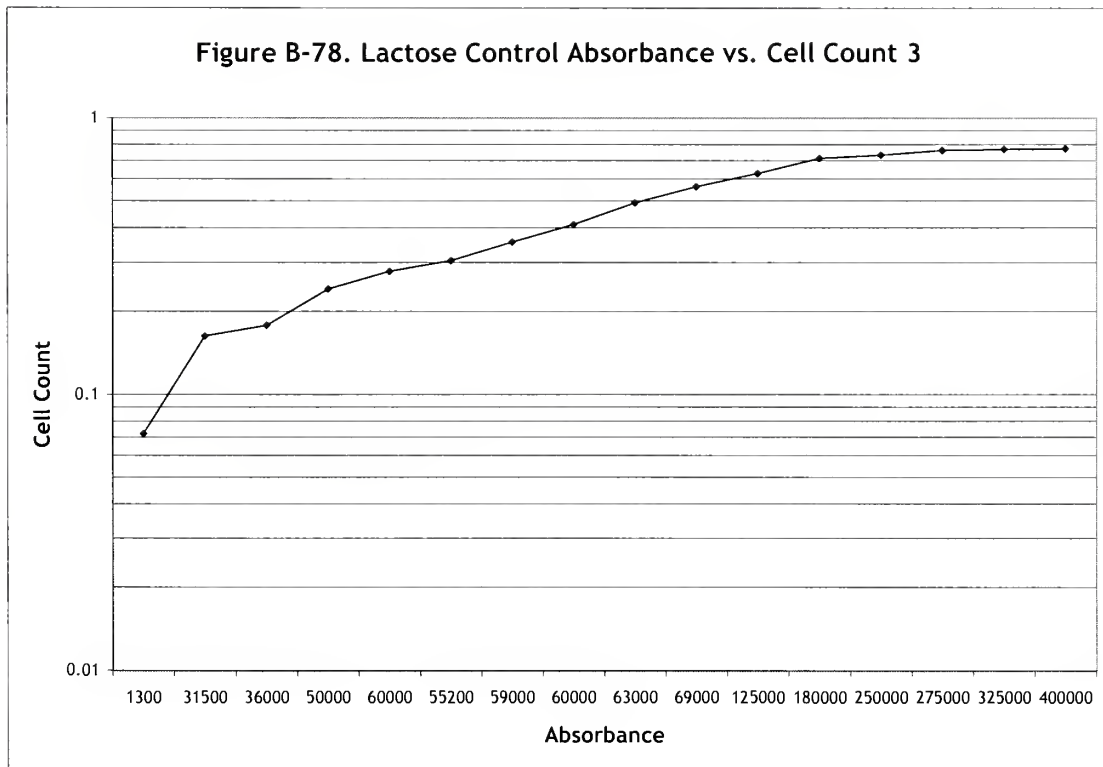
Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1300	0.081	1.21	0.830	1300	0.081
1080	31500	0.145	1.40	0.716	31500	0.145
1260	48600	0.152	1.42	0.705	48600	0.152
1320	50000	0.165	1.46	0.684	50000	0.165
1380	48000	0.239	1.73	0.577	48000	0.239
1500	60000	0.362	2.30	0.435	60000	0.362
1560	57600	0.391	2.46	0.406	57600	0.391
1620	57600	0.411	2.58	0.388	57600	0.411
1680	58800	0.424	2.65	0.377	58800	0.424
1800	69000	0.485	3.05	0.327	69000	0.485
1860	90000	0.508	3.22	0.310	90000	0.508
1920	180000	0.612	4.09	0.244	180000	0.612
1980	210000	0.653	4.50	0.222	210000	0.653
2040	300000	0.708	5.11	0.196	300000	0.708
2100	330000	0.711	5.14	0.195	330000	0.711
2160	420000	0.715	5.19	0.193	420000	0.715



Cell Count increases from 90000 to 180000

Difference in absorbance = (0.612 - 0.508) = 0.104

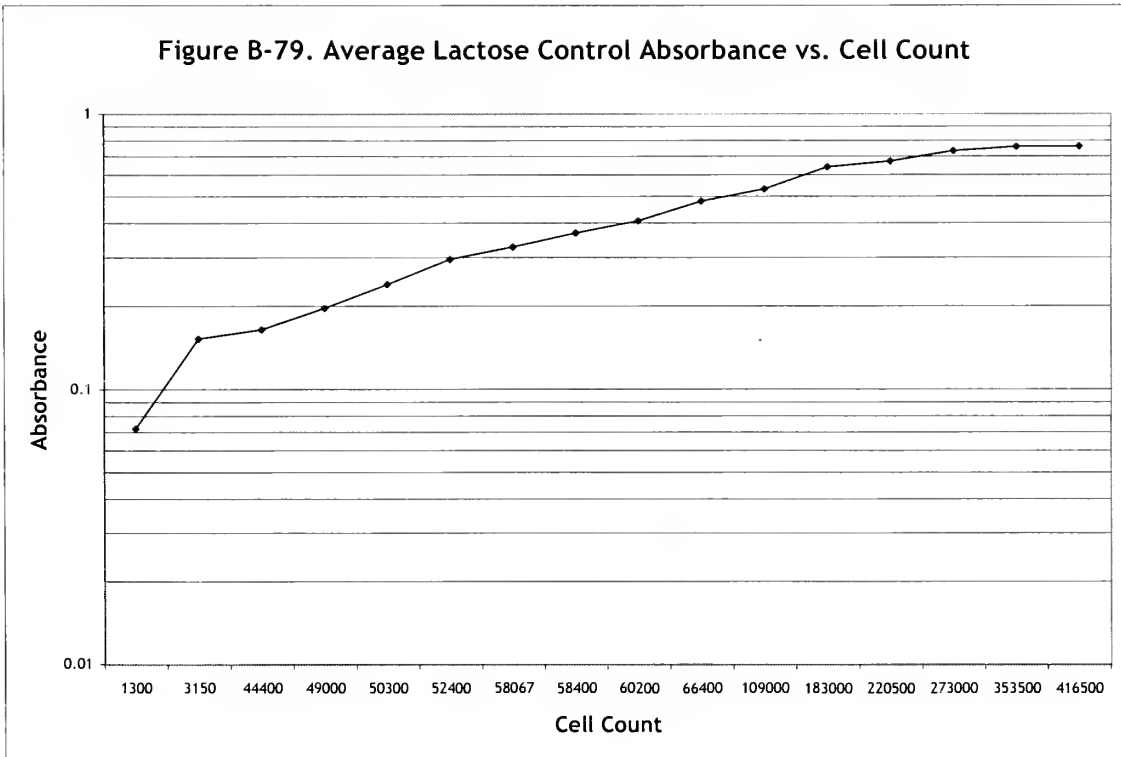
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1300	0.072	1.18	0.847	1300	0.072
1080	31500	0.163	1.46	0.687	31500	0.163
1260	36000	0.178	1.51	0.664	36000	0.178
1320	50000	0.241	1.74	0.574	50000	0.241
1380	60000	0.279	1.90	0.526	60000	0.279
1500	55200	0.305	2.02	0.495	55200	0.305
1560	59000	0.355	2.26	0.442	59000	0.355
1620	60000	0.411	2.58	0.388	60000	0.411
1680	63000	0.492	3.10	0.322	63000	0.492
1800	69000	0.563	3.66	0.274	69000	0.563
1860	125000	0.628	4.25	0.236	125000	0.628
1920	180000	0.713	5.16	0.194	180000	0.713
1980	250000	0.732	5.40	0.185	250000	0.732
2040	275000	0.763	5.79	0.173	275000	0.763
2100	325000	0.768	5.86	0.171	325000	0.768
2160	400000	0.773	5.93	0.169	400000	0.773



Cell Count increases from 125000 to 250000

Difference in absorbance = (0.732 - 0.628) = 0.104

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1300	0.072	1.18	0.847	1300	0.072
1080	3150	0.153	1.42	0.703	3150	0.153
1260	44400	0.165	1.46	0.684	44400	0.165
1320	49000	0.197	1.58	0.635	49000	0.197
1380	50300	0.240	1.74	0.576	50300	0.24
1500	52400	0.296	1.98	0.506	52400	0.296
1560	58067	0.328	2.13	0.470	58067	0.328
1620	58400	0.368	2.33	0.429	58400	0.368
1680	60200	0.408	2.56	0.391	60200	0.408
1800	66400	0.481	3.03	0.330	66400	0.481
1860	109000	0.532	3.41	0.294	109000	0.532
1920	183000	0.641	4.38	0.228	183000	0.641
1980	220500	0.673	4.71	0.212	220500	0.673
2040	273000	0.734	5.42	0.184	273000	0.734
2100	353500	0.759	5.74	0.174	353500	0.759
2160	416500	0.762	5.78	0.173	416500	0.762

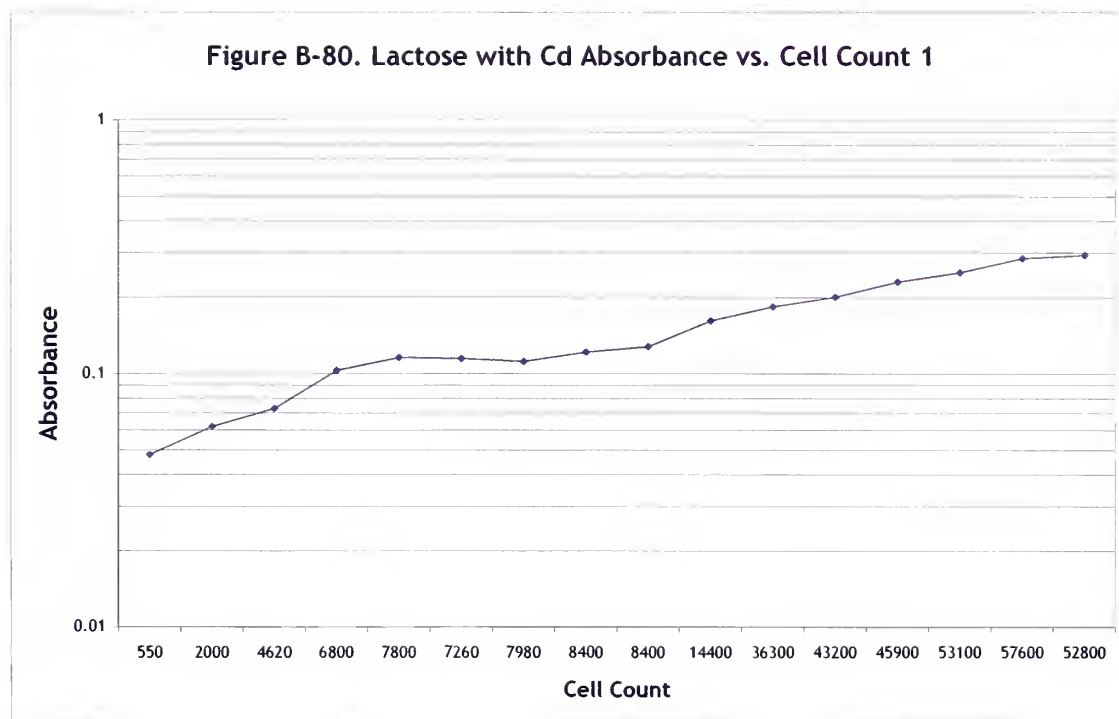


Cell Count increases from 109000 to 220500

Difference in absorbance = (0.673 - 0.532) = 0.141

LACTOSE WITH Cd CELL COUNT

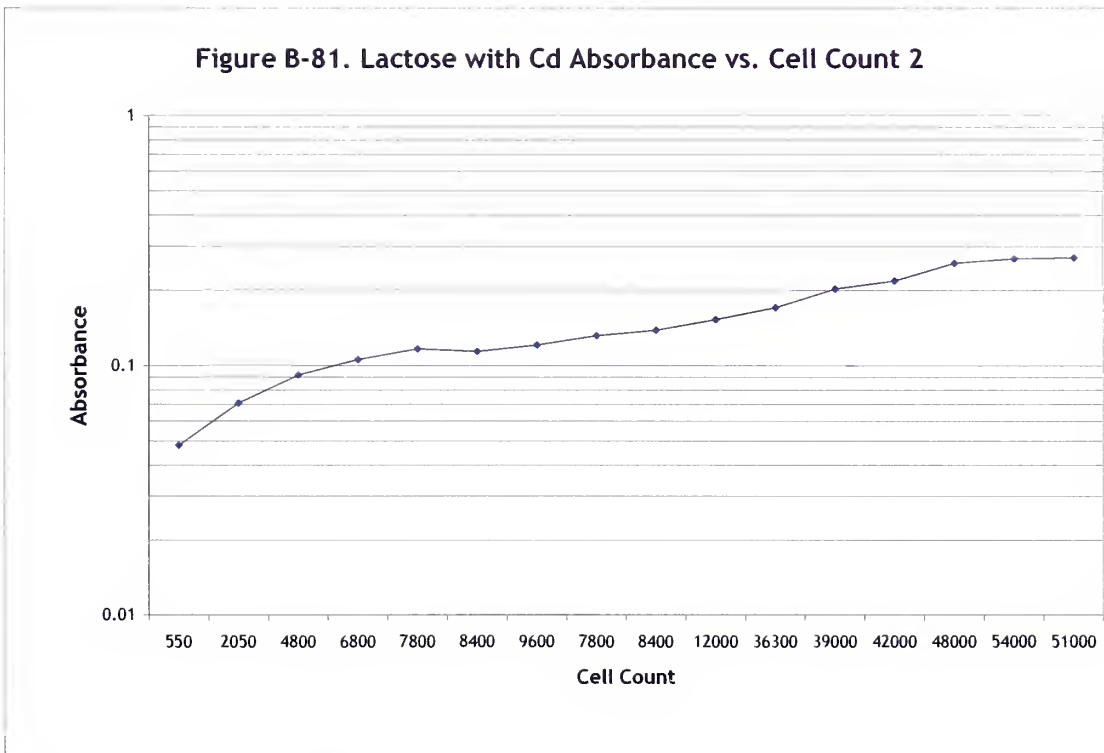
Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	550	0.048	1.12	0.895	550	0.048
1080	2000	0.062	1.15	0.867	2000	0.062
1260	4620	0.073	1.18	0.845	4620	0.073
1320	6800	0.103	1.27	0.789	6800	0.103
1380	7800	0.116	1.31	0.766	7800	0.116
1500	7260	0.115	1.30	0.767	7260	0.115
1560	7980	0.112	1.29	0.773	7980	0.112
1620	8400	0.122	1.32	0.755	8400	0.122
1680	8400	0.128	1.34	0.745	8400	0.128
1800	14400	0.162	1.45	0.689	14400	0.162
1860	36300	0.184	1.53	0.655	36300	0.184
1920	43200	0.201	1.59	0.630	43200	0.201
1980	45900	0.231	1.70	0.587	45900	0.231
2040	53100	0.251	1.78	0.561	53100	0.251
2100	57600	0.286	1.93	0.518	57600	0.286
2160	52800	0.294	1.97	0.508	52800	0.294



Cell Count increases from 7260 to 14400

Difference in absorbance = (0.162 - 0.115) = 0.047

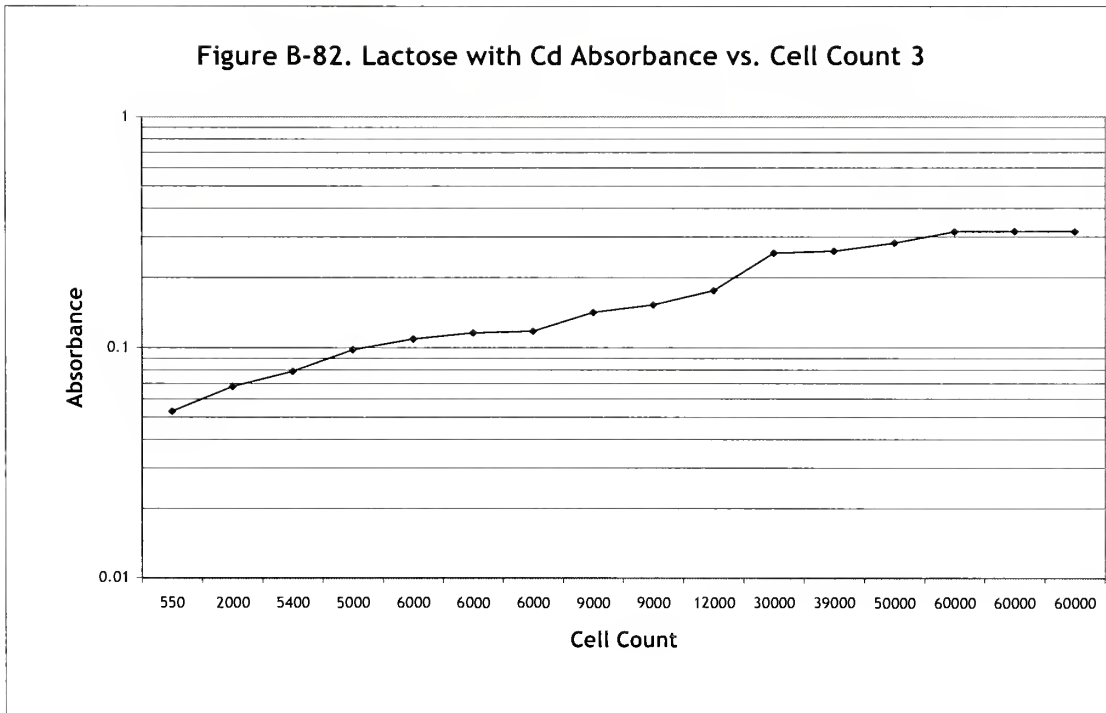
Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	550	0.048	1.12	0.895	550	0.048
1080	2050	0.071	1.18	0.849	2050	0.071
1260	4800	0.092	1.24	0.809	4800	0.092
1320	6800	0.106	1.28	0.783	6800	0.106
1380	7800	0.117	1.31	0.764	7800	0.117
1500	8400	0.114	1.30	0.769	8400	0.114
1560	9600	0.121	1.32	0.757	9600	0.121
1620	7800	0.132	1.36	0.738	7800	0.132
1680	8400	0.139	1.38	0.726	8400	0.139
1800	12000	0.153	1.42	0.703	12000	0.153
1860	36300	0.171	1.48	0.675	36300	0.171
1920	39000	0.203	1.60	0.627	39000	0.203
1980	42000	0.219	1.66	0.604	42000	0.219
2040	48000	0.258	1.81	0.552	48000	0.258
2100	54000	0.269	1.86	0.538	54000	0.269
2160	51000	0.271	1.87	0.536	51000	0.271



Cell Count increases from 4800 to 9600

Difference in absorbance = (0.121 - 0.071) = 0.05

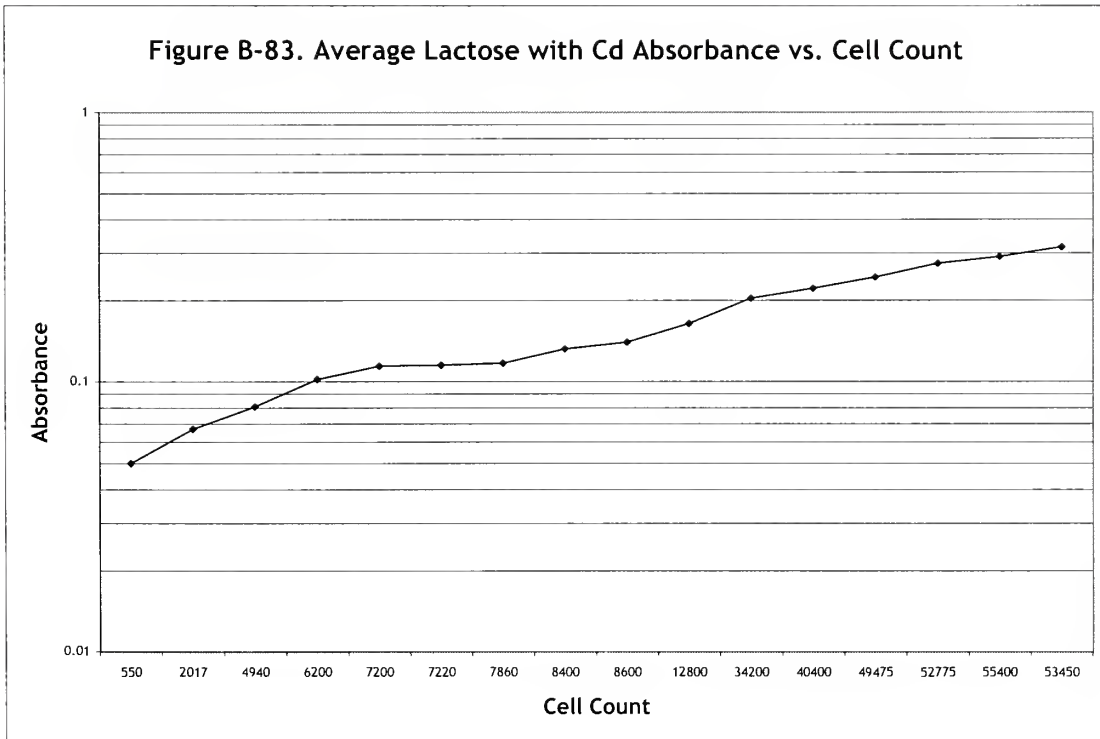
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	550	0.053	1.13	0.885	550	0.053
1080	2000	0.068	1.17	0.855	2000	0.068
1260	5400	0.079	1.20	0.834	5400	0.079
1320	5000	0.098	1.25	0.798	5000	0.098
1380	6000	0.109	1.29	0.778	6000	0.109
1500	6000	0.116	1.31	0.766	6000	0.116
1560	6000	0.118	1.31	0.762	6000	0.118
1620	9000	0.142	1.39	0.721	9000	0.142
1680	9000	0.153	1.42	0.703	9000	0.153
1800	12000	0.176	1.50	0.667	12000	0.176
1860	30000	0.256	1.80	0.555	30000	0.256
1920	39000	0.261	1.82	0.548	39000	0.261
1980	50000	0.283	1.92	0.521	50000	0.283
2040	60000	0.316	2.07	0.483	60000	0.316
2100	60000	0.317	2.07	0.482	60000	0.317
2160	60000	0.316	2.07	0.483	60000	0.316



Cell Count increases from 30000 to 60000

Difference in absorbance = (0.316 - 0.256) = 0.06

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	550	0.050	1.12	0.892	550	0.05
1080	2017	0.067	1.17	0.857	2017	0.067
1260	4940	0.081	1.21	0.829	4940	0.081
1320	6200	0.102	1.27	0.790	6200	0.102
1380	7200	0.114	1.30	0.769	7200	0.114
1500	7220	0.115	1.30	0.767	7220	0.115
1560	7860	0.117	1.31	0.764	7860	0.117
1620	8400	0.132	1.36	0.738	8400	0.132
1680	8600	0.140	1.38	0.724	8600	0.14
1800	12800	0.164	1.46	0.686	12800	0.164
1860	34200	0.204	1.60	0.626	34200	0.204
1920	40400	0.222	1.67	0.600	40400	0.222
1980	49475	0.244	1.76	0.570	49475	0.244
2040	52775	0.275	1.88	0.531	52775	0.275
2100	55400	0.291	1.95	0.512	55400	0.291
2160	53450	0.294	1.97	0.509	53450	0.316

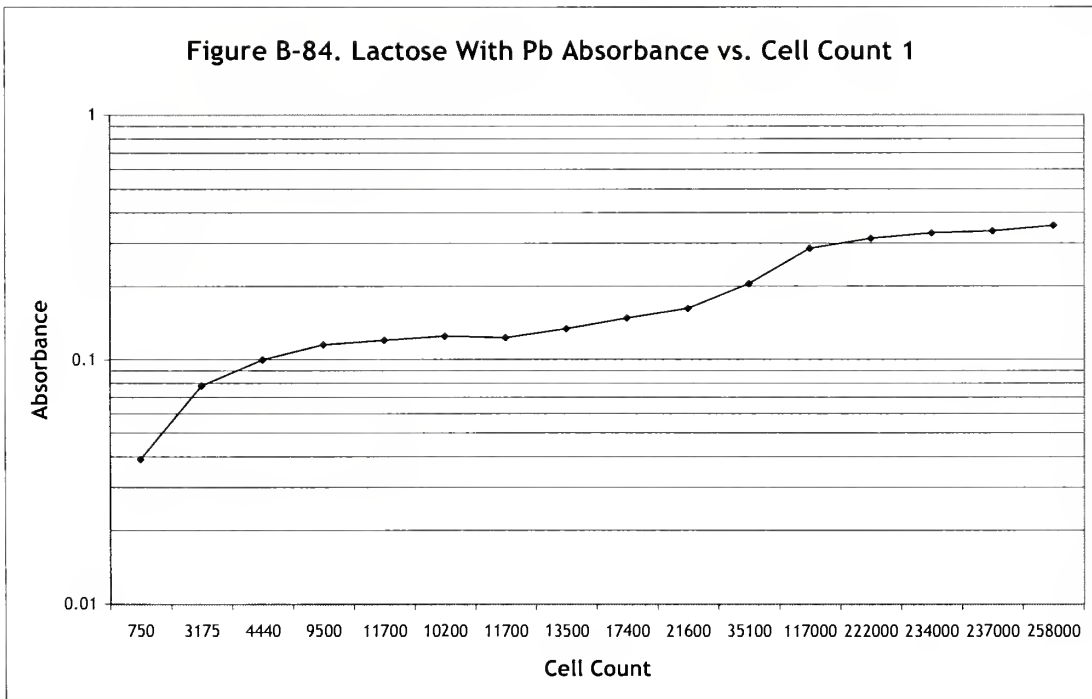


Cell Count increases from 6200 to 12800

Difference in absorbance = (0.164 - 0.102) = 0.062

LACTOSE WITH Pb CELL COUNT

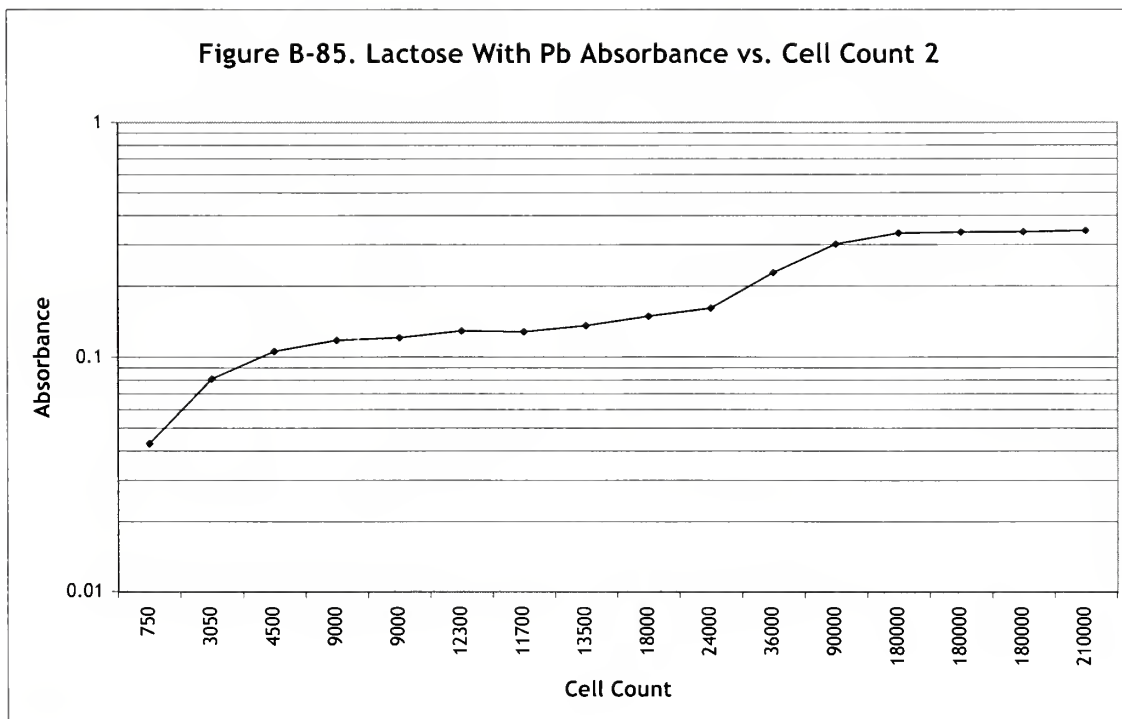
Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	750	0.039	1.09	0.914	750	0.039
1080	3175	0.078	1.20	0.836	3175	0.078
1260	4440	0.1	1.26	0.794	4440	0.1
1320	9500	0.115	1.30	0.767	9500	0.115
1380	11700	0.12	1.32	0.759	11700	0.12
1500	10200	0.125	1.33	0.750	10200	0.125
1560	11700	0.123	1.33	0.753	11700	0.123
1620	13500	0.134	1.36	0.735	13500	0.134
1680	17400	0.148	1.41	0.711	17400	0.148
1800	21600	0.162	1.45	0.689	21600	0.162
1860	35100	0.205	1.60	0.624	35100	0.205
1920	117000	0.286	1.93	0.518	117000	0.286
1980	222000	0.314	2.06	0.485	222000	0.314
2040	234000	0.331	2.14	0.467	234000	0.331
2100	237000	0.337	2.17	0.460	237000	0.337
2160	258000	0.355	2.26	0.442	258000	0.355



Cell Count increases from 17400 to 35100

Difference in absorbance = (0.205 - 0.148) = 0.057

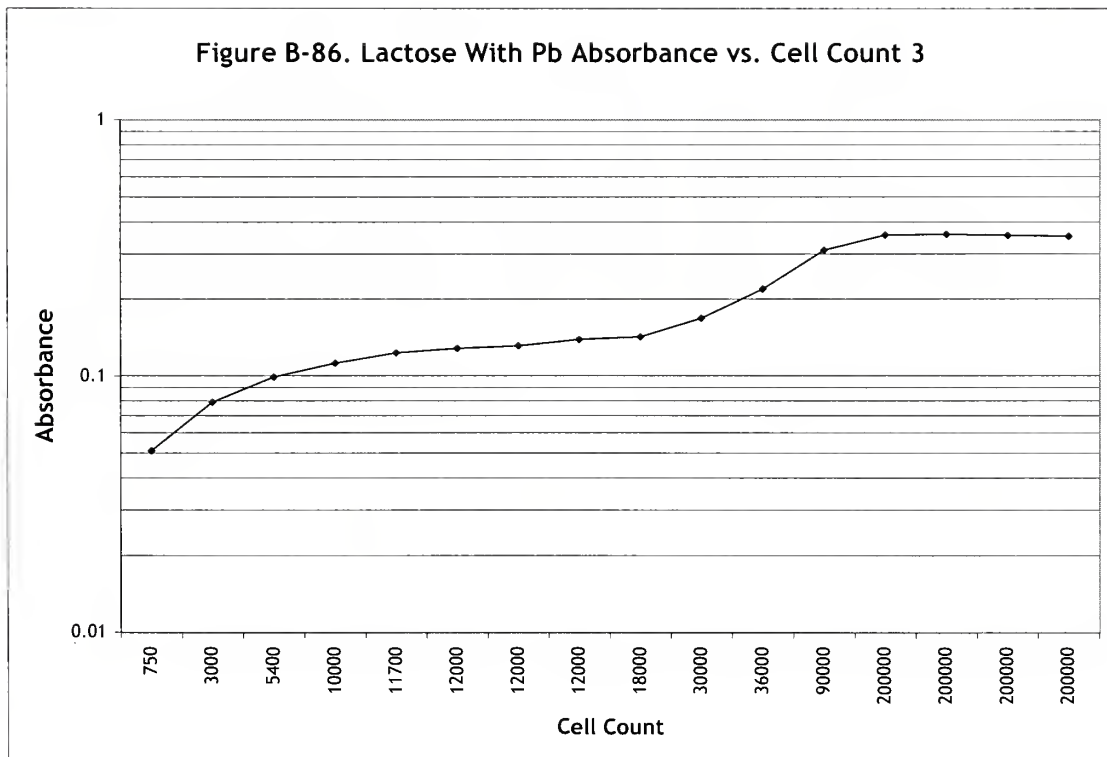
Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	750	0.043	1.10	0.906	750	0.043
1080	3050	0.081	1.21	0.830	3050	0.081
1260	4500	0.106	1.28	0.783	4500	0.106
1320	9000	0.118	1.31	0.762	9000	0.118
1380	9000	0.121	1.32	0.757	9000	0.121
1500	12300	0.129	1.35	0.743	12300	0.129
1560	11700	0.128	1.34	0.745	11700	0.128
1620	13500	0.136	1.37	0.731	13500	0.136
1680	18000	0.149	1.41	0.710	18000	0.149
1800	24000	0.161	1.45	0.690	24000	0.161
1860	36000	0.228	1.69	0.592	36000	0.228
1920	90000	0.302	2.00	0.499	90000	0.302
1980	180000	0.336	2.17	0.461	180000	0.336
2040	180000	0.339	2.18	0.458	180000	0.339
2100	180000	0.341	2.19	0.456	180000	0.341
2160	210000	0.345	2.21	0.452	210000	0.345



Cell Count increases from 18000 to 36000

Difference in absorbance = (0.228 - 0.149) = 0.079

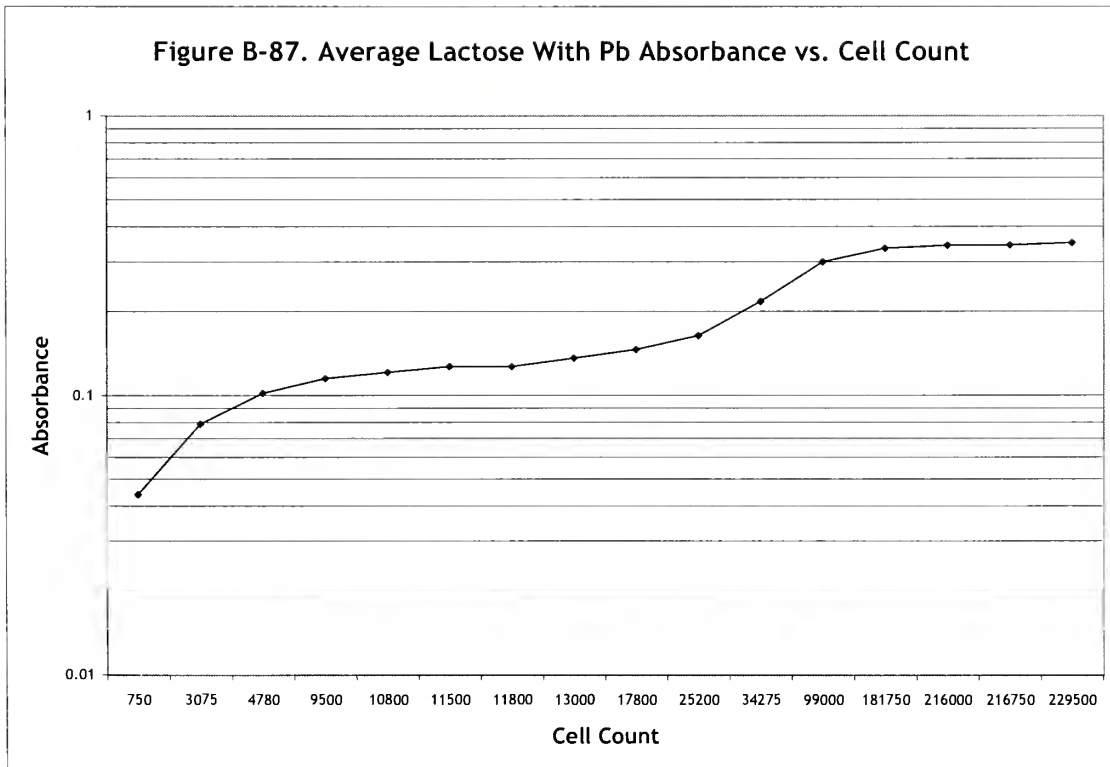
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	750	0.051	1.12	0.889	750	0.051
1080	3000	0.079	1.20	0.834	3000	0.079
1260	5400	0.099	1.26	0.796	5400	0.099
1320	10000	0.112	1.29	0.773	10000	0.112
1380	11700	0.123	1.33	0.753	11700	0.123
1500	12000	0.128	1.34	0.745	12000	0.128
1560	12000	0.131	1.35	0.740	12000	0.131
1620	12000	0.139	1.38	0.726	12000	0.139
1680	18000	0.142	1.39	0.721	18000	0.142
1800	30000	0.168	1.47	0.679	30000	0.168
1860	36000	0.219	1.66	0.604	36000	0.219
1920	90000	0.311	2.05	0.489	90000	0.311
1980	200000	0.356	2.27	0.441	200000	0.356
2040	200000	0.358	2.28	0.439	200000	0.358
2100	200000	0.355	2.26	0.442	200000	0.355
2160	200000	0.352	2.25	0.445	200000	0.352



Cell Count increases from 18000 to 36000

Difference in absorbance = (0.219 - 0.142) = 0.077

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	750	0.044	1.11	0.903	750	0.044
1080	3075	0.079	1.20	0.833	3075	0.079
1260	4780	0.102	1.26	0.791	4780	0.102
1320	9500	0.115	1.30	0.767	9500	0.115
1380	10800	0.121	1.32	0.756	10800	0.121
1500	11500	0.127	1.34	0.746	11500	0.127
1560	11800	0.127	1.34	0.746	11800	0.127
1620	13000	0.136	1.37	0.731	13000	0.136
1680	17800	0.146	1.40	0.714	17800	0.146
1800	25200	0.164	1.46	0.686	25200	0.164
1860	34275	0.217	1.65	0.606	34275	0.217
1920	99000	0.300	1.99	0.502	99000	0.3
1980	181750	0.335	2.16	0.462	181750	0.335
2040	216000	0.343	2.20	0.454	216000	0.343
2100	216750	0.344	2.21	0.453	216750	0.344
2160	229500	0.351	2.24	0.446	229500	0.351

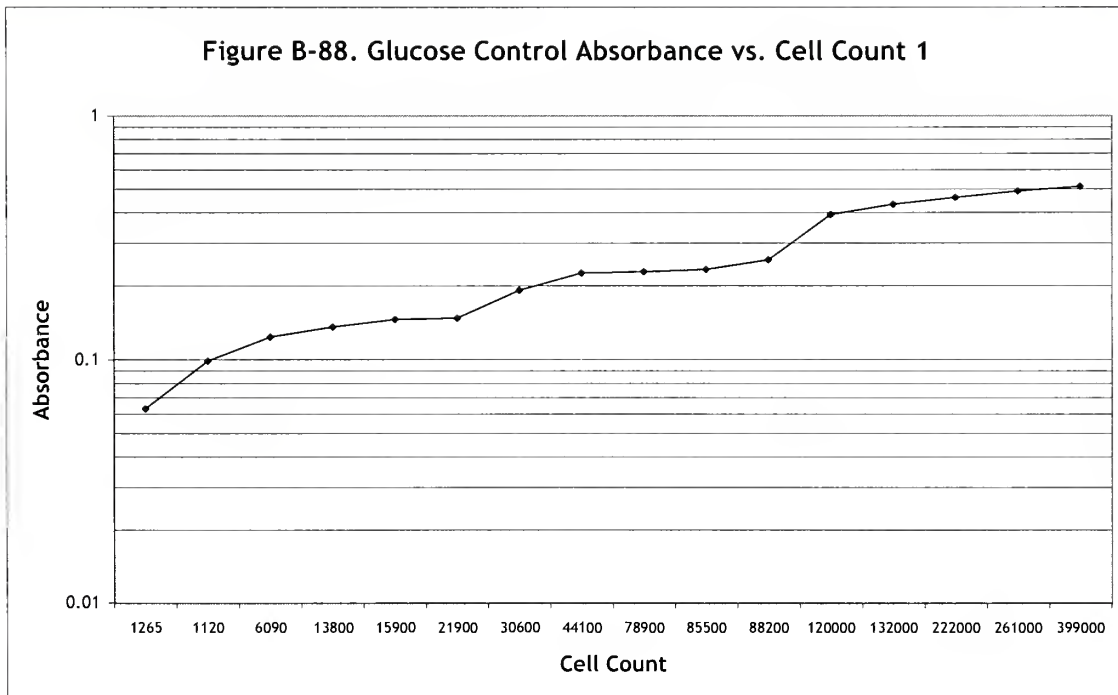


Cell Count increases from 13000 to 25200

Difference in absorbance = (0.164 - 0.136) = 0.028

GLUCOSE ONLY CELL COUNT

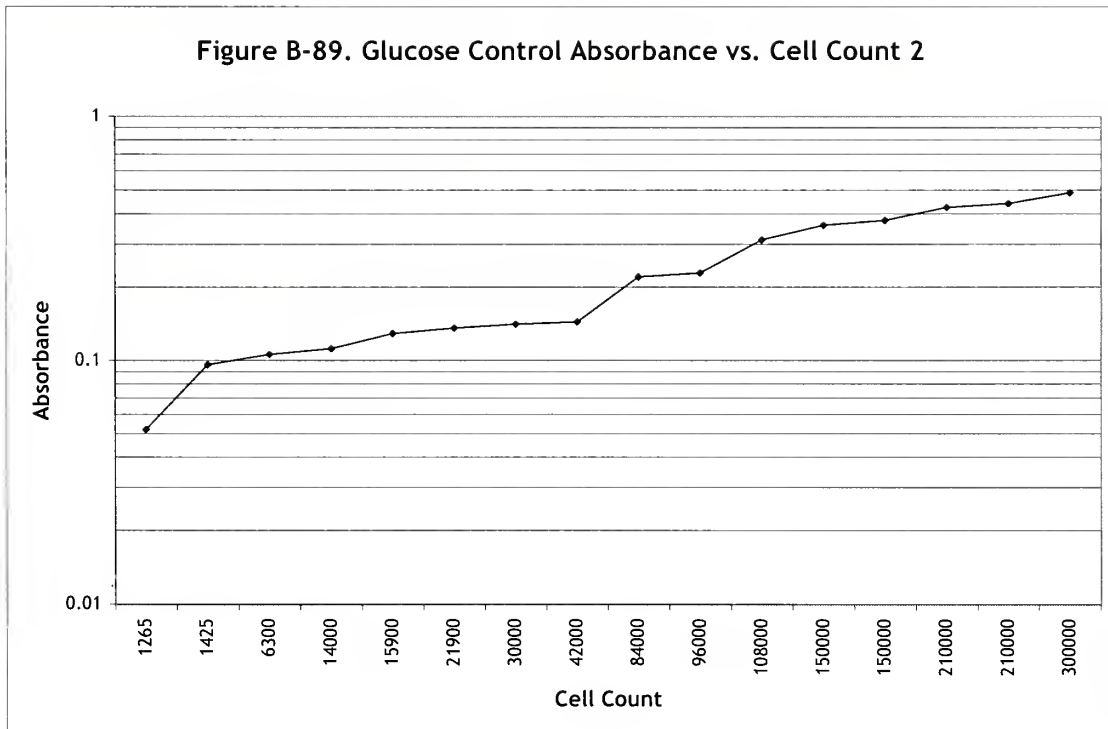
Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1265	0.063	1.16	0.865	1265	0.063
1080	1120	0.099	1.26	0.796	1120	0.099
1260	6090	0.124	1.33	0.752	6090	0.124
1320	13800	0.136	1.37	0.731	13800	0.136
1380	15900	0.146	1.40	0.714	15900	0.146
1500	21900	0.148	1.41	0.711	21900	0.148
1560	30600	0.192	1.56	0.643	30600	0.192
1620	44100	0.226	1.68	0.594	44100	0.226
1680	78900	0.229	1.69	0.590	78900	0.229
1800	85500	0.234	1.71	0.583	85500	0.234
1860	88200	0.256	1.80	0.555	88200	0.256
1920	120000	0.393	2.47	0.405	120000	0.393
1980	132000	0.434	2.72	0.368	132000	0.434
2040	222000	0.462	2.90	0.345	222000	0.462
2100	261000	0.491	3.10	0.323	261000	0.491
2160	399000	0.513	3.26	0.307	399000	0.513



Cell Count increases from 21900 to 44100

Difference in absorbance = (0.226 - 0.148) = 0.078

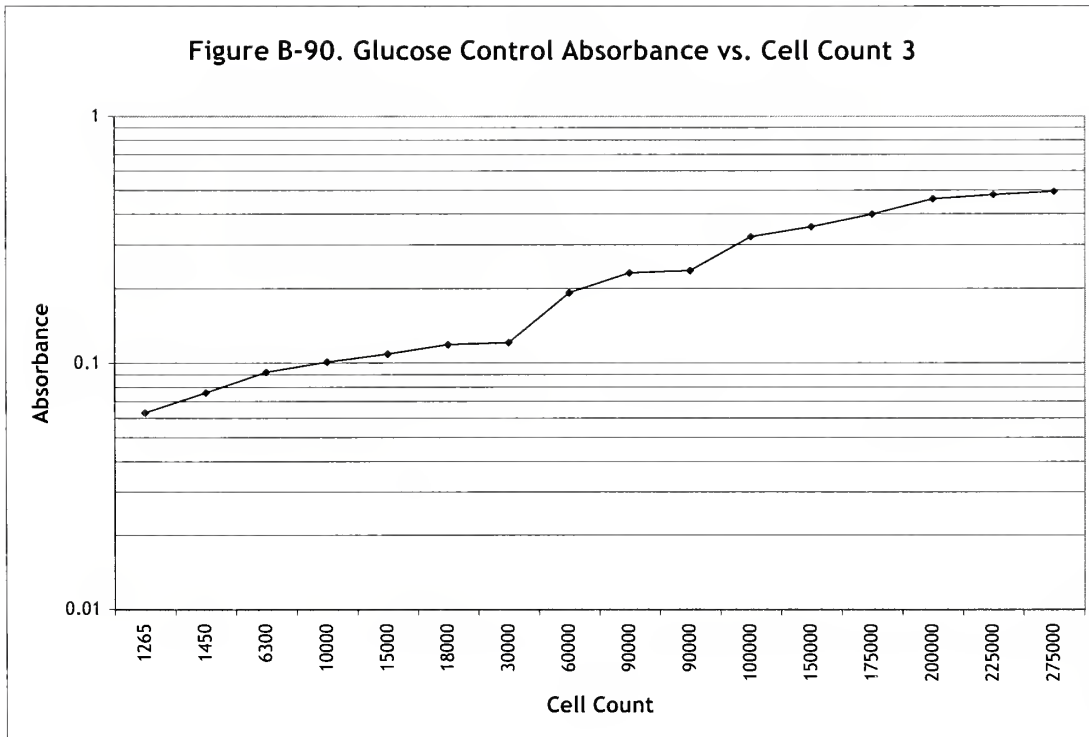
Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1265	0.052	1.13	0.887	1265	0.052
1080	1425	0.096	1.25	0.802	1425	0.096
1260	6300	0.106	1.28	0.783	6300	0.106
1320	14000	0.112	1.29	0.773	14000	0.112
1380	15900	0.129	1.35	0.743	15900	0.129
1500	21900	0.136	1.37	0.731	21900	0.136
1560	30000	0.141	1.38	0.723	30000	0.141
1620	42000	0.144	1.39	0.718	42000	0.144
1680	84000	0.221	1.66	0.601	84000	0.221
1800	96000	0.229	1.69	0.590	96000	0.229
1860	108000	0.313	2.06	0.486	108000	0.313
1920	150000	0.359	2.29	0.438	150000	0.359
1980	150000	0.376	2.38	0.421	150000	0.376
2040	210000	0.425	2.66	0.376	210000	0.425
2100	210000	0.441	2.76	0.362	210000	0.441
2160	300000	0.488	3.08	0.325	300000	0.488



Cell Count increases from 42000 to 84000

Difference in absorbance = (0.221 - 0.144) = 0.077

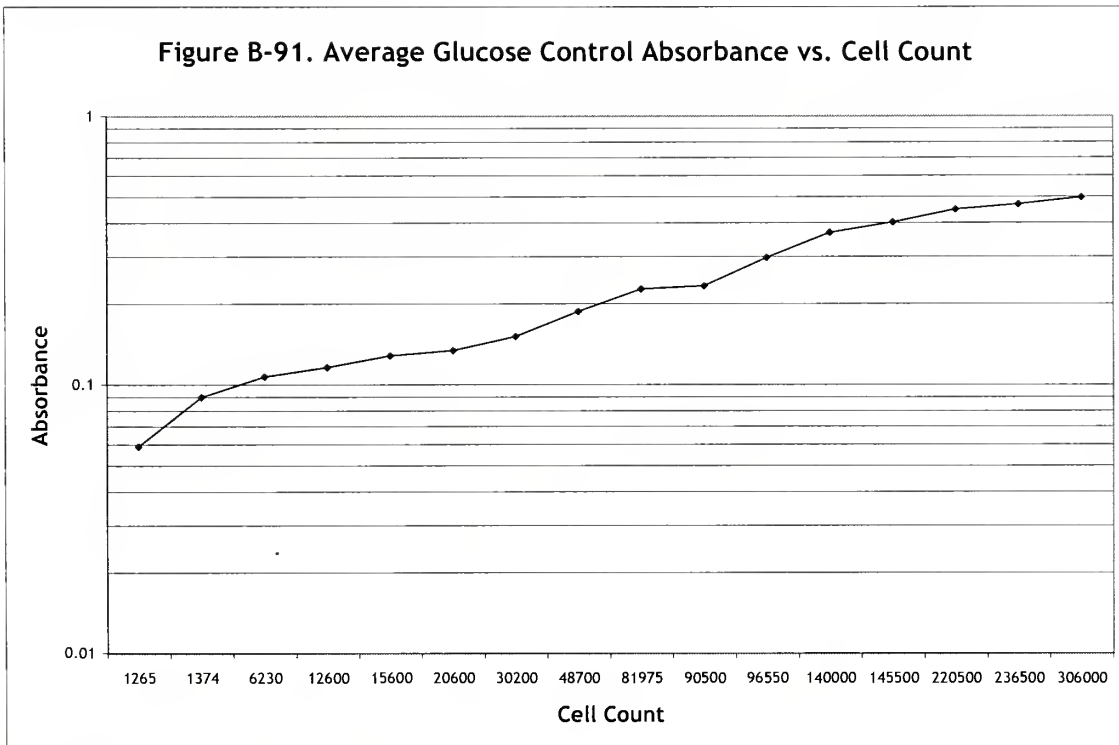
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1265	0.063	1.16	0.865	1265	0.063
1080	1450	0.076	1.19	0.839	1450	0.076
1260	6300	0.092	1.24	0.809	6300	0.092
1320	10000	0.101	1.26	0.793	10000	0.101
1380	15000	0.109	1.29	0.778	15000	0.109
1500	18000	0.119	1.32	0.760	18000	0.119
1560	30000	0.121	1.32	0.757	30000	0.121
1620	60000	0.192	1.56	0.643	60000	0.192
1680	90000	0.231	1.70	0.587	90000	0.231
1800	90000	0.236	1.72	0.581	90000	0.236
1860	100000	0.323	2.10	0.475	100000	0.323
1920	150000	0.355	2.26	0.442	150000	0.355
1980	175000	0.399	2.51	0.399	175000	0.399
2040	200000	0.461	2.89	0.346	200000	0.461
2100	225000	0.479	3.01	0.332	225000	0.479
2160	275000	0.493	3.11	0.321	275000	0.493



Cell Count increases from 30000 to 60000

Difference in absorbance = (0.192 - 0.121) = 0.071

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1265	0.059	1.15	0.872	1265	0.059
1080	1374	0.090	1.23	0.812	1374	0.09
1260	6230	0.107	1.28	0.781	6230	0.107
1320	12600	0.116	1.31	0.765	12600	0.116
1380	15600	0.128	1.34	0.745	15600	0.128
1500	20600	0.134	1.36	0.734	20600	0.134
1560	30200	0.151	1.42	0.706	30200	0.151
1620	48700	0.187	1.54	0.650	48700	0.187
1680	81975	0.227	1.69	0.593	81975	0.227
1800	90500	0.233	1.71	0.585	90500	0.233
1860	96550	0.297	1.98	0.504	96550	0.297
1920	140000	0.369	2.34	0.428	140000	0.369
1980	145500	0.403	2.53	0.395	145500	0.403
2040	220500	0.449	2.81	0.355	220500	0.449
2100	236500	0.470	2.95	0.339	236500	0.47
2160	306000	0.498	3.15	0.318	306000	0.498

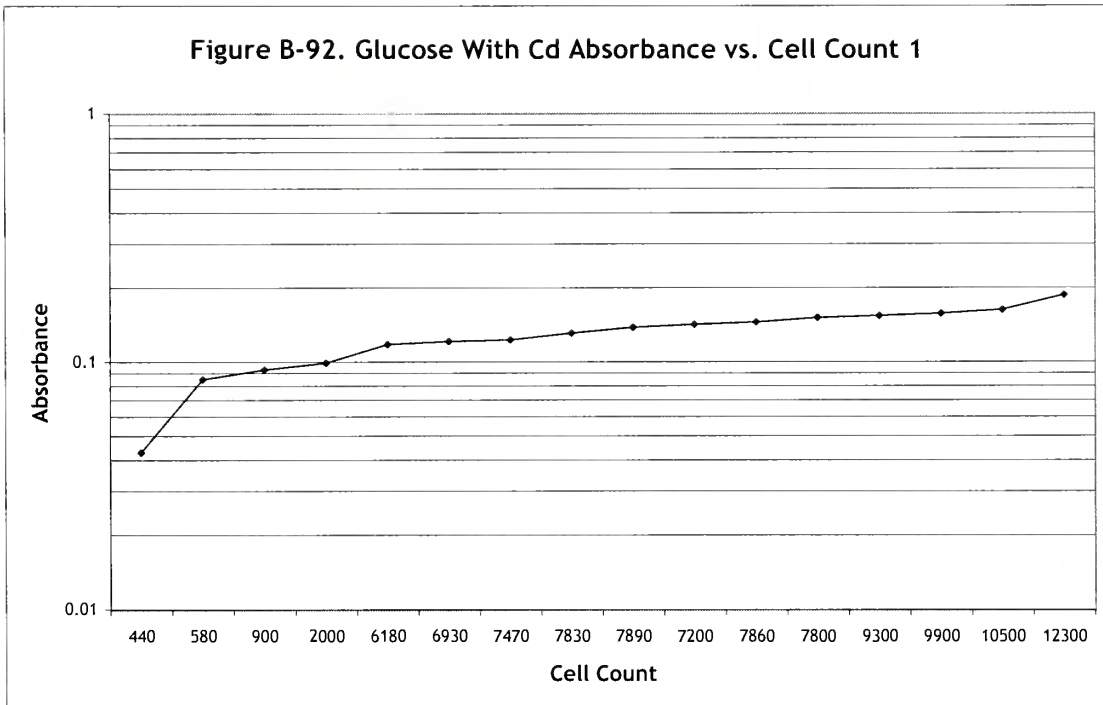


Cell Count increases from 15600 to 30200

Difference in absorbance = (0.151 - 0.128) = 0.023

GLUCOSE WITH Cd CELL COUNT

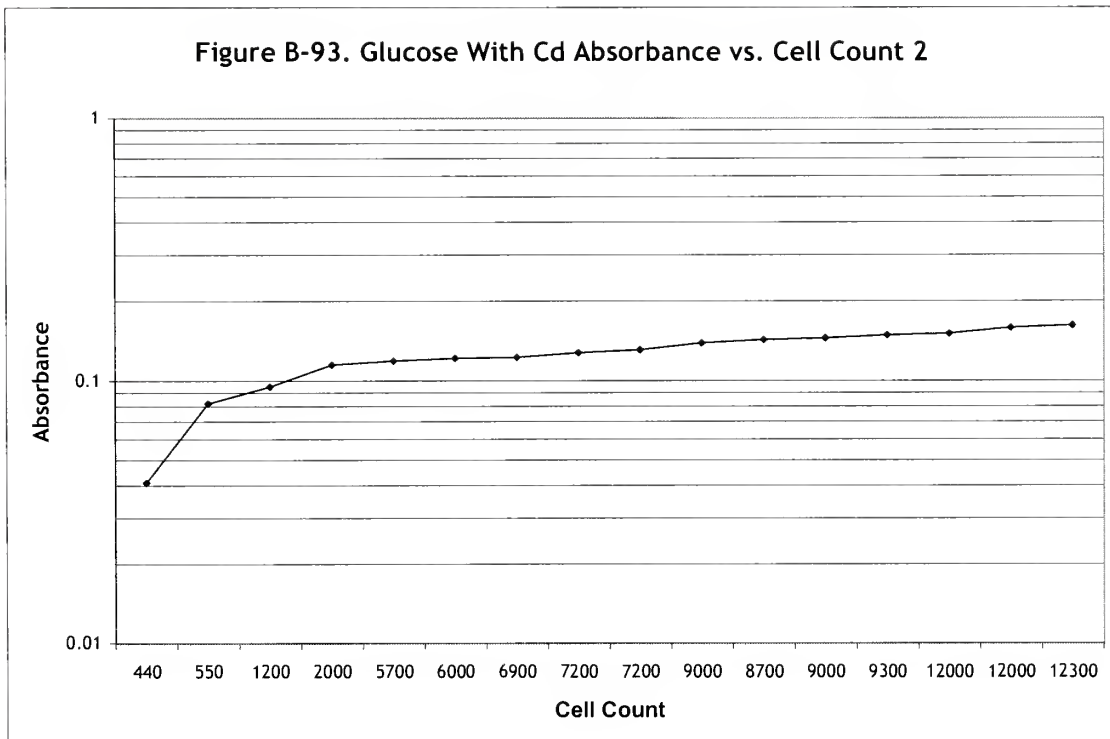
Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	440	0.043	1.10	0.906	440	0.043
1080	580	0.085	1.22	0.822	580	0.085
1260	900	0.093	1.24	0.807	900	0.093
1320	2000	0.099	1.26	0.796	2000	0.099
1380	6180	0.118	1.31	0.762	6180	0.118
1500	6930	0.121	1.32	0.757	6930	0.121
1560	7470	0.123	1.33	0.753	7470	0.123
1620	7830	0.131	1.35	0.740	7830	0.131
1680	7890	0.138	1.37	0.728	7890	0.138
1800	7200	0.142	1.39	0.721	7200	0.142
1860	7860	0.145	1.40	0.716	7860	0.145
1920	7800	0.151	1.42	0.706	7800	0.151
1980	9300	0.154	1.43	0.701	9300	0.154
2040	9900	0.157	1.44	0.697	9900	0.157
2100	10500	0.163	1.46	0.687	10500	0.163
2160	12300	0.187	1.54	0.650	12300	0.187



Cell Count increases from 6180 to 12300

Difference in absorbance = $(0.187 - 0.118) = 0.069$

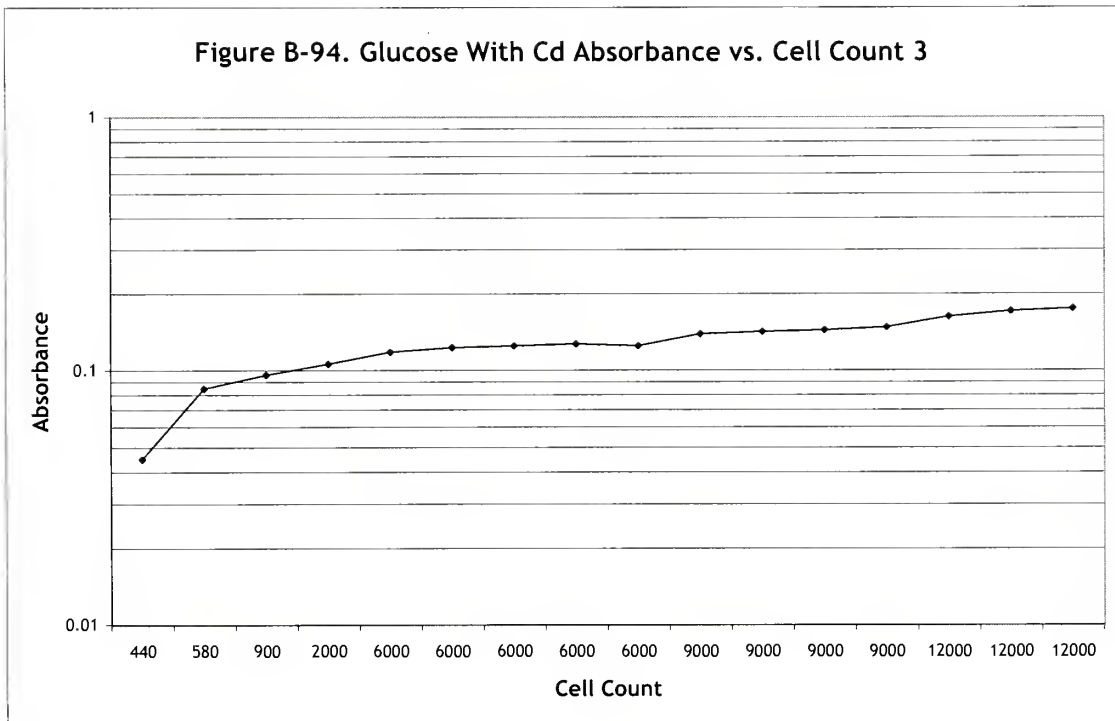
Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	440	0.041	1.10	0.910	440	0.041
1080	550	0.082	1.21	0.828	550	0.082
1260	1200	0.095	1.24	0.804	1200	0.095
1320	2000	0.115	1.30	0.767	2000	0.115
1380	5700	0.119	1.32	0.760	5700	0.119
1500	6000	0.122	1.32	0.755	6000	0.122
1560	6900	0.123	1.33	0.753	6900	0.123
1620	7200	0.128	1.34	0.745	7200	0.128
1680	7200	0.131	1.35	0.740	7200	0.131
1800	9000	0.139	1.38	0.726	9000	0.139
1860	8700	0.143	1.39	0.719	8700	0.143
1920	9000	0.145	1.40	0.716	9000	0.145
1980	9300	0.149	1.41	0.710	9300	0.149
2040	12000	0.162	1.45	0.689	12000	0.151
2100	12000	0.168	1.47	0.679	12000	0.159
2160	12300	0.171	1.48	0.675	12300	0.162



Cell Count increases from 6000 to 12000

Difference in absorbance = (0.162 - 0.122) = 0.04

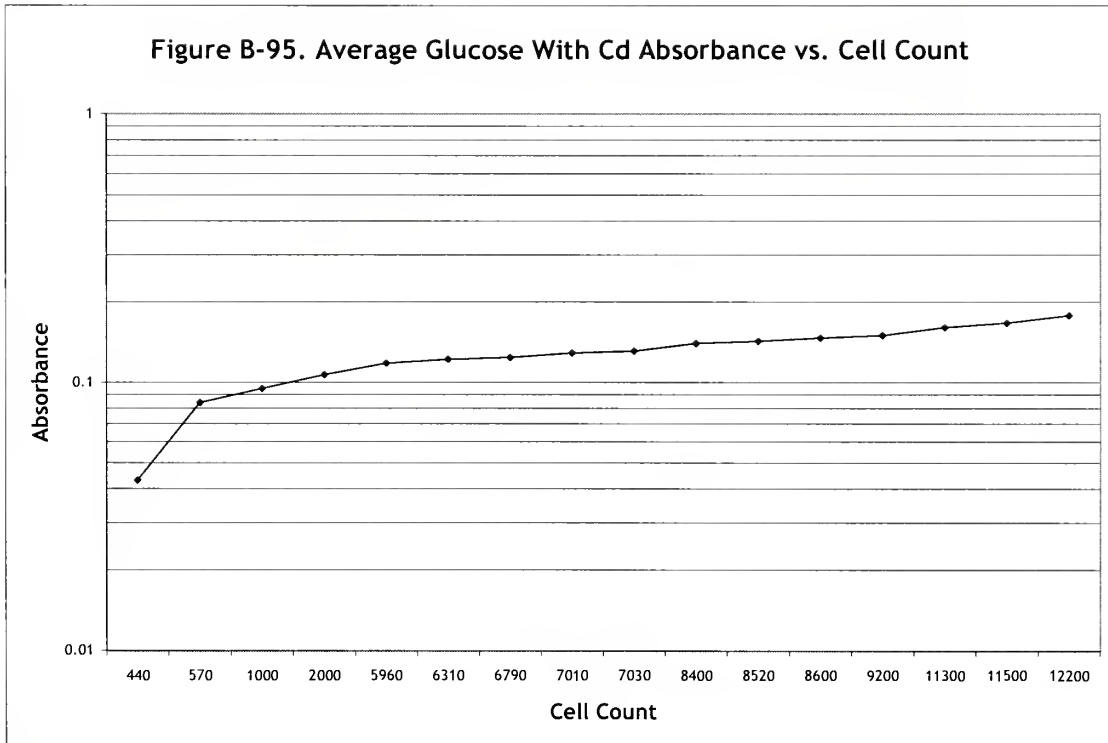
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	440	0.045	1.11	0.902	440	0.045
1080	580	0.085	1.22	0.822	580	0.085
1260	900	0.096	1.25	0.802	900	0.096
1320	2000	0.106	1.28	0.783	2000	0.106
1380	6000	0.118	1.31	0.762	6000	0.118
1500	6000	0.123	1.33	0.753	6000	0.123
1560	6000	0.125	1.33	0.750	6000	0.125
1620	6000	0.127	1.34	0.746	6000	0.127
1680	6000	0.125	1.33	0.750	6000	0.125
1800	9000	0.139	1.38	0.726	9000	0.139
1860	9000	0.142	1.39	0.721	9000	0.142
1920	9000	0.144	1.39	0.718	9000	0.144
1980	9000	0.148	1.41	0.711	9000	0.148
2040	12000	0.163	1.46	0.687	12000	0.163
2100	12000	0.171	1.48	0.675	12000	0.171
2160	12000	0.175	1.50	0.668	12000	0.175



Cell Count increases from 6000 to 12000

Difference in absorbance = (0.163 - 0.127) = 0.038

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	440	0.043	1.10	0.906	440	0.043
1080	570	0.084	1.21	0.824	570	0.084
1260	1000	0.095	1.24	0.804	1000	0.095
1320	2000	0.107	1.28	0.782	2000	0.107
1380	5960	0.118	1.31	0.761	5960	0.118
1500	6310	0.122	1.32	0.755	6310	0.122
1560	6790	0.124	1.33	0.752	6790	0.124
1620	7010	0.129	1.34	0.744	7010	0.129
1680	7030	0.131	1.35	0.739	7030	0.131
1800	8400	0.140	1.38	0.724	8400	0.14
1860	8520	0.143	1.39	0.719	8520	0.143
1920	8600	0.147	1.40	0.713	8600	0.147
1980	9200	0.150	1.41	0.707	9200	0.15
2040	11300	0.161	1.45	0.691	11300	0.161
2100	11500	0.167	1.47	0.680	11500	0.167
2160	12200	0.178	1.51	0.664	12200	0.178

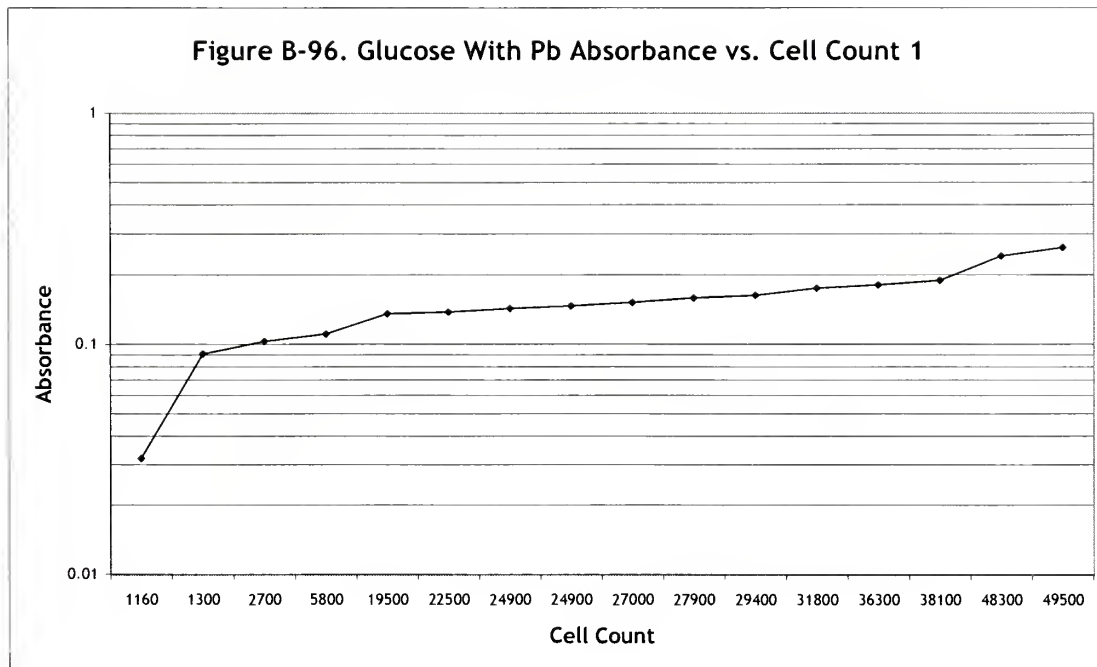


Cell Count increases from 5960 to 11500

Difference in absorbance = (0.161 - 0.118) = 0.043

GLUCOSE WITH Pb CELL COUNT

Time(min)	Cell Count 1	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1160	0.032	1.08	0.929	1160	0.032
1080	1300	0.091	1.23	0.811	1300	0.091
1260	2700	0.103	1.27	0.789	2700	0.103
1320	5800	0.111	1.29	0.774	5800	0.111
1380	19500	0.136	1.37	0.731	19500	0.136
1500	22500	0.138	1.37	0.728	22500	0.138
1560	24900	0.143	1.39	0.719	24900	0.143
1620	24900	0.147	1.40	0.713	24900	0.147
1680	27000	0.152	1.42	0.705	27000	0.152
1800	27900	0.159	1.44	0.693	27900	0.159
1860	29400	0.163	1.46	0.687	29400	0.163
1920	31800	0.175	1.50	0.668	31800	0.175
1980	36300	0.181	1.52	0.659	36300	0.181
2040	38100	0.189	1.55	0.647	38100	0.189
2100	48300	0.241	1.74	0.574	48300	0.241
2160	49500	0.262	1.83	0.547	49500	0.262

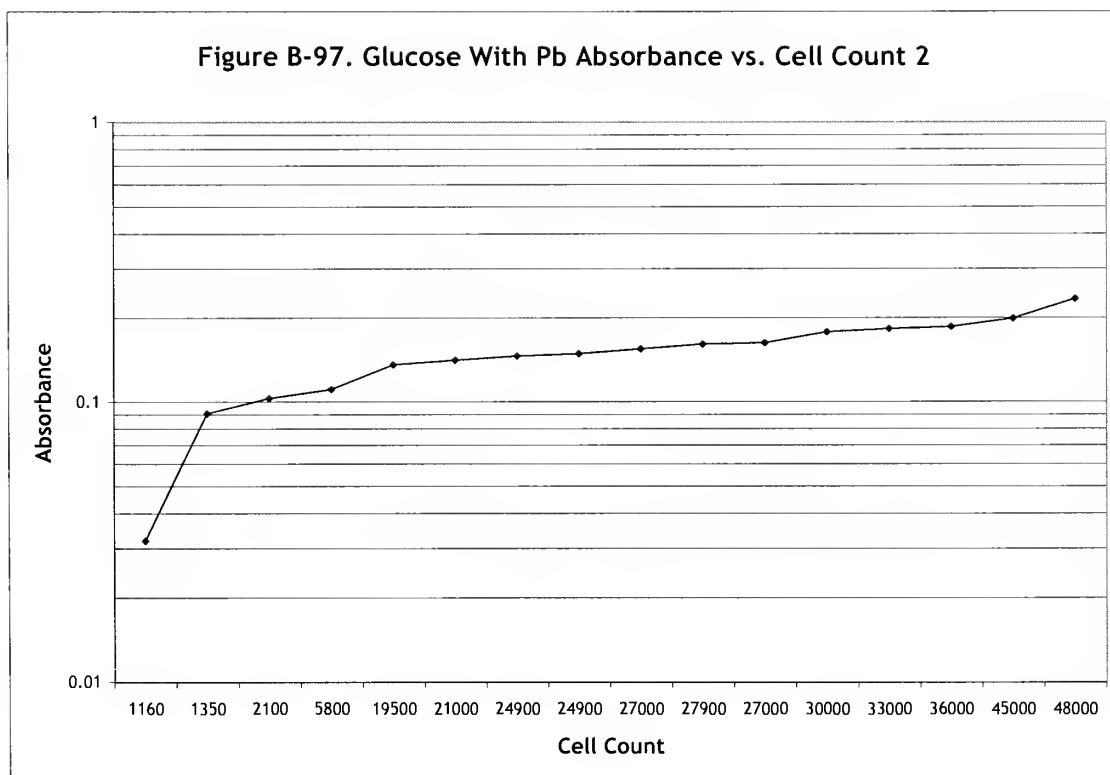


Cell Count increases from 19500 to 38100

Difference in absorbance = (0.169 - 0.136) = 0.053

Time(min)	Cell Count 2	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1160	0.032	1.08	0.929	1160	0.032
1080	1350	0.091	1.23	0.811	1350	0.091
1260	2100	0.103	1.27	0.789	2100	0.103
1320	5800	0.111	1.29	0.774	5800	0.111
1380	19500	0.136	1.37	0.731	19500	0.136
1500	21000	0.141	1.38	0.723	21000	0.141
1560	24900	0.146	1.40	0.714	24900	0.146
1620	24900	0.149	1.41	0.710	24900	0.149
1680	27000	0.155	1.43	0.700	27000	0.155
1800	27900	0.161	1.45	0.690	27900	0.161
1860	27000	0.163	1.46	0.687	27000	0.163
1920	30000	0.178	1.51	0.664	30000	0.178
1980	33000	0.183	1.52	0.656	33000	0.183
2040	36000	0.186	1.53	0.652	36000	0.186
2100	45000	0.199	1.58	0.632	45000	0.199
2160	48000	0.234	1.71	0.583	48000	0.234

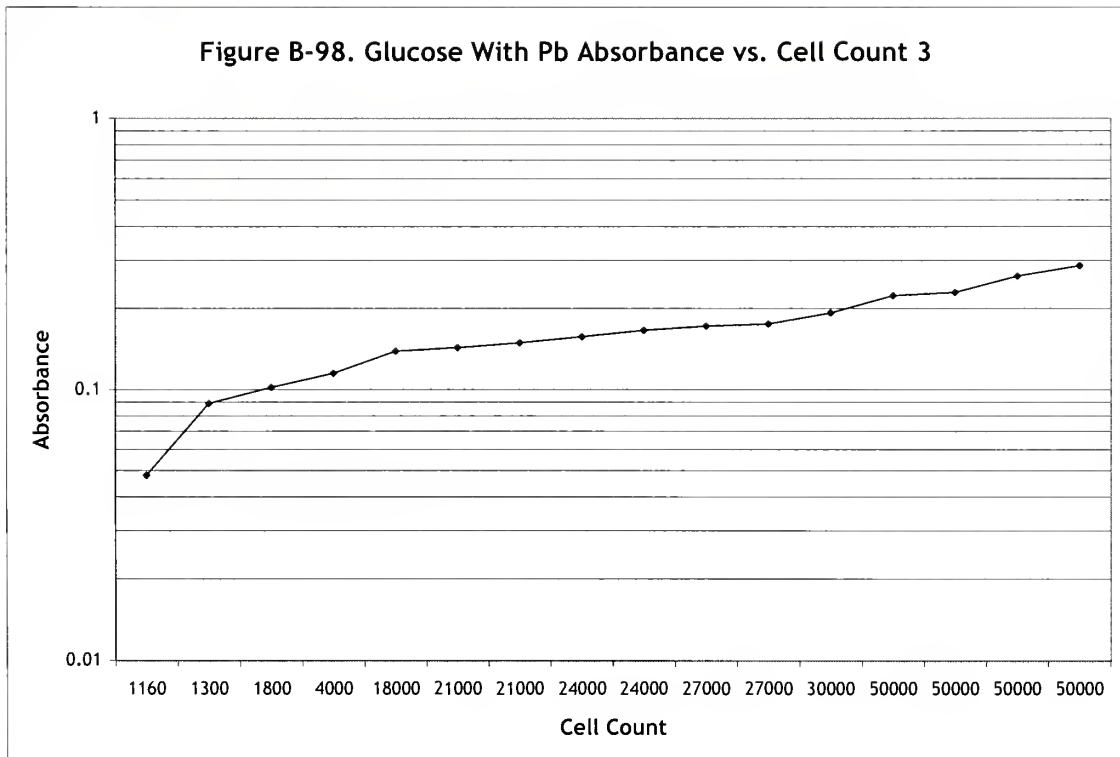
Figure B-97. Glucose With Pb Absorbance vs. Cell Count 2



Cell Count increases from 21000 to 45000

Difference in absorbance = (0.199 - 0.141) = 0.058

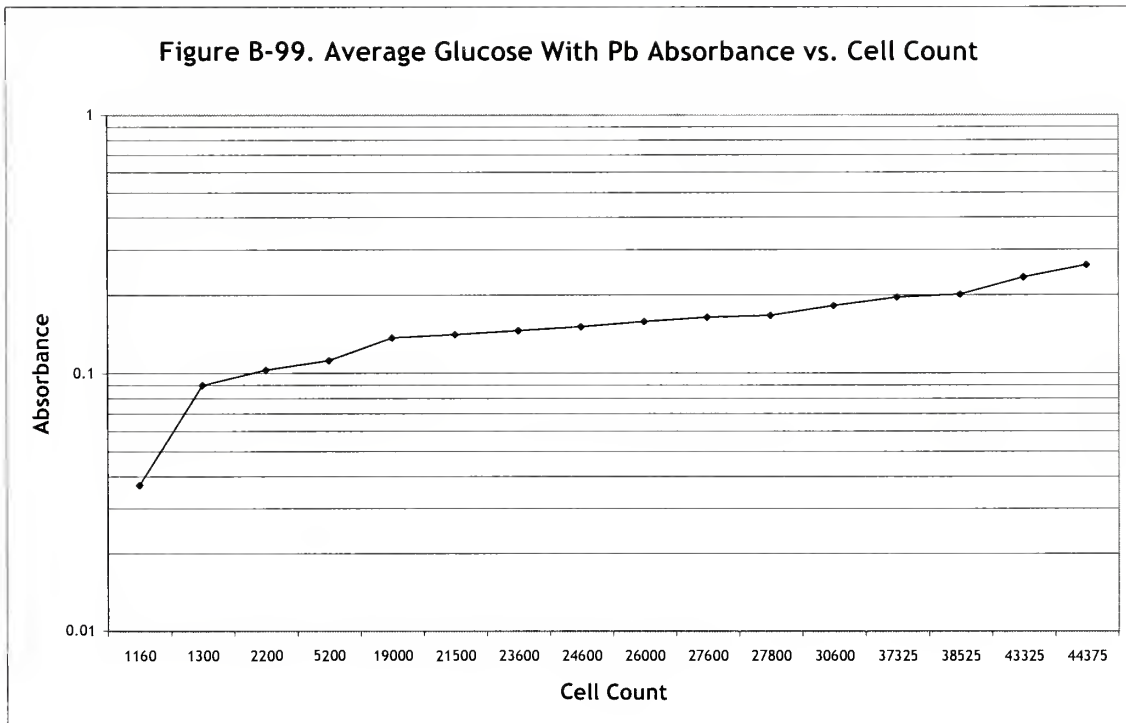
Time(min)	Cell Count 3	Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1160	0.048	1.12	0.895	1160	0.048
1080	1300	0.089	1.23	0.815	1300	0.089
1260	1800	0.102	1.26	0.791	1800	0.102
1320	4000	0.115	1.30	0.767	4000	0.115
1380	18000	0.139	1.38	0.726	18000	0.139
1500	21000	0.143	1.39	0.719	21000	0.143
1560	21000	0.149	1.41	0.710	21000	0.149
1620	24000	0.157	1.44	0.697	24000	0.157
1680	24000	0.166	1.47	0.682	24000	0.166
1800	27000	0.172	1.49	0.673	27000	0.172
1860	27000	0.175	1.50	0.668	27000	0.175
1920	30000	0.192	1.56	0.643	30000	0.192
1980	50000	0.223	1.67	0.598	50000	0.223
2040	50000	0.229	1.69	0.590	50000	0.229
2100	50000	0.263	1.83	0.546	50000	0.263
2160	50000	0.287	1.94	0.516	50000	0.287



Cell Count increases from 24000 to 50000

Difference in absorbance = (0.223 - 0.166) = 0.057

Time(min)	Average Cell Count	Average Absorbance	Antilog	Transmittance	Cell Count	Absorbance
420	1160	0.037	1.09	0.918	1160	0.037
1080	1300	0.090	1.23	0.812	1300	0.09
1260	2200	0.103	1.27	0.789	2200	0.103
1320	5200	0.112	1.30	0.772	5200	0.112
1380	19000	0.137	1.37	0.729	19000	0.137
1500	21500	0.141	1.38	0.723	21500	0.141
1560	23600	0.146	1.40	0.714	23600	0.146
1620	24600	0.151	1.42	0.706	24600	0.151
1680	26000	0.158	1.44	0.696	26000	0.158
1800	27600	0.164	1.46	0.685	27600	0.164
1860	27800	0.167	1.47	0.681	27800	0.167
1920	30600	0.182	1.52	0.658	30600	0.182
1980	37325	0.196	1.57	0.637	37325	0.196
2040	38525	0.201	1.59	0.629	38525	0.201
2100	43325	0.234	1.72	0.583	43325	0.234
2160	44375	0.261	1.82	0.548	44375	0.261



Cell Count increases from 19000 to 38525

Difference in absorbance = (0.201 - 0.137) = 0.064

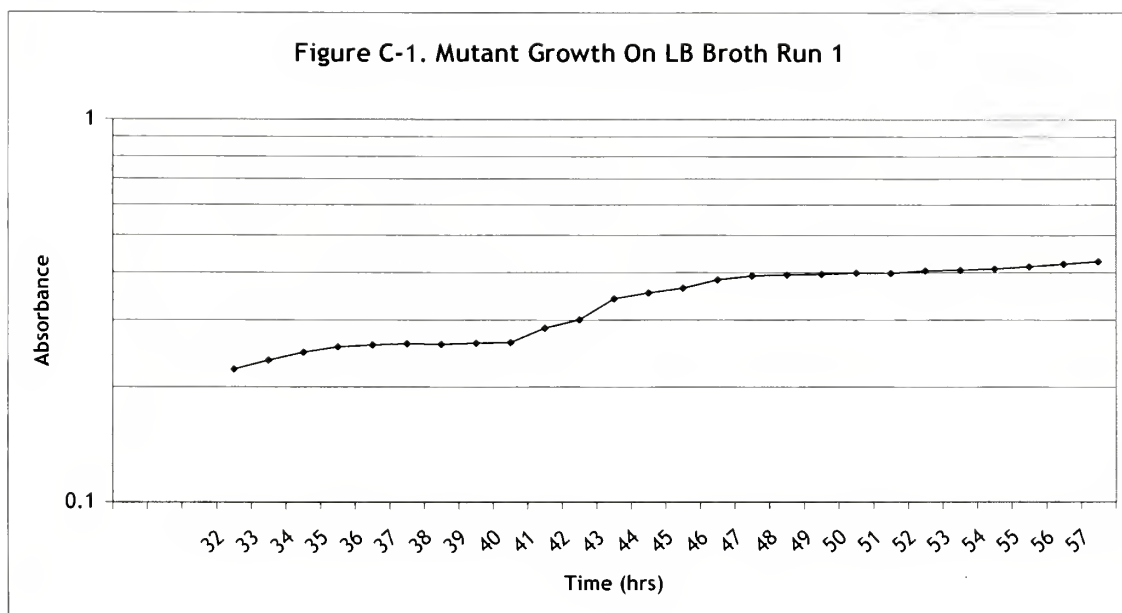
APPENDIX C

All of the doubling times for suspected mutant, donor and recipient strains are presented here

MUTANT growth rates on LB broth

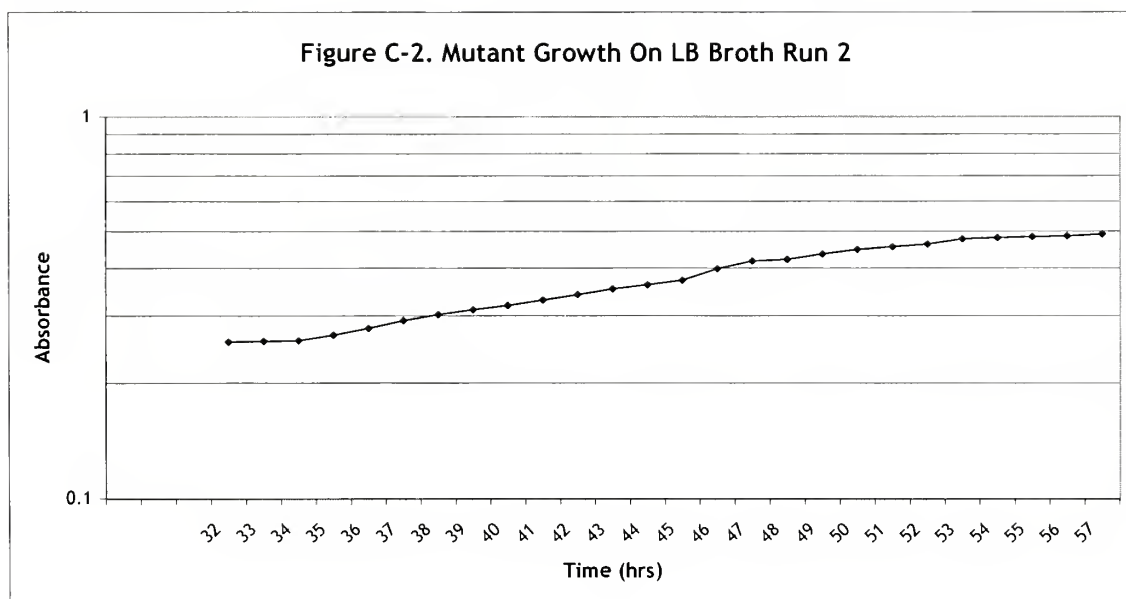
Time(mi)	Time(hrs)	Run1	Run2	Run3	Average
0					
30					
60	1	0.021	0.019	0.018	0.019
120	2	0.032	0.024	0.029	0.028
180	3	0.041	0.038	0.037	0.039
240	4	0.047	0.052	0.045	0.048
300	5	0.054	0.059	0.051	0.055
360	6	0.056	0.063	0.062	0.060
420	7	0.064	0.071	0.068	0.068
480	8	0.075	0.076	0.079	0.077
540	9	0.085	0.079	0.086	0.083
600	10	0.090	0.084	0.097	0.090
660	11	0.091	0.089	0.098	0.093
720	12	0.096	0.102	0.101	0.100
780	13	0.101	0.104	0.105	0.103
840	14	0.107	0.107	0.111	0.108
900	15	0.113	0.109	0.123	0.115
960	16	0.122	0.114	0.132	0.123
1020	17	0.134	0.118	0.139	0.130
1080	18	0.139	0.129	0.147	0.138
1140	19	0.142	0.135	0.153	0.143
1200	20	0.149	0.148	0.159	0.152
1260	21	0.154	0.159	0.163	0.159
1320	22	0.171	0.163	0.168	0.167
1380	23	0.173	0.171	0.185	0.176
1440	24	0.178	0.183	0.189	0.183
1500	25	0.181	0.194	0.193	0.189
1560	26	0.185	0.216	0.212	0.204
1620	27	0.192	0.228	0.228	0.216
1680	28	0.196	0.244	0.231	0.224
1740	29	0.215	0.251	0.236	0.234
1800	30	0.217	0.255	0.244	0.239
1860	31	0.221	0.257	0.253	0.244

1920	32	0.223	0.256	0.255	0.245
1980	33	0.235	0.257	0.259	0.250
2040	34	0.247	0.258	0.261	0.255
2100	35	0.255	0.267	0.265	0.262
2160	36	0.258	0.278	0.269	0.268
2220	37	0.26	0.291	0.272	0.274
2280	38	0.259	0.302	0.278	0.280
2340	39	0.261	0.311	0.283	0.285
2400	40	0.262	0.319	0.299	0.293
2460	41	0.286	0.33	0.312	0.309
2520	42	0.301	0.341	0.314	0.319
2580	43	0.342	0.353	0.321	0.339
2640	44	0.354	0.362	0.331	0.349
2700	45	0.364	0.372	0.338	0.358
2760	46	0.383	0.399	0.34	0.374
2820	47	0.392	0.418	0.341	0.384
2880	48	0.394	0.422	0.352	0.389
2940	49	0.396	0.436	0.361	0.398
3000	50	0.399	0.448	0.375	0.407
3060	51	0.398	0.456	0.389	0.414
3120	52	0.404	0.463	0.396	0.421
3180	53	0.406	0.478	0.404	0.429
3240	54	0.409	0.481	0.412	0.434
3300	55	0.415	0.484	0.423	0.441
3360	56	0.421	0.486	0.436	0.448
3420	57	0.428	0.492	0.448	0.456



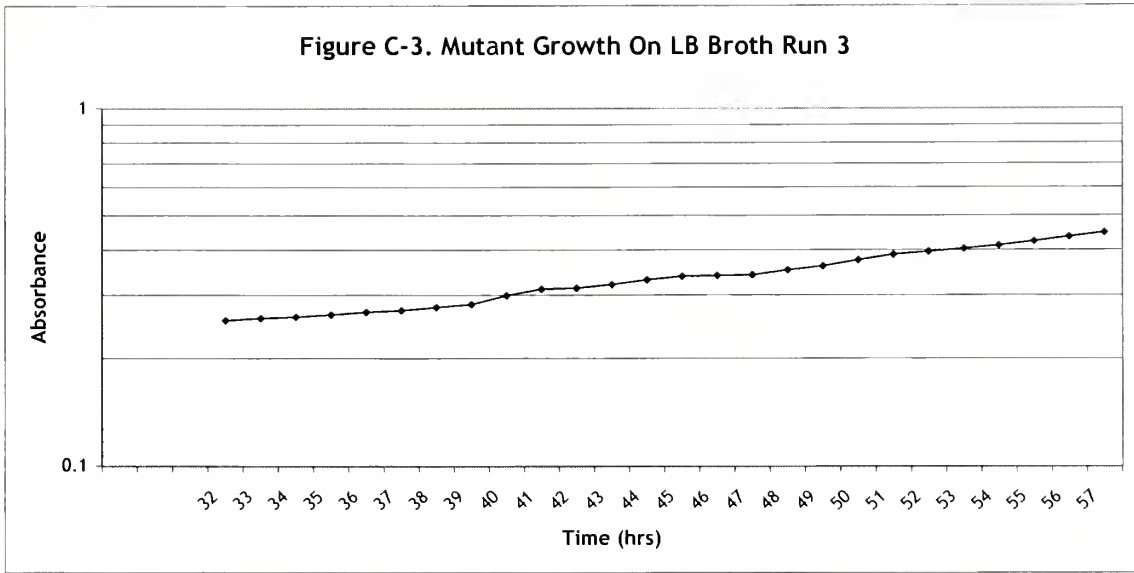
Absorbance increases from 0.107 to 0.215

Actual growth rate = (29 hrs - 14 hrs) = 15 hrs



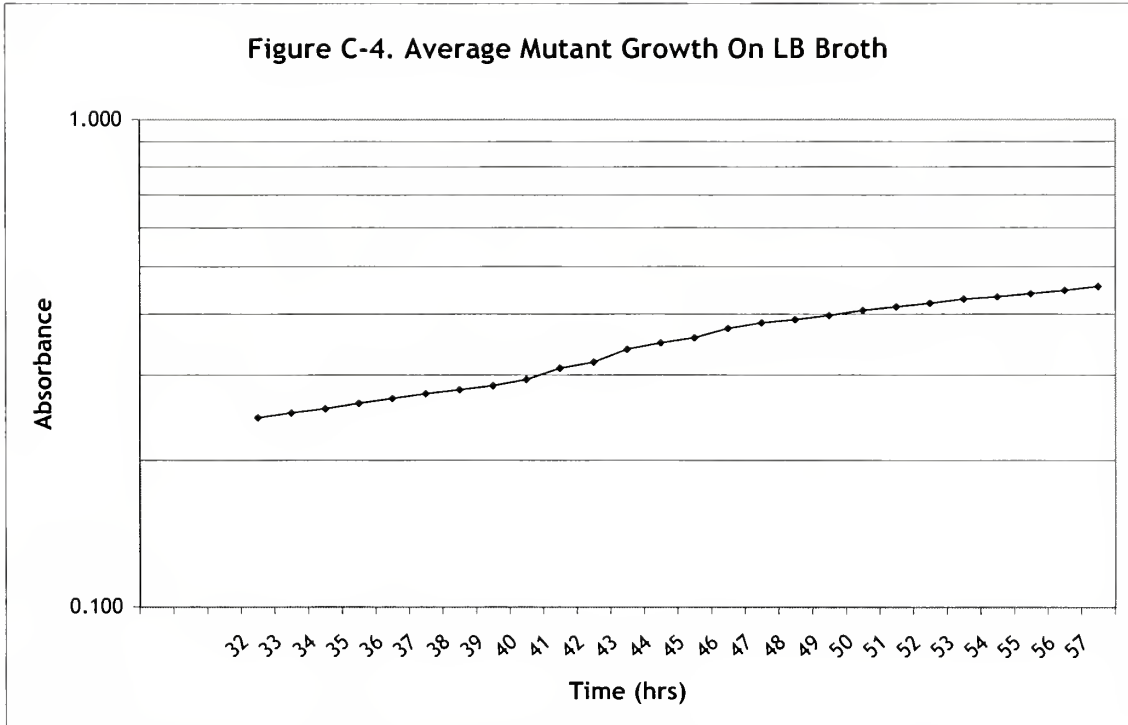
Absorbance increases from 0.109 to 0.216

Actual growth rate = (26 hrs - 15 hrs) = 11 hrs



Absorbance increases from 0.105 to 0.212

Actual growth rate = (26 hrs - 13 hrs) = 13 hrs

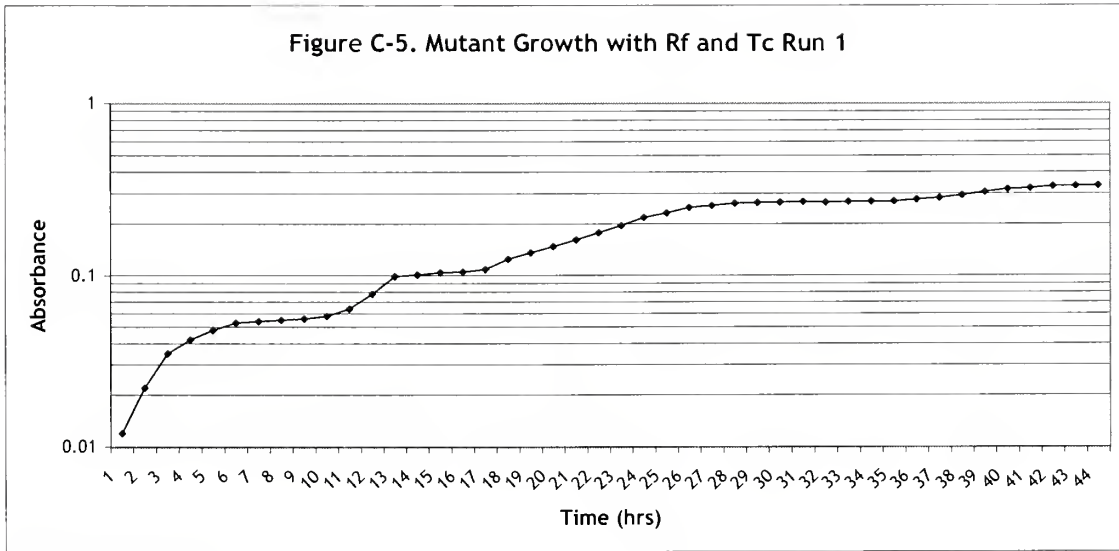


Absorbance increases from 0.108 to 0.216

Actual growth rate = (27 hrs - 14 hrs) = 13 hrs

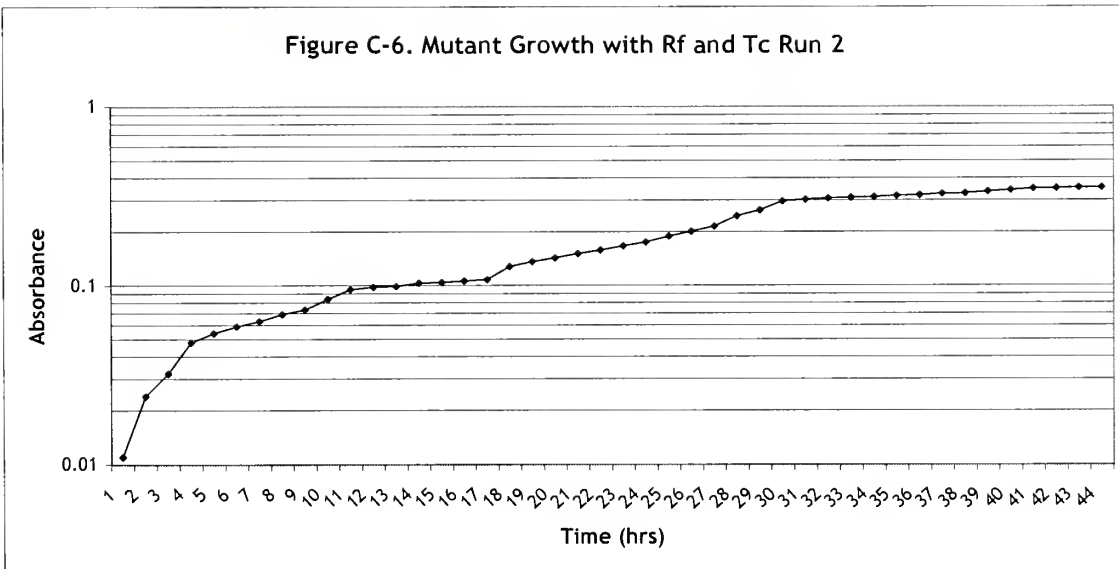
MUTANT growth rates on LB broth with Rif and Tc

Time(min)	Time(hrs)	Run1	Run2	Run3	Average
0					
30					
60	1	0.012	0.011	0.009	0.011
120	2	0.022	0.024	0.018	0.021
180	3	0.035	0.032	0.028	0.032
240	4	0.042	0.048	0.056	0.049
300	5	0.048	0.054	0.063	0.055
360	6	0.053	0.059	0.072	0.061
420	7	0.054	0.063	0.084	0.067
480	8	0.055	0.069	0.090	0.071
540	9	0.056	0.073	0.092	0.074
600	10	0.058	0.084	0.097	0.080
660	11	0.064	0.095	0.102	0.087
720	12	0.078	0.098	0.112	0.096
780	13	0.099	0.099	0.120	0.106
840	14	0.101	0.103	0.129	0.111
900	15	0.104	0.104	0.138	0.115
960	16	0.105	0.106	0.143	0.118
1020	17	0.109	0.108	0.152	0.123
1080	18	0.125	0.128	0.167	0.140
1140	19	0.136	0.136	0.171	0.148
1200	20	0.148	0.143	0.179	0.157
1260	21	0.162	0.151	0.186	0.166
1320	22	0.178	0.158	0.198	0.178
1380	23	0.196	0.167	0.243	0.202
1440	24	0.218	0.175	0.279	0.224
1500	25	0.232	0.189	0.282	0.234
1560	26	0.249	0.201	0.288	0.246
1620	27	0.256	0.215	0.293	0.255
1680	28	0.264	0.245	0.297	0.269
1740	29	0.266	0.264	0.301	0.277
1800	30	0.267	0.297	0.307	0.290
1860	31	0.269	0.303	0.309	0.294
1920	32	0.267	0.308	0.313	0.296
1980	33	0.269	0.311	0.311	0.297
2040	34	0.27	0.313	0.315	0.299
2100	35	0.271	0.318	0.317	0.302
2160	36	0.278	0.321	0.318	0.306
2220	37	0.284	0.326	0.331	0.314
2280	38	0.293	0.328	0.346	0.322
2340	39	0.306	0.336	0.358	0.333
2400	40	0.318	0.342	0.372	0.344
2460	41	0.323	0.349	0.382	0.351
2520	42	0.331	0.351	0.387	0.356
2580	43	0.332	0.353	0.398	0.361
2640	44	0.334	0.354	0.401	0.363



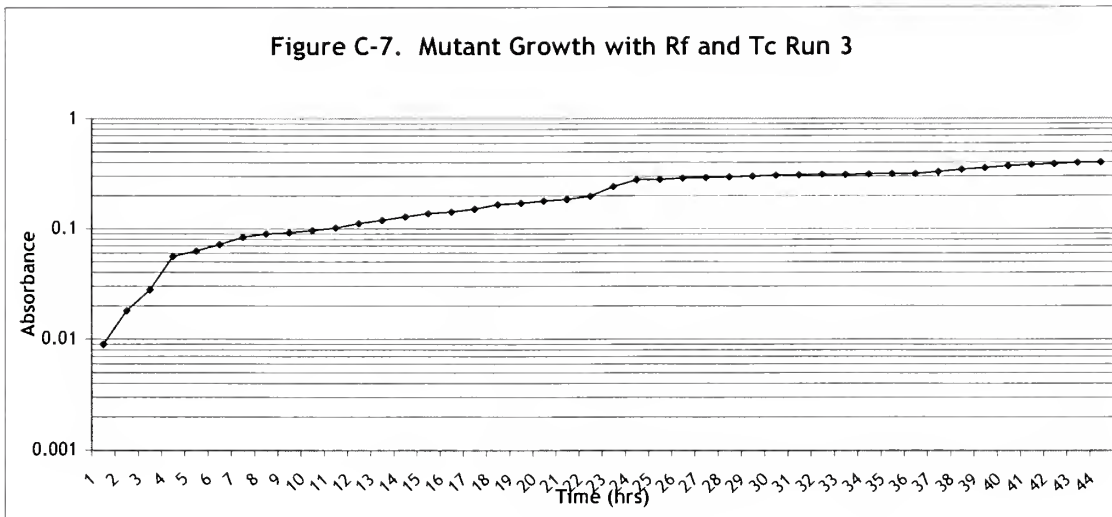
Absorbance increases from 0.109 to 0.218

Actual growth rate = (24 hrs - 17 hrs) = 7 hrs



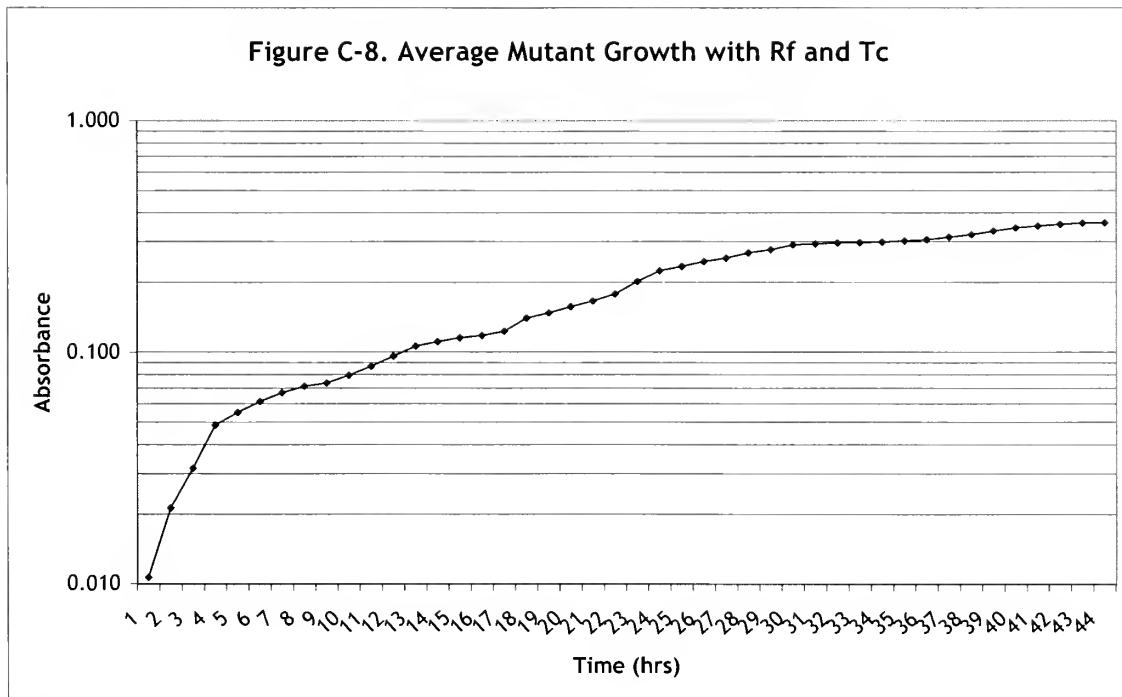
Absorbance increases from 0.108 to 0.215

Actual growth rate = (27 hrs - 17 hrs) = 10 hrs



Absorbance increases from 0.120 to 0.243

Actual growth rate = (23 hrs - 13 hrs) = 10 hrs

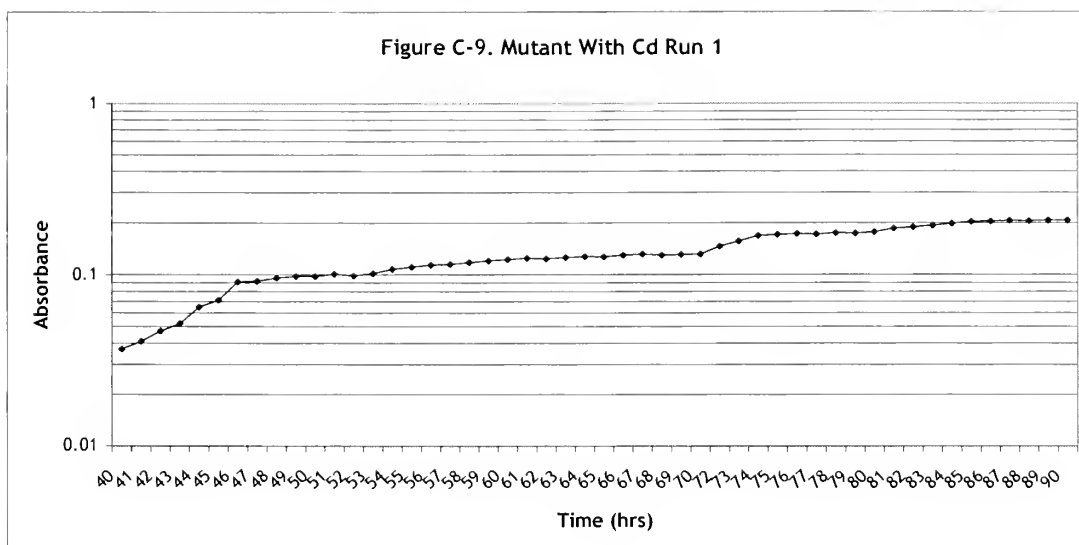


Absorbance increases from 0.111 to 0.224

Actual growth rate = (24 hrs - 14 hrs) = 10 hrs

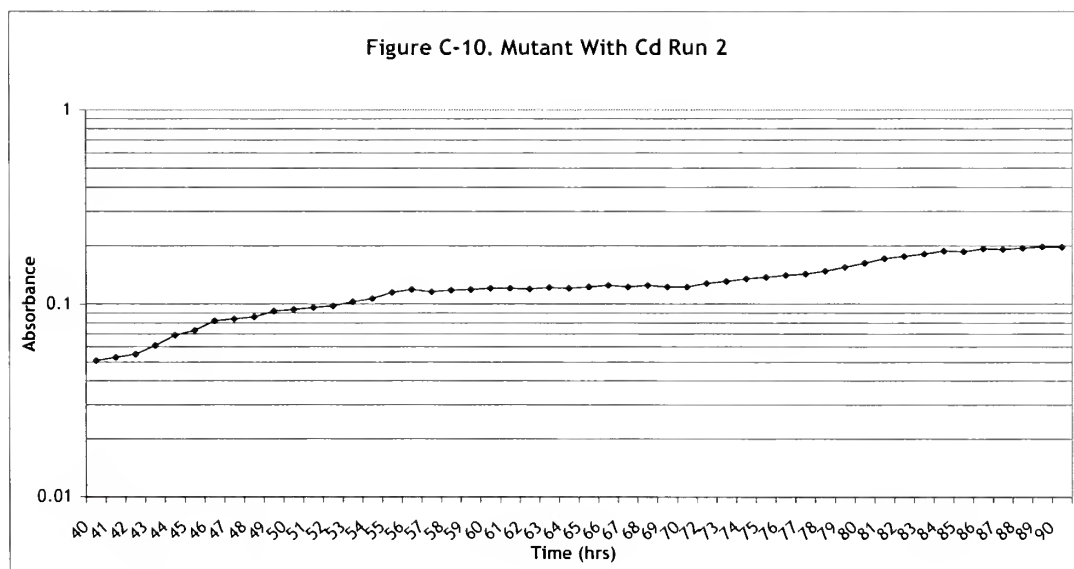
MUTANT growth rates with Cd

50*10 ⁻⁶ M CdCl ₂				
Time(hrs)	Run1	Run2	Run3	Average
40	0.037	0.051	0.052	0.047
41	0.041	0.053	0.06	0.051
42	0.047	0.055	0.062	0.055
43	0.052	0.061	0.069	0.061
44	0.065	0.069	0.075	0.070
45	0.071	0.073	0.077	0.074
46	0.091	0.082	0.079	0.084
47	0.092	0.084	0.083	0.086
48	0.096	0.086	0.088	0.090
49	0.098	0.092	0.086	0.092
50	0.098	0.094	0.091	0.094
51	0.101	0.096	0.095	0.097
52	0.099	0.098	0.096	0.098
53	0.102	0.103	0.098	0.101
54	0.108	0.107	0.101	0.105
55	0.111	0.115	0.104	0.110
56	0.114	0.119	0.106	0.113
57	0.115	0.116	0.11	0.114
58	0.118	0.118	0.115	0.117
59	0.121	0.119	0.117	0.119
60	0.123	0.121	0.123	0.122
61	0.125	0.121	0.124	0.123
62	0.124	0.12	0.126	0.123
63	0.126	0.122	0.129	0.126
64	0.128	0.121	0.128	0.126
65	0.127	0.123	0.134	0.128
66	0.13	0.125	0.136	0.130
67	0.132	0.123	0.139	0.131
68	0.13	0.125	0.141	0.132
69	0.131	0.123	0.143	0.132
70	0.132	0.123	0.144	0.133
72	0.146	0.128	0.152	0.142
73	0.157	0.131	0.158	0.149
74	0.169	0.135	0.163	0.156
75	0.171	0.138	0.165	0.158
76	0.173	0.141	0.168	0.161
77	0.172	0.143	0.173	0.163
78	0.175	0.148	0.177	0.167
79	0.174	0.155	0.179	0.169
80	0.177	0.163	0.183	0.174
81	0.186	0.172	0.186	0.181
82	0.189	0.177	0.189	0.185
83	0.193	0.182	0.188	0.188
84	0.198	0.188	0.194	0.193
85	0.203	0.187	0.198	0.196
86	0.204	0.193	0.206	0.201
87	0.206	0.192	0.209	0.202
88	0.205	0.195	0.211	0.204
89	0.207	0.198	0.213	0.206
90	0.207	0.197	0.215	0.206



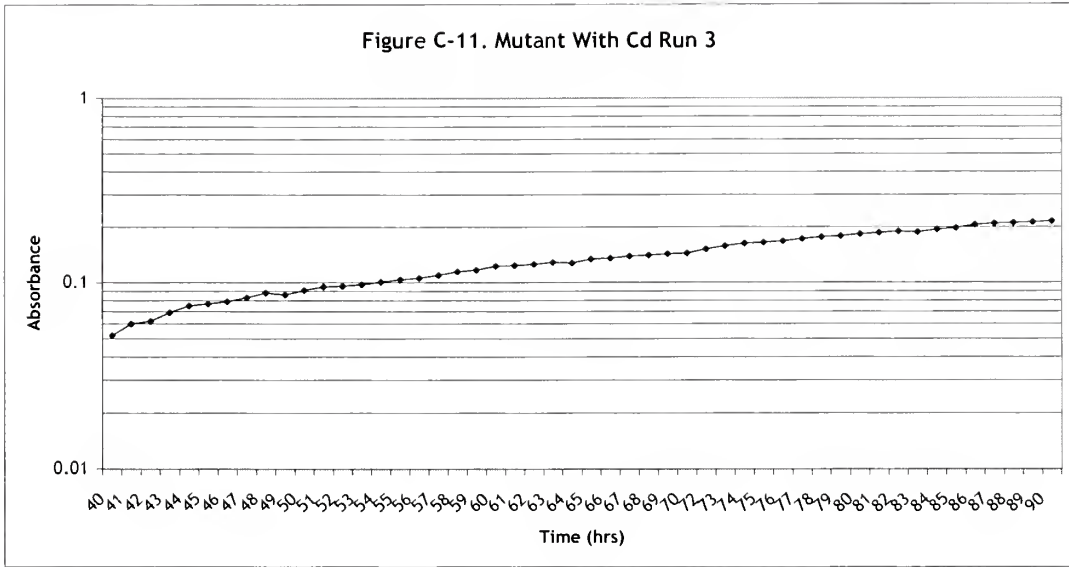
Absorbance increases from 0.102 to 0.204

Actual growth rate = (86 hrs - 53 hrs) = 33 hrs



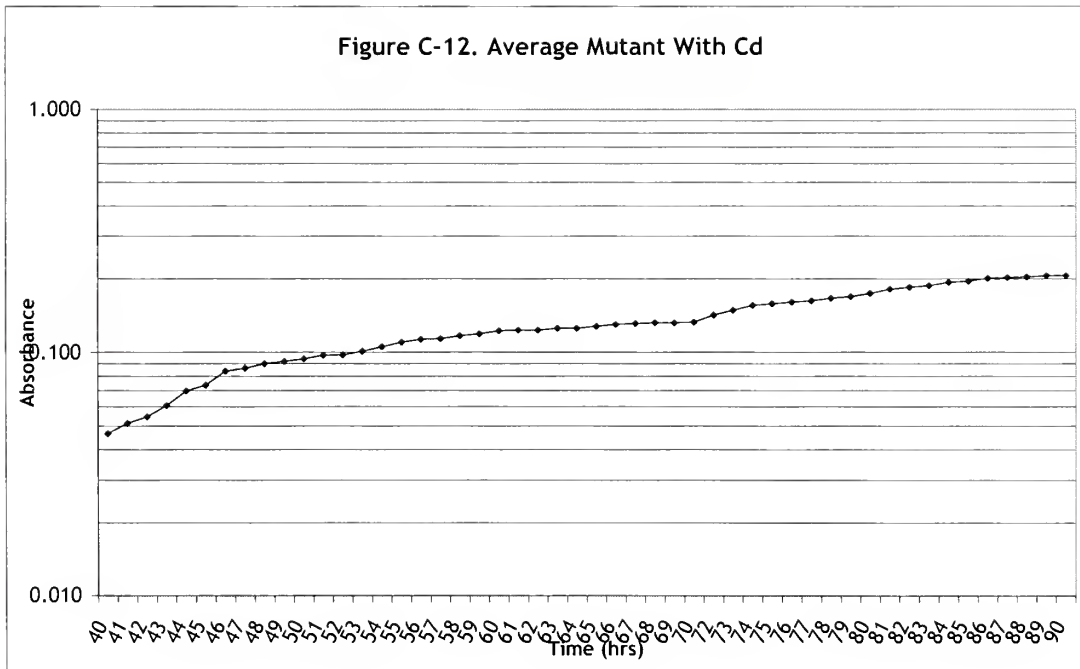
Absorbance increases from 0.096 to 0.192

Actual growth rate = (87 hrs - 51 hrs) = 36 hrs



Absorbance increases from 0.104 to 0.209

Actual growth rate = (87 hrs - 55 hrs) = 32 hrs



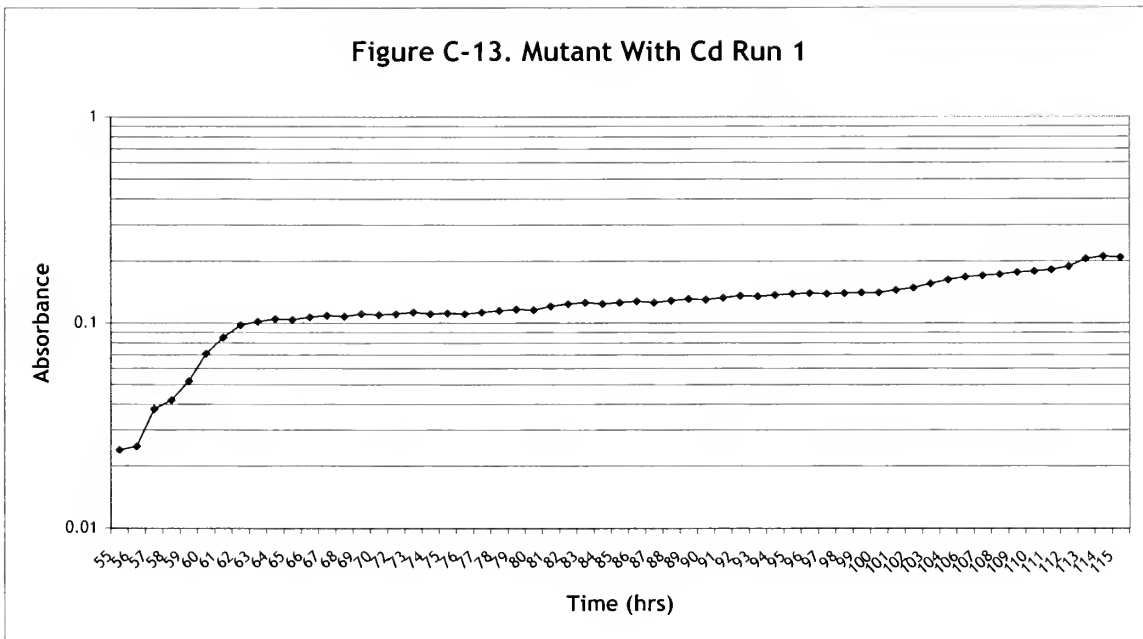
Absorbance increases from 0.101 to 0.202

Actual growth rate = (87 hrs - 53 hrs) = 34 hrs

MUTANT growth rates with Cd

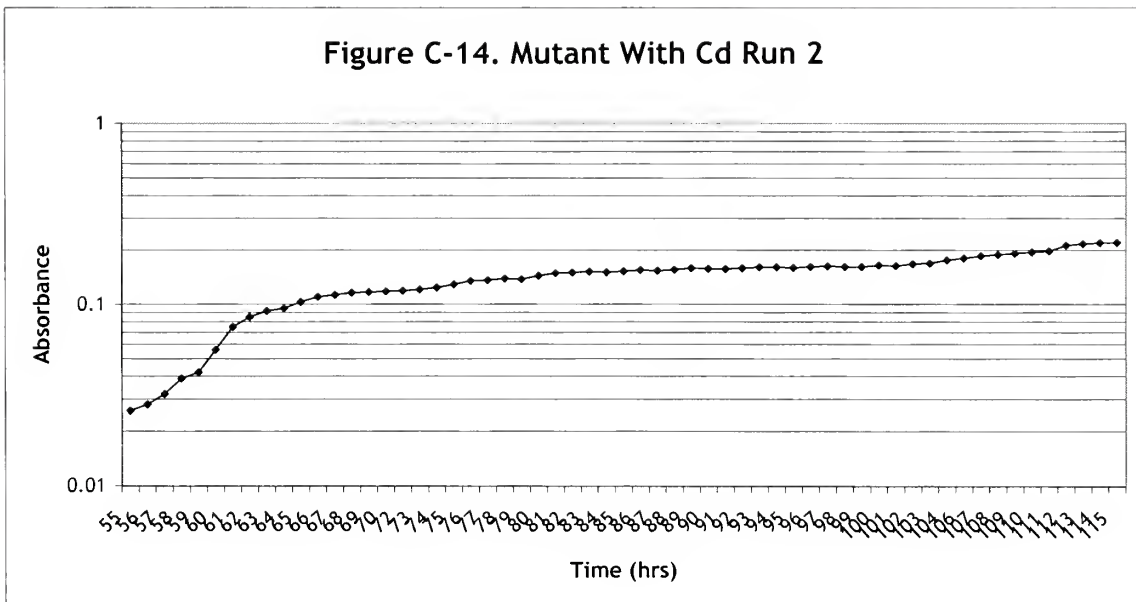
100*10 ⁻⁶ M CdCl ₂				
Time(hrs)	Run1	Run2	Run3	Average
55	0.024	0.026	0.021	0.024
56	0.025	0.028	0.028	0.027
57	0.038	0.032	0.035	0.035
58	0.042	0.039	0.039	0.040
59	0.052	0.042	0.042	0.045
60	0.071	0.056	0.063	0.063
61	0.085	0.075	0.075	0.078
62	0.098	0.085	0.085	0.089
63	0.102	0.092	0.093	0.096
64	0.105	0.095	0.102	0.101
65	0.104	0.103	0.105	0.104
66	0.107	0.11	0.109	0.109
67	0.109	0.113	0.108	0.110
68	0.108	0.116	0.108	0.111
69	0.111	0.117	0.11	0.113
70	0.11	0.118	0.112	0.113
72	0.111	0.119	0.111	0.114
73	0.113	0.121	0.113	0.116
74	0.111	0.124	0.114	0.116
75	0.112	0.129	0.115	0.119
76	0.111	0.135	0.114	0.120
77	0.113	0.136	0.115	0.121
78	0.115	0.139	0.116	0.123
79	0.117	0.138	0.118	0.124
80	0.116	0.144	0.119	0.126
81	0.121	0.149	0.117	0.129
82	0.124	0.15	0.12	0.131
83	0.126	0.152	0.122	0.133
84	0.124	0.151	0.123	0.133
85	0.126	0.153	0.124	0.134
86	0.128	0.155	0.128	0.137
87	0.126	0.154	0.13	0.137
88	0.129	0.156	0.129	0.138
89	0.131	0.159	0.131	0.140
90	0.13	0.158	0.135	0.141
91	0.133	0.157	0.137	0.142
92	0.136	0.159	0.139	0.145
93	0.135	0.161	0.141	0.146

94	0.137	0.161	0.143	0.147
95	0.139	0.16	0.147	0.149
96	0.14	0.162	0.151	0.151
97	0.139	0.163	0.154	0.152
98	0.14	0.162	0.156	0.153
99	0.141	0.162	0.155	0.153
100	0.141	0.165	0.158	0.155
101	0.145	0.164	0.163	0.157
102	0.149	0.168	0.168	0.162
103	0.156	0.169	0.167	0.164
104	0.163	0.176	0.174	0.171
106	0.168	0.181	0.178	0.176
107	0.171	0.186	0.181	0.179
108	0.173	0.189	0.184	0.182
109	0.177	0.192	0.188	0.186
110	0.179	0.195	0.193	0.189
111	0.182	0.198	0.195	0.192
112	0.189	0.213	0.198	0.200
113	0.206	0.217	0.211	0.211
114	0.211	0.22	0.218	0.216
115	0.209	0.221	0.216	0.215



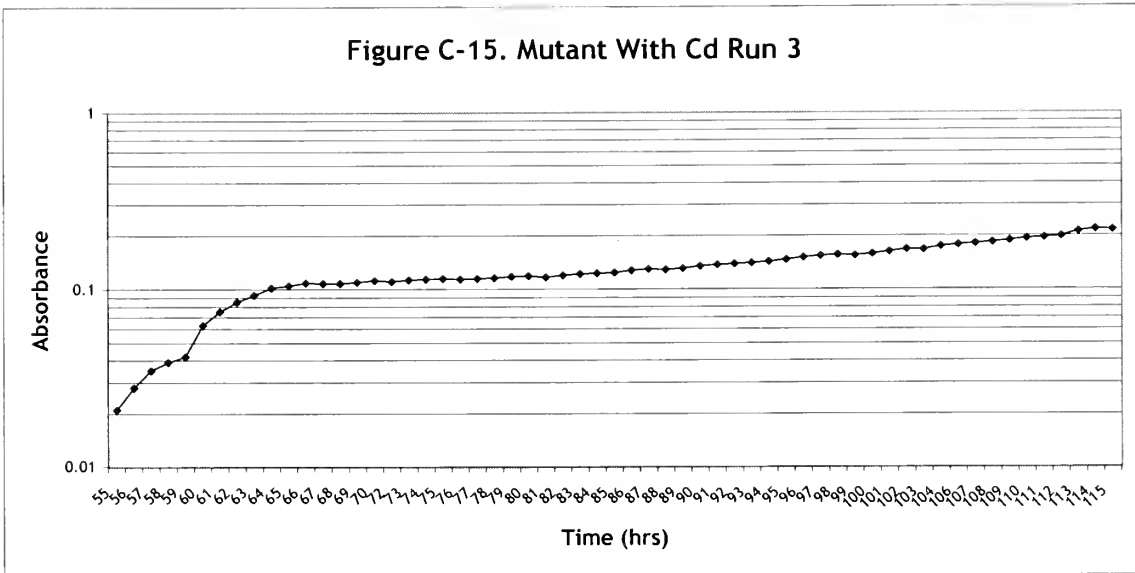
Absorbance increases from 0.105 to 0.211

Actual growth rate = (114 hrs - 64 hrs) = 50 hrs



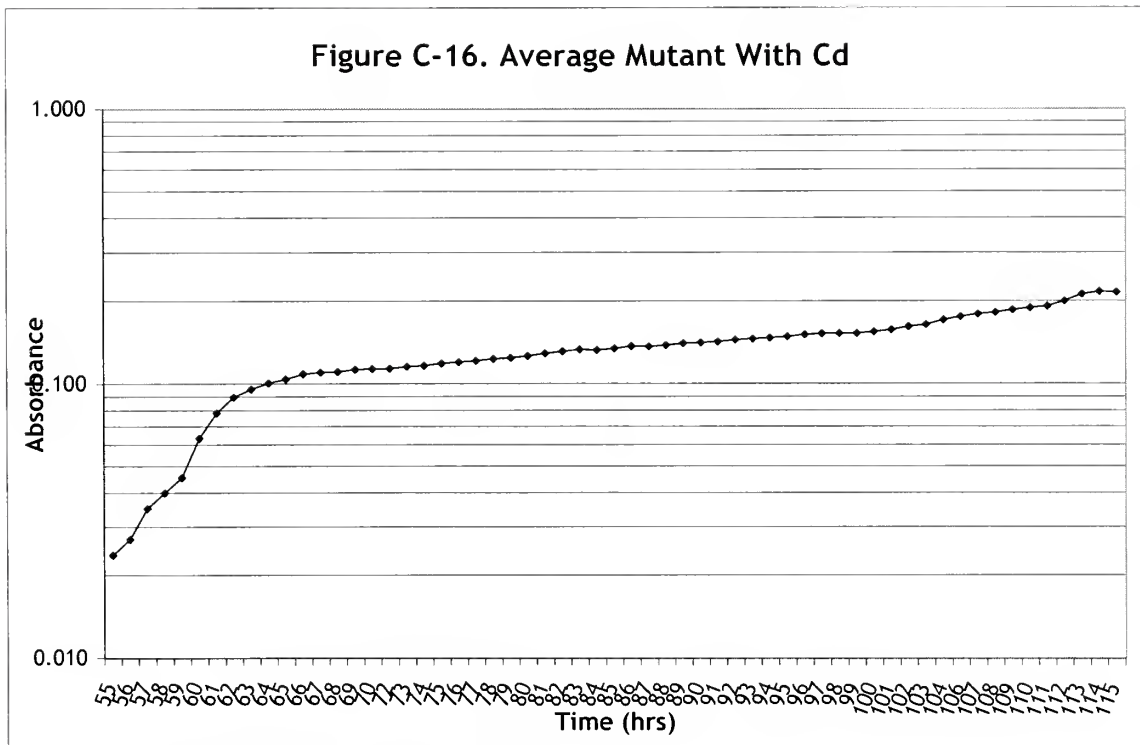
Absorbance increases from 0.111 to 0.221

Actual growth rate = (115 hrs - 66 hrs) = 49 hrs



Absorbance increases from 0.105 to 0.211

Actual growth rate = (113 hrs - 65 hrs) = 48 hrs

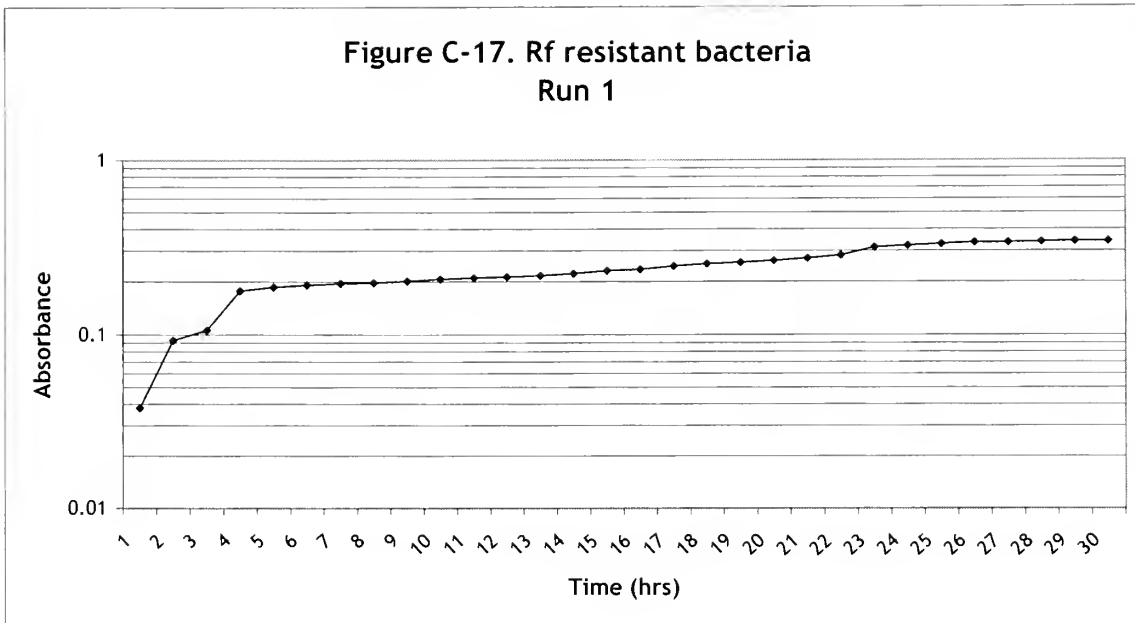


Absorbance increases from 0.109 to 0.216

Actual growth rate = (114 hrs - 66 hrs) = 48 hrs

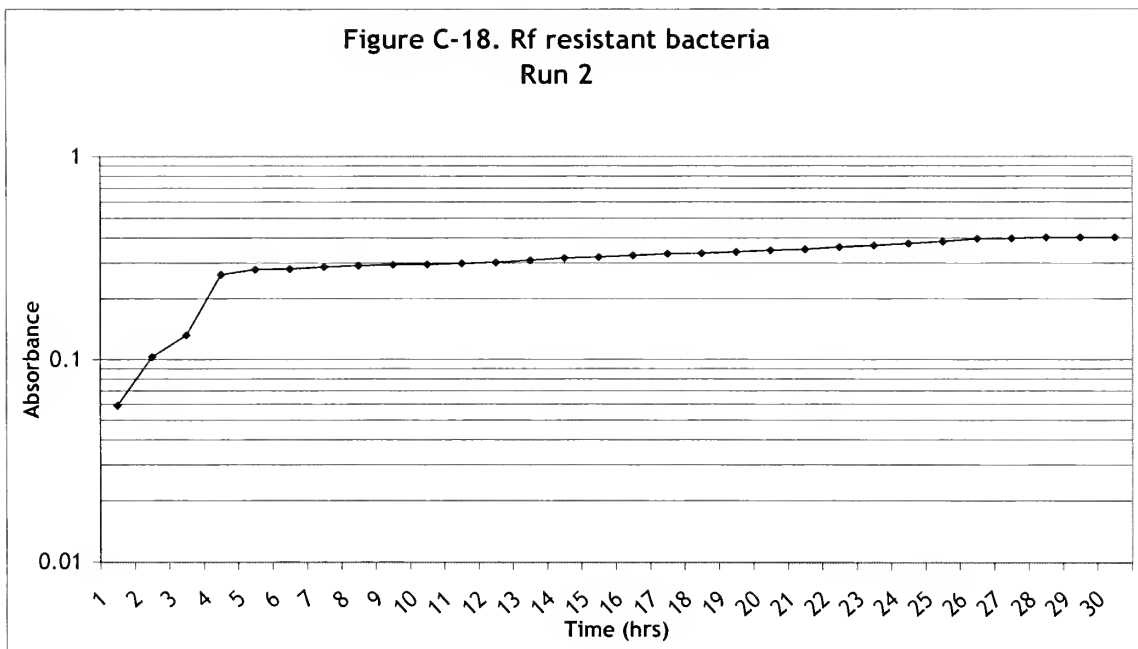
Recipient (Rif resistant *E. coli*) growth rates on LB broth + Rf

Time(min)	Time(hrs)	Run1	Run2	Run3	Average
0					
30					
60	1	0.038	0.059	0.056	0.051
120	2	0.093	0.103	0.099	0.098
180	3	0.106	0.132	0.128	0.122
240	4	0.178	0.263	0.196	0.212
300	5	0.187	0.279	0.236	0.234
360	6	0.192	0.281	0.255	0.243
420	7	0.196	0.288	0.268	0.251
480	8	0.198	0.292	0.275	0.255
540	9	0.202	0.295	0.279	0.259
600	10	0.207	0.296	0.283	0.262
660	11	0.211	0.299	0.291	0.267
720	12	0.214	0.303	0.298	0.272
780	13	0.217	0.311	0.307	0.278
840	14	0.223	0.319	0.314	0.285
900	15	0.232	0.322	0.317	0.290
960	16	0.236	0.328	0.321	0.295
1020	17	0.246	0.334	0.325	0.302
1080	18	0.253	0.337	0.329	0.306
1140	19	0.258	0.341	0.331	0.310
1200	20	0.264	0.348	0.334	0.315
1260	21	0.272	0.352	0.338	0.321
1320	22	0.283	0.361	0.345	0.330
1380	23	0.315	0.367	0.351	0.344
1440	24	0.321	0.376	0.359	0.352
1500	25	0.328	0.384	0.371	0.361
1560	26	0.335	0.396	0.383	0.371
1620	27	0.336	0.398	0.385	0.373
1680	28	0.339	0.402	0.389	0.377
1740	29	0.341	0.403	0.393	0.379
1800	30	0.342	0.402	0.393	0.379



Absorbance increases from 0.093 to 0.187

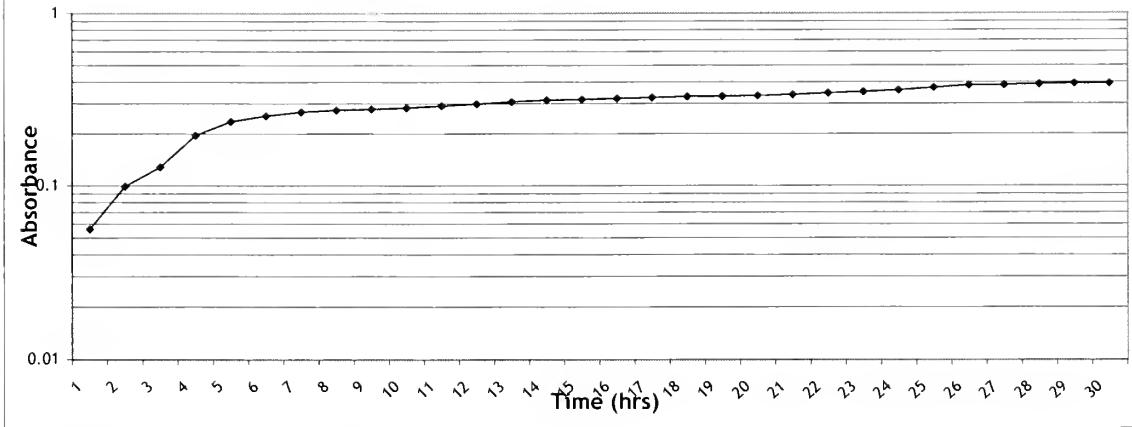
Actual growth rate = $(5 \text{ hrs} - 2 \text{ hrs}) = 3 \text{ hrs}$



Absorbance increases from 0.132 to 0.263

Actual growth rate = $(4 \text{ hrs} - 3 \text{ hrs}) = 1 \text{ hrs}$

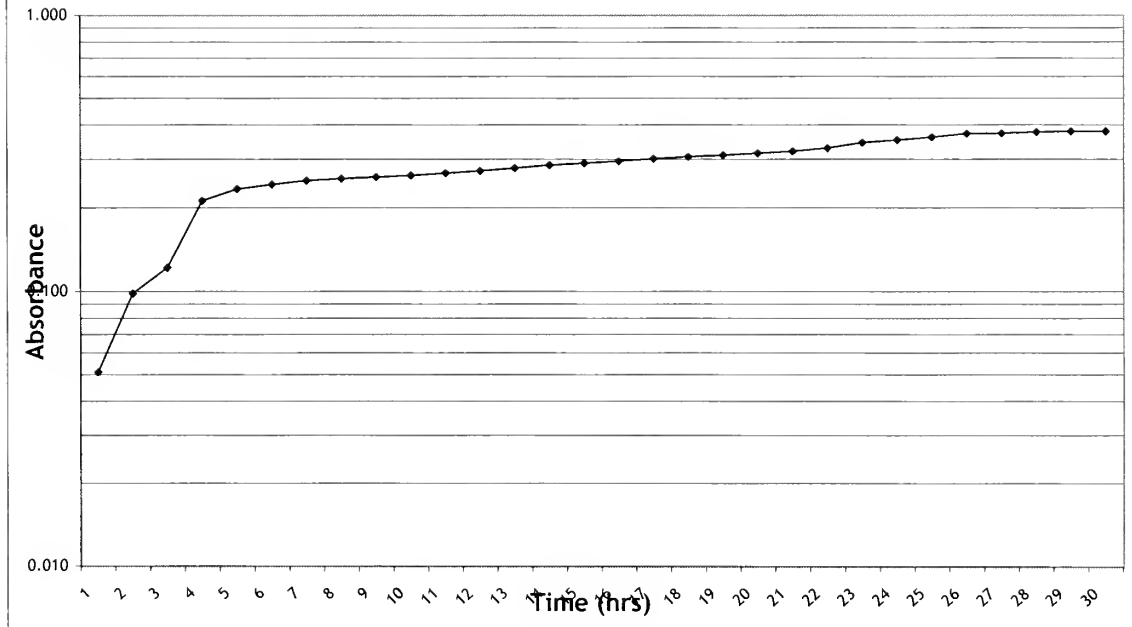
Figure C-19. Rf resistant bacteria
Run 3



Absorbance increases from 0.128 to 0.255

Actual growth rate = (6 hrs - 3 hrs) = 3 hrs

Figure C-20. Rf resistant bacteria
Average run

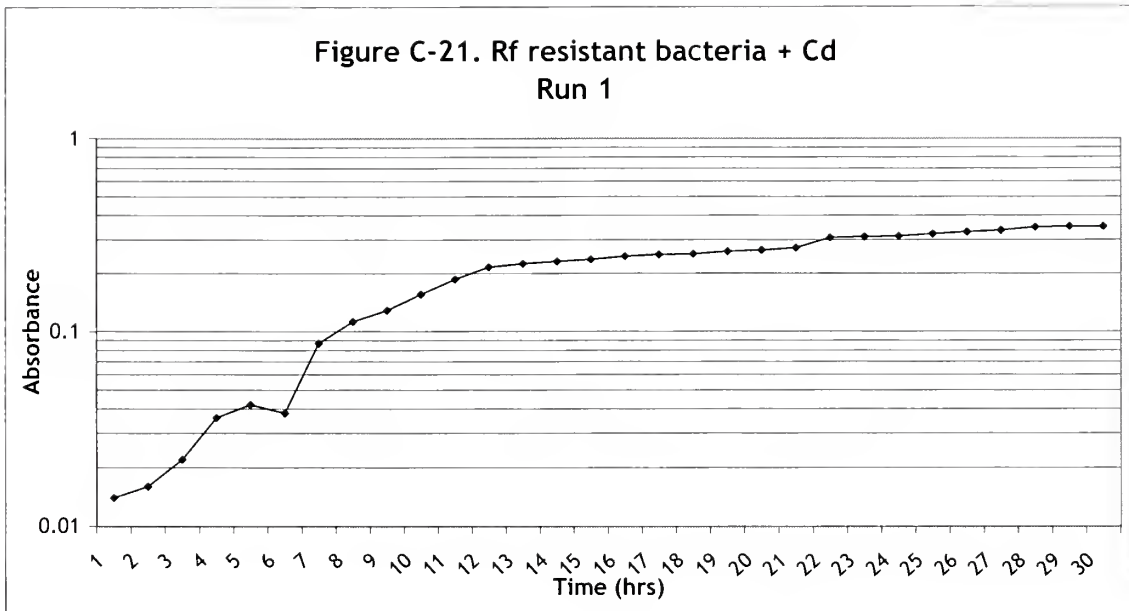


Absorbance increases from 0.122 to 0.243

Actual growth rate = (6 hrs - 3 hrs) = 3 hrs

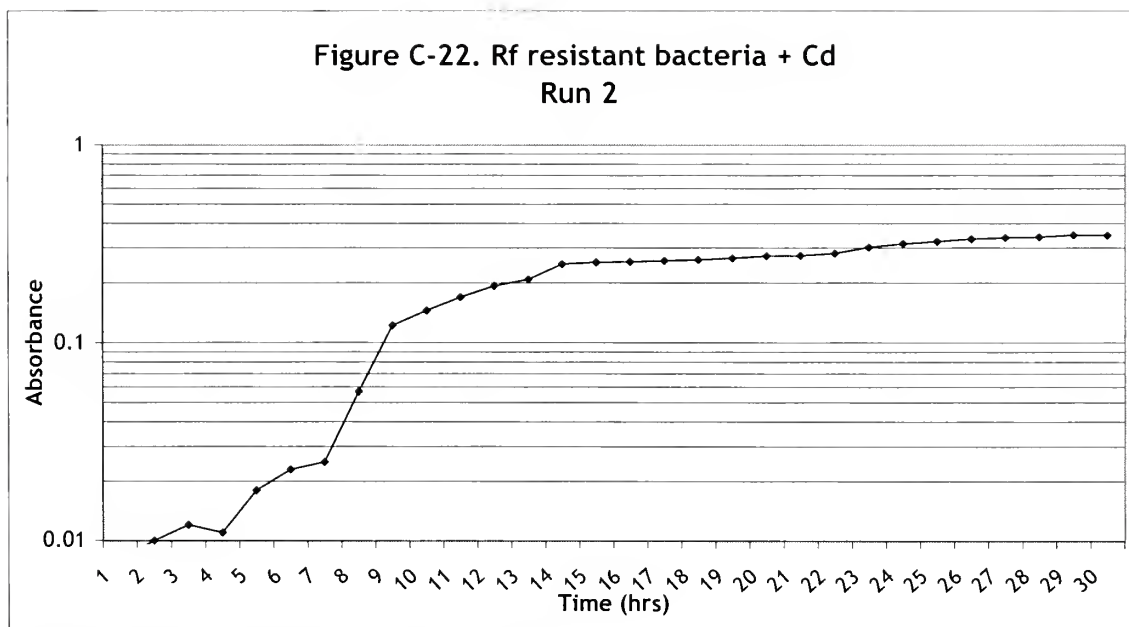
Recipient (Rif resistant *E.coli*) growth rates on LB broth + Rf + Cd

Time(min)	Time(hrs)	Run1	Run2	Run3	Average
0					
30					
60	1	0.014	0.008	0.008	0.010
120	2	0.016	0.01	0.009	0.012
180	3	0.022	0.012	0.014	0.016
240	4	0.036	0.011	0.018	0.022
300	5	0.042	0.018	0.026	0.029
360	6	0.038	0.023	0.059	0.040
420	7	0.087	0.025	0.091	0.068
480	8	0.113	0.057	0.116	0.095
540	9	0.129	0.122	0.121	0.124
600	10	0.156	0.145	0.151	0.151
660	11	0.187	0.169	0.188	0.181
720	12	0.216	0.193	0.192	0.200
780	13	0.226	0.208	0.218	0.217
840	14	0.232	0.249	0.235	0.239
900	15	0.237	0.254	0.246	0.246
960	16	0.246	0.256	0.252	0.251
1020	17	0.251	0.259	0.255	0.255
1080	18	0.253	0.261	0.261	0.258
1140	19	0.261	0.266	0.269	0.265
1200	20	0.265	0.273	0.282	0.273
1260	21	0.272	0.275	0.287	0.278
1320	22	0.308	0.282	0.296	0.295
1380	23	0.311	0.303	0.312	0.309
1440	24	0.313	0.315	0.325	0.318
1500	25	0.322	0.324	0.332	0.326
1560	26	0.329	0.334	0.338	0.334
1620	27	0.336	0.338	0.348	0.341
1680	28	0.349	0.341	0.351	0.347
1740	29	0.352	0.349	0.349	0.350
1800	30	0.351	0.347	0.353	0.350



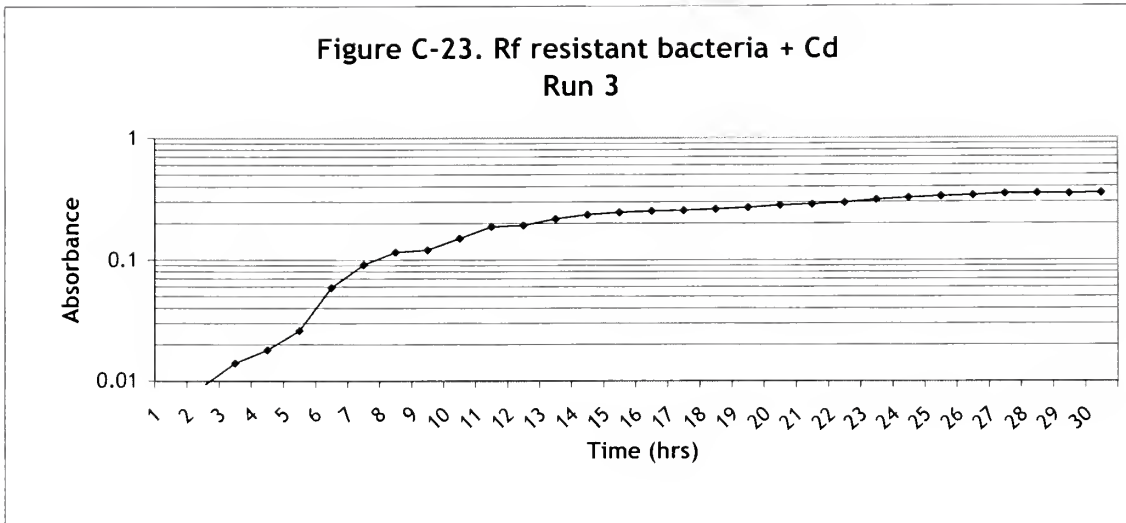
Absorbance increases from 0.113 to 0.226

Actual growth rate = (13 hrs - 8 hrs) = 5 hrs



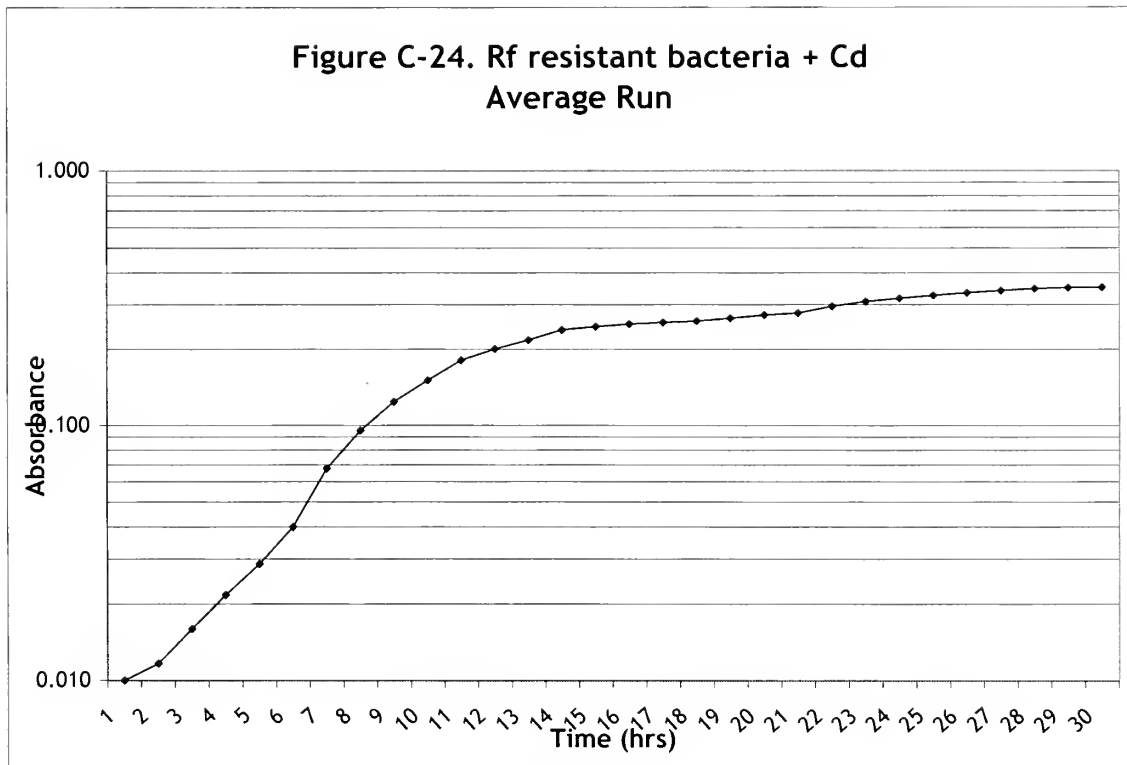
Absorbance increases from 0.122 to 0.249

Actual growth rate = (14 hrs - 9 hrs) = 5 hrs



Absorbance increases from 0.116 to 0.235

Actual growth rate = (14 hrs - 8 hrs) = 6 hrs

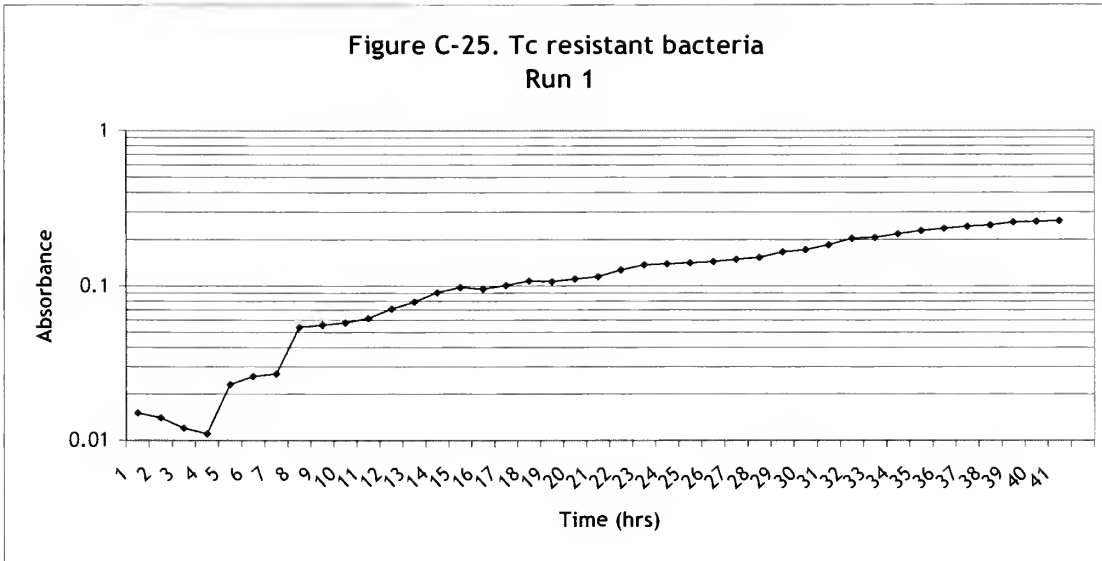


Absorbance increases from 0.124 to 0.246

Actual growth rate = (15 hrs - 9 hrs) = 6 hrs

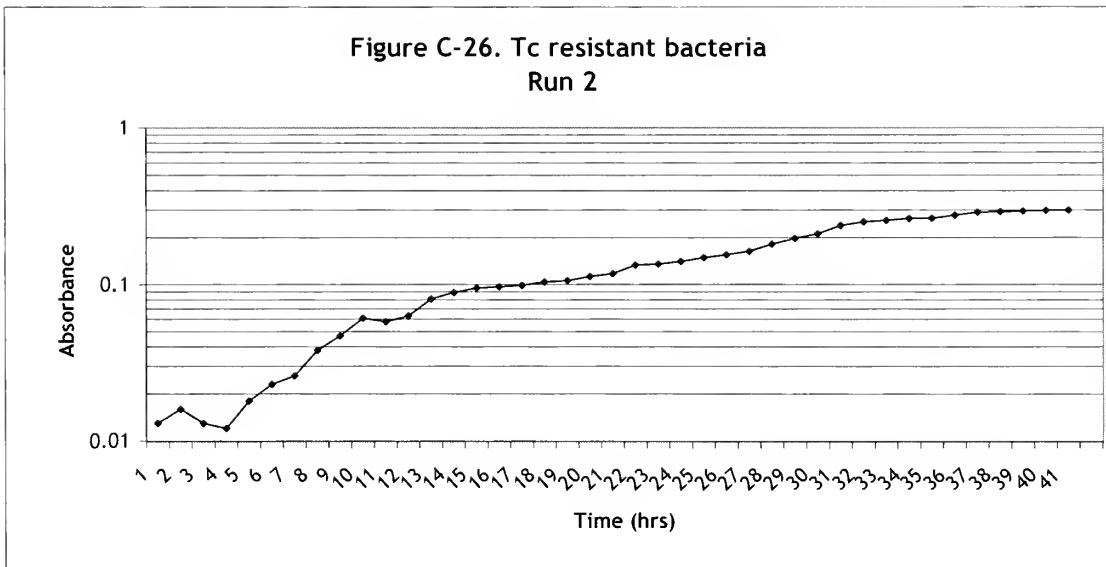
Donor (Tc resistant *E.coli*) growth rate on LB broth + Tc

Time(min)	Time(hrs)	Run1	Run2	Run3	Average
0					
30					
60	1	0.015	0.013	0.018	0.015
120	2	0.014	0.016	0.021	0.017
180	3	0.012	0.013	0.017	0.014
240	4	0.011	0.012	0.014	0.012
300	5	0.023	0.018	0.021	0.021
360	6	0.026	0.023	0.025	0.025
420	7	0.027	0.026	0.024	0.026
480	8	0.054	0.038	0.042	0.045
540	9	0.056	0.047	0.045	0.049
600	10	0.058	0.061	0.052	0.057
660	11	0.06	0.058	0.055	0.090
720	12	0.071	0.063	0.062	0.065
780	13	0.079	0.081	0.071	0.077
840	14	0.091	0.089	0.091	0.090
900	15	0.098	0.095	0.099	0.097
960	16	0.096	0.097	0.098	0.097
1020	17	0.101	0.099	0.103	0.101
1080	18	0.108	0.104	0.105	0.106
1140	19	0.107	0.106	0.103	0.105
1200	20	0.111	0.113	0.109	0.111
1260	21	0.115	0.118	0.114	0.116
1320	22	0.127	0.134	0.129	0.130
1380	23	0.137	0.136	0.133	0.135
1440	24	0.139	0.141	0.139	0.140
1500	25	0.141	0.149	0.145	0.145
1560	26	0.144	0.156	0.159	0.153
1620	27	0.149	0.164	0.167	0.160
1680	28	0.153	0.182	0.178	0.171
1740	29	0.166	0.198	0.193	0.186
1800	30	0.172	0.212	0.205	0.196
1860	31	0.185	0.239	0.211	0.212
1920	32	0.203	0.253	0.232	0.229
1980	33	0.206	0.258	0.248	0.237
2040	34	0.218	0.266	0.255	0.246
2100	35	0.228	0.267	0.265	0.253
2160	36	0.236	0.278	0.269	0.261
2220	37	0.243	0.291	0.272	0.269
2280	38	0.248	0.294	0.278	0.273
2340	39	0.259	0.296	0.283	0.279
2400	40	0.261	0.299	0.288	0.283
2460	41	0.263	0.301	0.294	0.286



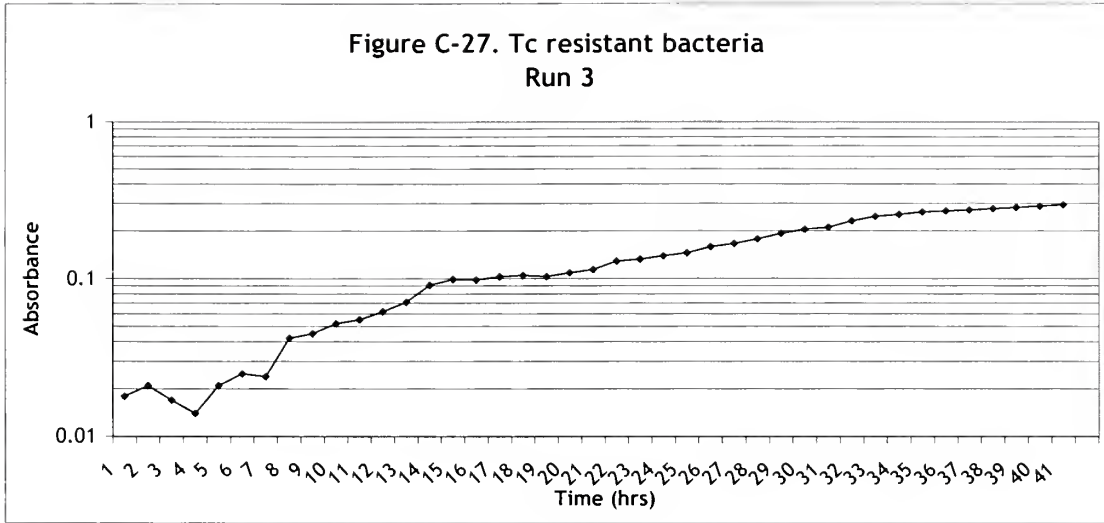
Absorbance increases from 0.101 to 0.203

Actual growth rate = (32 hrs - 17 hrs) = 15 hrs



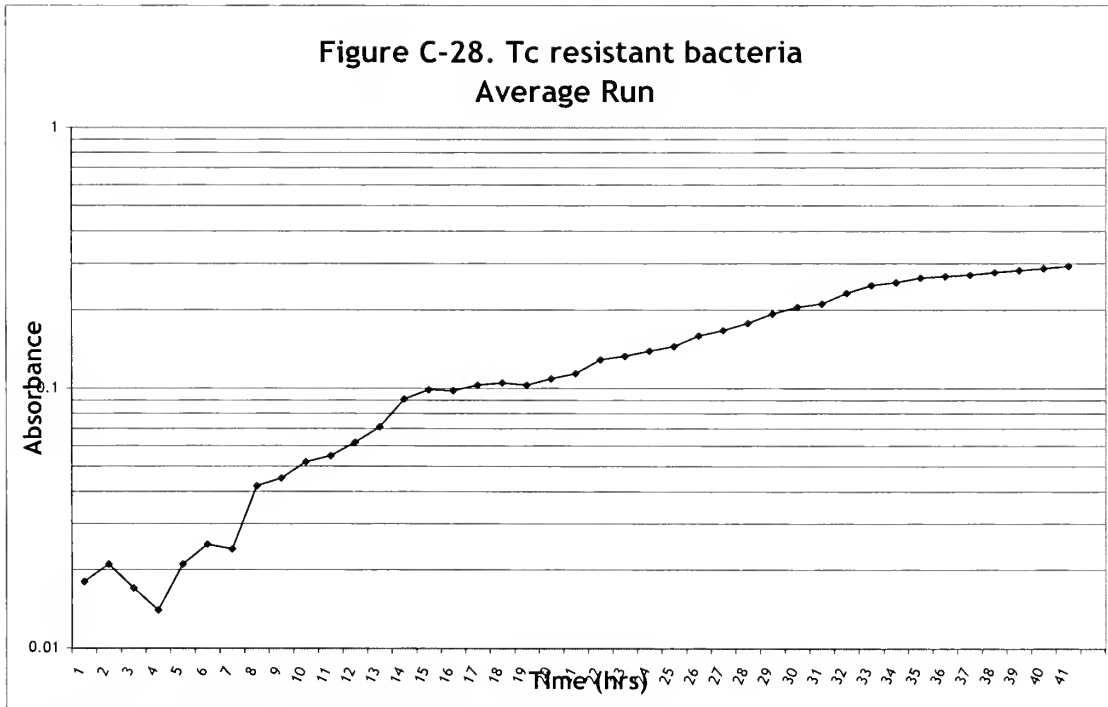
Absorbance increases from 0.106 to 0.212

Actual growth rate = (30 hrs - 19 hrs) = 11 hrs



Absorbance increases from 0.106 to 0.212

Actual growth rate = (31 hrs - 18 hrs) = 13 hrs

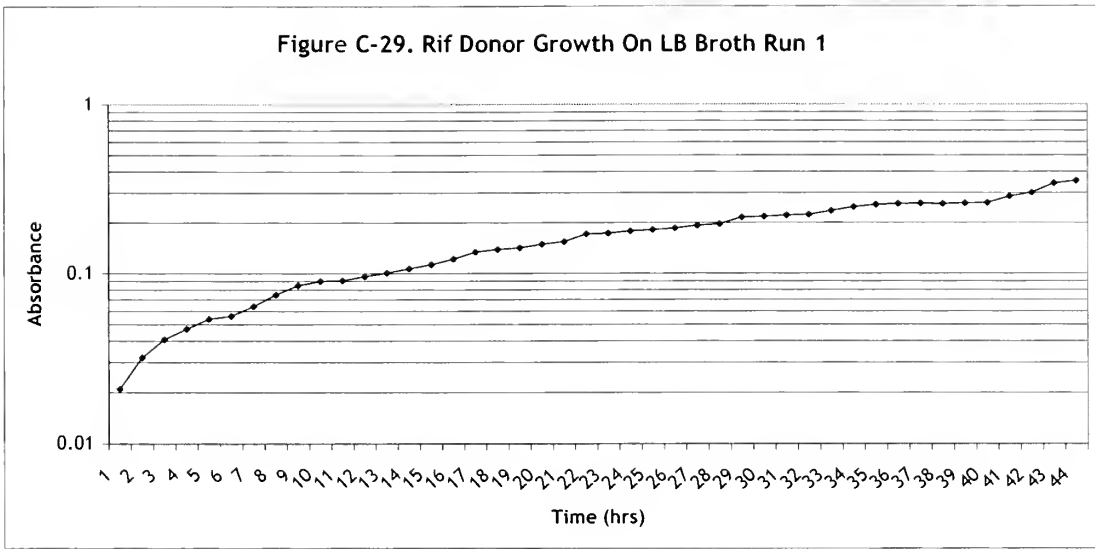


Absorbance increases from 0.106 to 0.212

Actual growth rate = (31 hrs - 18 hrs) = 13 hrs

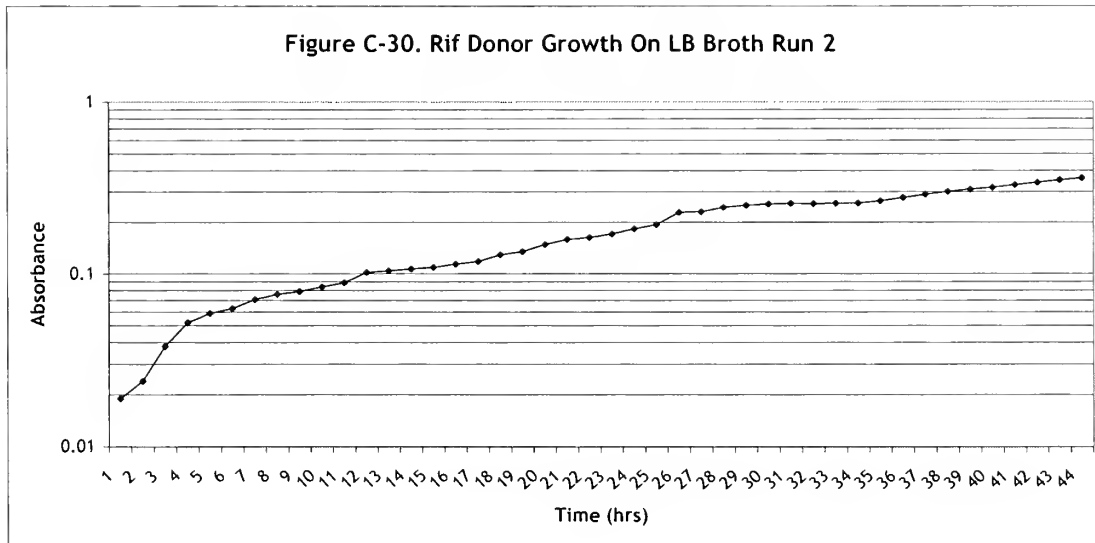
Donor (Rif resistant) growth rate on LB broth + Rif

Time(min)	Time(hrs)	Run1	Run2	Run3	Average
0					
30					
60	1	0.021	0.019	0.018	0.019
120	2	0.032	0.024	0.029	0.028
180	3	0.041	0.038	0.037	0.039
240	4	0.047	0.052	0.045	0.048
300	5	0.054	0.059	0.051	0.055
360	6	0.056	0.063	0.062	0.060
420	7	0.064	0.071	0.068	0.068
480	8	0.075	0.076	0.079	0.077
540	9	0.085	0.079	0.086	0.083
600	10	0.090	0.084	0.097	0.090
660	11	0.091	0.089	0.098	0.093
720	12	0.096	0.102	0.101	0.100
780	13	0.101	0.104	0.105	0.103
840	14	0.107	0.107	0.121	0.112
900	15	0.113	0.109	0.123	0.115
960	16	0.122	0.114	0.132	0.123
1020	17	0.134	0.118	0.139	0.130
1080	18	0.139	0.129	0.147	0.138
1140	19	0.142	0.135	0.153	0.143
1200	20	0.149	0.148	0.159	0.152
1260	21	0.154	0.159	0.163	0.159
1320	22	0.171	0.163	0.168	0.167
1380	23	0.173	0.171	0.185	0.176
1440	24	0.178	0.183	0.189	0.183
1500	25	0.181	0.194	0.193	0.189
1560	26	0.185	0.228	0.212	0.208
1620	27	0.192	0.23	0.228	0.217
1680	28	0.196	0.244	0.231	0.224
1740	29	0.215	0.251	0.236	0.234
1800	30	0.217	0.255	0.243	0.238
1860	31	0.221	0.257	0.253	0.244
1920	32	0.223	0.256	0.255	0.245
1980	33	0.235	0.257	0.259	0.250
2040	34	0.247	0.258	0.261	0.255
2100	35	0.255	0.267	0.265	0.262
2160	36	0.258	0.278	0.269	0.268
2220	37	0.26	0.291	0.272	0.274
2280	38	0.259	0.302	0.278	0.280
2340	39	0.261	0.311	0.283	0.285
2400	40	0.262	0.319	0.299	0.293
2460	41	0.286	0.33	0.312	0.309
2520	42	0.301	0.341	0.314	0.319
2580	43	0.342	0.353	0.321	0.339
2640	44	0.354	0.362	0.331	0.349



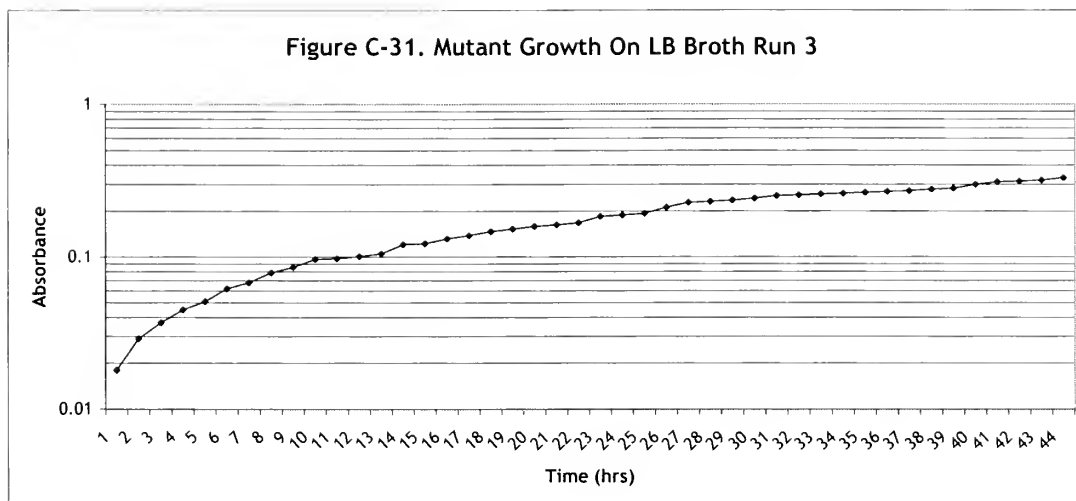
Absorbance increases from 0.107 to 0.215

Actual growth rate = (29 hrs - 14 hrs) = 15 hrs



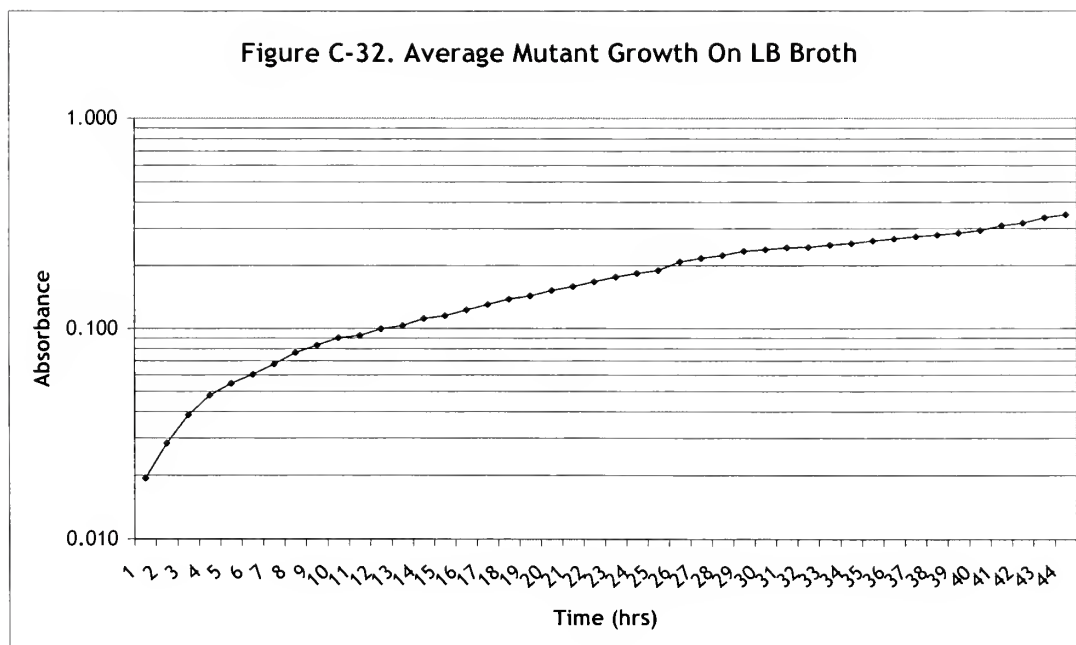
Absorbance increases from 0.114 to 0.228

Actual growth rate = (27 hrs - 17 hrs) = 10 hrs



Absorbance increases from 0.121 to 0.243

Actual growth rate = (30 hrs - 14 hrs) = 16 hrs



Absorbance increases from 0.114 to 0.228

Actual growth rate = (28 hrs - 14 hrs) = 14 hrs

WILD TYPE, DONOR, RECIPIENT, AND MUTANT GROWTH RATES
AT VERY LOW CONCENTRATIONS OF Cd.

TABLE C-1. SUMMARY OF WILD TYPE, Tc and Rif DONOR, RECIPIENT, AND
MUTANT AT VERY LOW CONCENTRATIONS OF Cd.

	R1	R2	R3	AVG
WILD TYPE	60min	60min	60min	60min
WILD TYPE + Cd1	60min	60min	60min	60min
WILD TYPE + Cd2	60min	60min	60min	60min
WILD TYPE + Cd3	90min	90min	90min	90min

	R1	R2	R3	AVG
Rif resistant bacteria (recipient)	3 hrs	1 hr	3 hrs	3 hrs
Rif resistant bacteria (recipient) + Cd3	5 hrs	5 hr	6 hrs	6 hrs

	R1	R2	R3	AVG
Tc resistant bacteria(donor)	15 hrs	11 hrs	13 hrs	13 hrs
Tc resistant bacteria(donor) + Cd3	no growth			

	R1	R2	R3	AVG
Rif resistant donor	15 hrs	10 hr	16 hrs	14 hrs
Rif resistant donor + Cd3	no growth			

	R1	R2	R3	AVG
MUTANT on LB	15hrs	11hrs	13hrs	13hrs
MUTANT on LB + Rf + Tc	7hrs	10hrs	10hrs	10hrs
MUTANT + Rf + Tc + Cd1	33hrs	36hrs	32hrs	34hrs
MUTANT + Rf + Tc + Cd2	50hrs	49hrs	48hrs	48hrs
MUTANT + Rf + Tc + Cd3	no growth			

Cd1= 50×10^{-6} M

Cd2= 100×10^{-6} M

Cd3= 300×10^{-6} M

MIC for mutant was 300×10^{-6} M

Figure C-33. Appearance of plasmids in mutant and donor strain

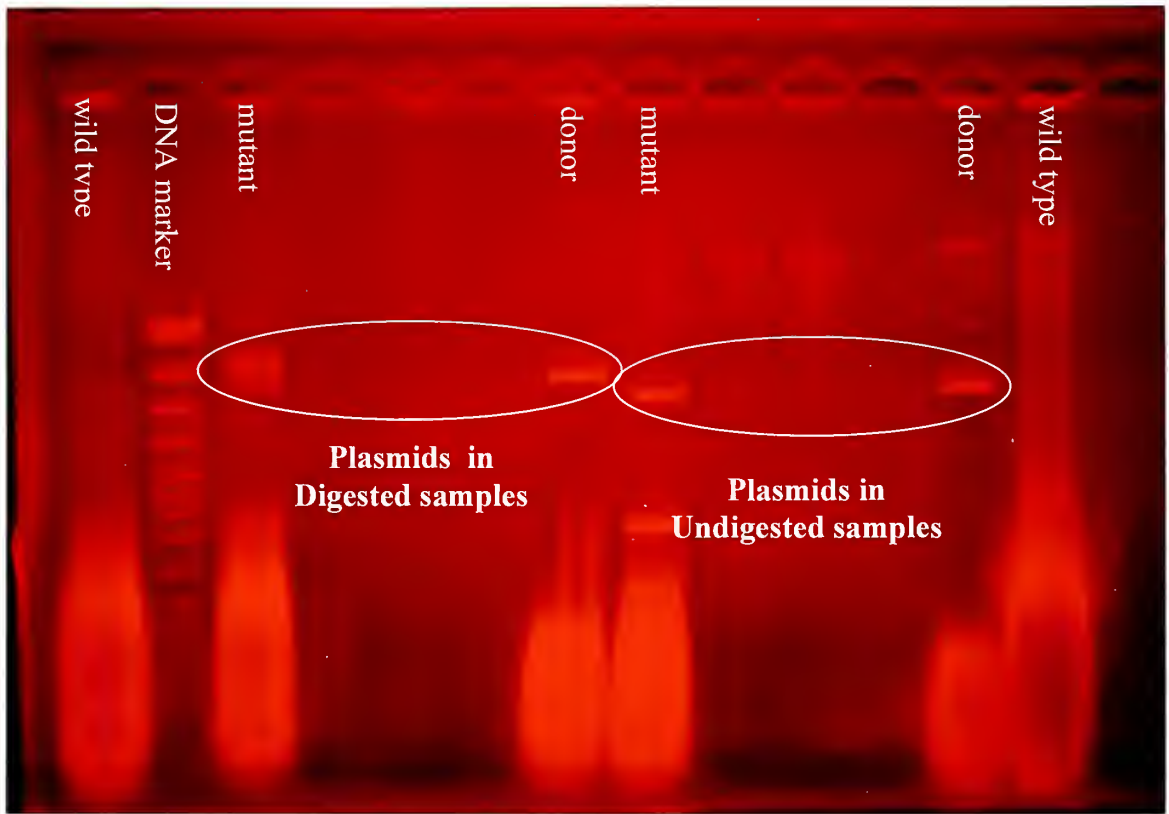


Figure C-34. Appearance of plasmids in mutant and donor strain

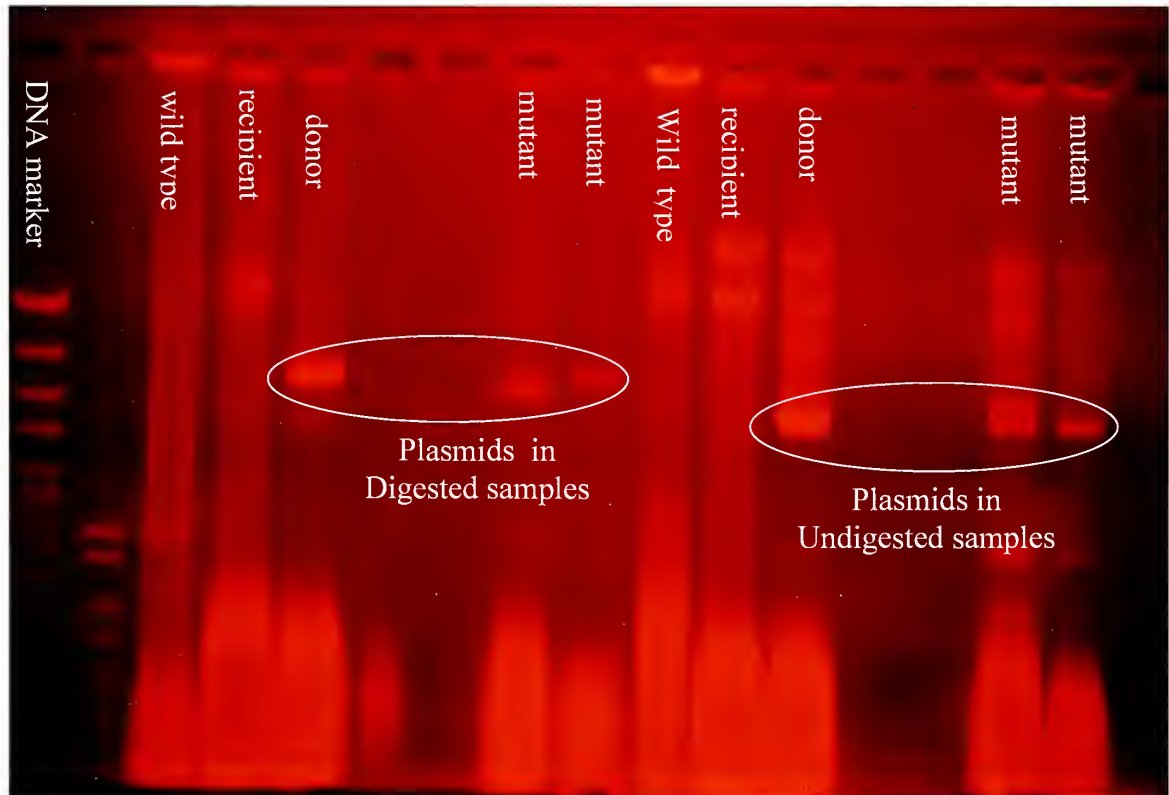


Figure C-35. Appearance of plasmids in mutant and donor strain

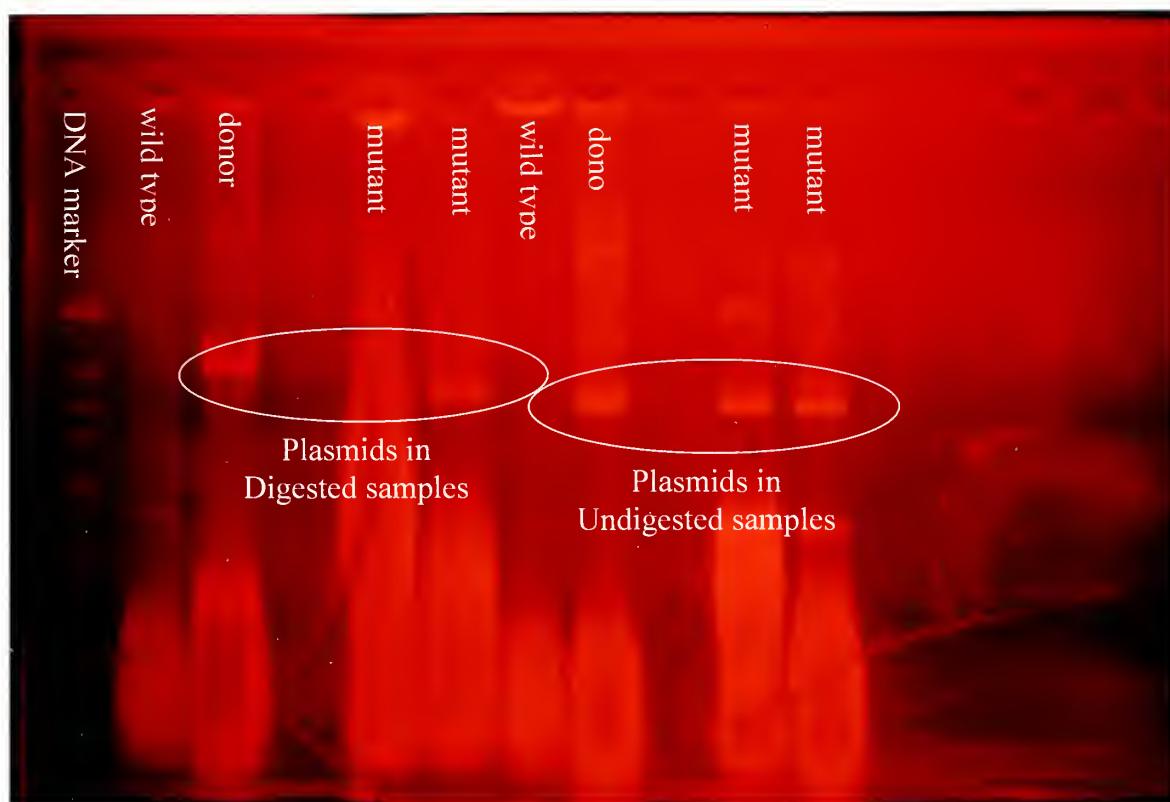


Figure C-36. Appearance of plasmids in mutant and donor strain

